

Springer Theses

Recognizing Outstanding Ph.D. Research

Claire Louisa Tinker-Mill

Nanoscale Imaging and Characterisation of Amyloid- β



Springer

Springer Theses

Recognizing Outstanding Ph.D. Research

Aims and Scope

The series “Springer Theses” brings together a selection of the very best Ph.D. theses from around the world and across the physical sciences. Nominated and endorsed by two recognized specialists, each published volume has been selected for its scientific excellence and the high impact of its contents for the pertinent field of research. For greater accessibility to non-specialists, the published versions include an extended introduction, as well as a foreword by the student’s supervisor explaining the special relevance of the work for the field. As a whole, the series will provide a valuable resource both for newcomers to the research fields described, and for other scientists seeking detailed background information on special questions. Finally, it provides an accredited documentation of the valuable contributions made by today’s younger generation of scientists.

Theses are accepted into the series by invited nomination only and must fulfill all of the following criteria

- They must be written in good English.
- The topic should fall within the confines of Chemistry, Physics, Earth Sciences, Engineering and related interdisciplinary fields such as Materials, Nanoscience, Chemical Engineering, Complex Systems and Biophysics.
- The work reported in the thesis must represent a significant scientific advance.
- If the thesis includes previously published material, permission to reproduce this must be gained from the respective copyright holder.
- They must have been examined and passed during the 12 months prior to nomination.
- Each thesis should include a foreword by the supervisor outlining the significance of its content.
- The theses should have a clearly defined structure including an introduction accessible to scientists not expert in that particular field.

More information about this series at <http://www.springer.com/series/8790>

Claire Louisa Tinker-Mill

Nanoscale Imaging and Characterisation of Amyloid- β

Doctoral Thesis accepted by
Lancaster University, UK

 Springer

Author

Dr. Claire Louisa Tinker-Mill
Department of Physics
Lancaster University
Lancaster
UK

Supervisors

Prof. Oleg Kolosov
Department of Physics
Lancaster University
Lancaster
UK

Prof. David Allsop
Department of Biological Life Sciences
Lancaster University
Lancaster
UK

ISSN 2190-5053

Springer Theses

ISBN 978-3-319-39533-3

DOI 10.1007/978-3-319-39534-0

ISSN 2190-5061 (electronic)

ISBN 978-3-319-39534-0 (eBook)

Library of Congress Control Number: 2016942535

© Springer International Publishing Switzerland 2016

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, express or implied, with respect to the material contained herein or for any errors or omissions that may have been made.

Printed on acid-free paper

This Springer imprint is published by Springer Nature

The registered company is Springer International Publishing AG Switzerland

List of Publications

Academic Publications

Amyloid- β fibrils in Alzheimer's Disease are not inert when bound to copper ions but can degrade hydrogen peroxide and generate reactive oxygen species

Mayer, J., Tinker-Mill, C., Kolosov, O., Zhang, H., Tabner, B. & Allsop, D.

Journal of Biological Chemistry. 289, p. 12052–12062 11 p. 25/04/2014

Ultrasonic force microscopy for nanomechanical characterization of early and late-stage amyloid- β peptide aggregation

Tinker-Mill, C., Mayer, J., Allsop, D. & Kolosov, O.

Scientific Reports. 4, 7 p. 4004, 2014

A novel retro-inverso peptide inhibitor reduces amyloid deposition, oxidation and inflammation and stimulates neurogenesis in the APP^{swe}/PS1 Δ E9 mouse model of Alzheimer's Disease

Parthasarathy, V., McClean, P. L., Hölscher, C., Taylor, M., Tinker, C., Jones, G., Kolosov, O., Salvati, E., Gregori, M., Masserini, M. & Allsop, D.

PLoS ONE. 8, 1, 11 p. e54769, 2013

Also in preparation

Retro-inverso peptide inhibitory nanoparticles (PINPs) as potent inhibitors of aggregation of the Alzheimer's A β peptide

Gregori, M., Taylor, M., Tinker-Mill, C., Michael, M., Kolosov, O., Salvati, E., Re, F., Minniti, S., Zambelli, V., Masserini, M. & Allsop, D.

Comparison of photothermal and opto-acoustic response of nanoscale probes for Mid-IR photothermal microspectroscopy (PTMS) of nanostructured biological materials

Tovee, P.D., Tinker-Mill, C., Kjoller, K., Allsop, D., Weightman, P., Surman, M., Siggel-King, M., Wolski, A. & Kolosov, O.V.

Scanning thermal microscopy imaging of Amyloid- β

Tovee, P.D., Tinker-Mill, C., Allsop, D., & Kolosov, O.V.

Presentations at Conferences

Nanoscale SPM Characterisation of Nacre Aragonite Plates and Synthetic Human Amyloid Fibres.

Grishin, I., Tinker, C., Allsop, D., Robson, A & O.V. Kolosov.

Nanotech 2012 Santa Clara, California.

Proceedings of Nanotech—2012, TechConnect World 2012, pages 2, 940 ISBN: 978-1-4665-6278-3, NSTI, CRC press, Santa Clara, USA (2012).

Nanoscale morphology and nanomechanical characterisation of recombinant human Amyloid- β 1-42 via tapping mode and ultrasonic force microscopies.

Tinker, C., Allsop D. & Kolosov, O.

Seeing at the Nanoscale, July 2012, Bristol University.

Nanoscale morphology and nanomechanical characterisation of recombinant human Amyloid- β 1-42 via tapping mode and ultrasonic force microscopies.

Tinker, C., Allsop D. & Kolosov, O.

European Microscopy Conference, September 2012, Manchester.

Nanoscale dynamics of Amyloid- β fibres on poly-L-Lysine substrate in air and liquid environments via atomic force and ultrasonic force microscopy

Tinker, C., Allsop, D., Kolosova, K., Dinelli, F & Kolosov, O

Multifrequency Conference, October 2012, Madrid.

Nanoscale imaging of Alzheimer's Disease: Getting to the core of it

Tinker, C., Allsop, D., Robson, A. & Kolosov, O

Lancaster University Sci-Tech Christmas Conference, 17th December 2012, Lancaster.

Ultrasonic force microscopy studies of the nanoscale structure of Amyloid- β fibres in a liquid environment

Tinker C., Allsop, D., Kolosova, K., Dinelli, F. & Kolosov O.

Bristol University Nanomaterial Futures Conference, 17th February 2013, Bristol.

Nanostructure of Amyloid Fibres using Ultrasonic Force Microscopy

Tinker, C., Allsop D. & Kolosov, O.

Waterloo University talk as a visiting student, September 2013, Waterloo, Canada.

Nanomechanical and nanothermal mapping of initial stages of amyloid fibres formation

Kolosov, O., Tovee, P., Tinker-Mill, C. & Allsop, D.

Bristol Nanoscience Symposium 15/09/2014, Bristol.

Press Articles

<http://www.labnews.co.uk/news/sewing-machine-inspires-imaging-tool-for-alzheimers/>

21/04/14: (Lab News)

Tinker-Mill, C., Mayes, J., Allsop, D. & Kolosov, O.

Imaging tool gives insight into origins of Alzheimer's

2/04/14: (Medical Express)

Tinker-Mill, C., Mayes, J., Allsop, D. & Kolosov, O.

New imaging tool provides fresh insight into origins of Alzheimer's, Parkinson's disease

2/04/14: (News medical)

Tinker-Mill, C., Mayes, J., Allsop, D. & Kolosov, O.

Sewing machine' idea gives insight into origins of Alzheimer's

1/04/14: (USA Daily news) (Science daily) (Deccan chronicle) (Business Standard)

Tinker-Mill, C., Mayes, J., Allsop, D. & Kolosov, O.

Tools of the Trade

26/02/14: (BioTechniques - The International Journal of Life Science Methods)

Tinker-Mill, C., Mayes, J., Allsop, D. & Kolosov, O.

Awards

Juno Award for Research Excellence 2014

Juno Award for Research Excellence 2012

Supervisor's Foreword

It is my great pleasure and true honour to write this foreword to Claire Tinker-Mill's thesis. I was always amazed with Claire, a mainstream biologist by training, who has been able to successfully develop highly sophisticated physical methods for exploring nanoscale properties of materials that only a handful of research groups worldwide was privy to. Her work is an excellent example how a truly interdisciplinary exploration of nature should be conducted.

The subject of Claire's thesis came from my ongoing collaboration with Prof. David Allsop at Lancaster University Health and Medicine, who was working on uncovering molecular mechanisms of Alzheimer's disease and developing paradigms for the novel drugs that can stop or, better, reverse this debilitating disease. While there were multiple optical and chemical methods for detecting an aggregation of misfolded amyloid peptides, the known cause for the disease, the exact mechanisms of their aggregation and the source of pathological changes was not at all clear.

While researchers' ultimate goal would be to directly observe such folding, this was not an easy task, given that most relevant biomolecular interactions are happening in the liquid, and the interacting structures are of the order of few nanometres in size. When Claire joined my group, we were perfecting a rather powerful method for nanoscale characterisation and imaging of inorganic materials—e.g. semiconductor quantum dots or metallic interfaces—ultrasonic force microscopy or UFM. This approach combined a near-atomic resolution of scanning probe microscopy with the sensitivity of ultrasonic imaging to local mechanical properties of surface and near-surface areas.

Claire has successfully applied UFM for the detection of the early-stage toxic amyloid aggregates by combining UFM with methods for capturing all range of peptide assemblies from the monomers to the fully formed amyloid plaques. She explored a small universe of diverse substrates, coatings and buffers, to home in on the winning combination that allowed to capture practically all peptide components on the perfectly flat surface. The nanomechanical contrast provided by UFM allowed her to pinpoint toxic amyloid peptides and to reliably differentiate them

from the substrate. Claire also established that the nanoscale ultrasonic probe had another huge advantage—the minute but very-high-frequency ultrasonic vibrations used in UFM allowed a nanoscale tip of the probe to gently poke into the biological molecules, leaving them intact, while still identifying a wrongly folded ones. The approach successfully uncovered that a significant portion of the initial amyloid peptide even at later stages of assembly was in the small molecular weight stage that is regarded as triggers of cellular death in Alzheimer's disease. This work allowed to develop the novel retro-inverso peptide inhibitor based on nanoscale liposomes as carriers for the anti-Alzheimer's drugs. These have shown potential for 1000 times higher efficiency in scrubbing harmful fibres from biological fluids and reducing the peptide deposition in this disease, hence paving the way to the new paradigm for the future drugs.

While expanding this avenue of research would be more than sufficient for a solid Ph.D. thesis, Claire did not pause at all and explored other physical methods to study amyloid proteins. She used a relative of UFM—heterodyne force microscopy, or HFM, to measured time response of peptides. Claire found that pathological misfolded peptides have a characteristic time signature, linked to their mechanical relaxation that was different from the normal proteins, adding new method that may help to detect early stages of Alzheimer's disease in the future.

Finally during her Ph.D. work, Claire explored yet another powerful method allowing to directly identify biomolecules with nanoscale resolution—via combining chemical identification by infrared optical excitation with the scanning probe microscopy. As high-power tunable infrared sources are not easy to come by, she was studying amyloid aggregates on the Daresbury ALICE accelerator that powered a free electron laser, where she became a dedicated commissioner responsible for safety of this complex physical instrument. This work, instigated by Claire, can create an ultimate instrument for early detection of Alzheimer's disease.

Overall, Claire's thesis not only provides a deep insight into a novel scanning probe methodology for nanoscale exploration of biological molecules and their aggregates, but also shows an inspiring example of how to solve a complex interdisciplinary problem at the interface of biology, physics and chemistry. It will be a useful reference both for established researchers and graduate students working in these highly active areas of modern research.

Lancaster
May 2016

Prof. Oleg Kolosov

Abstract

In this work, several novel scanning probe microscopy (SPM) methods have been applied to the study of the amyloid peptide implicated in the pathogenesis of Alzheimer's disease (AD). Amyloid- β ($A\beta$) undergoes a hierarchy of aggregation following a structural transition, making it an ideal subject of studying with SPM.

The application of SPM-based techniques to biological samples has become increasingly common place. However, these techniques are not always immediately suitable for imaging delicate samples of proteins and adaptations must be made before imaging can be considered successful. AD is the most common form of dementia worldwide, and a growing concern for health authorities. As a result, it has attracted the attention of a wide range of disciplines. There has been much work conducted which combines the main pathogenic peptide, $A\beta$, with atomic force microscopy (AFM) in order to elucidate more about its aggregation behaviour; however, these techniques offer little more than structural comments, with only the most advanced forms of cryo-electron microscopy (EM) providing more details on the nanoscale. Presented here is a method for reliably and robustly producing samples of $A\beta$ by capturing them at various stages of aggregation, as well as the results of subsequent imaging by various methods of AFM. Each of the AFM techniques studied provides additional "added value" to the data which can typically be collected by AFM, either nanomechanical, elastic, thermal or spectroscopical.

By imaging samples of $A\beta$ with ultrasonic force microscopy, a detailed sub-structure to the morphology could be seen, which correlates well with the most advanced cryo-EM work. In addition, this technique was ideal for detecting the most toxic form of $A\beta$, early aggregates, in a sensitive and nondestructive fashion robustly differentiating them from the underlying layer of another peptide (poly-L-Lysine) that was designed to reliably capture the $A\beta$ aggregates. Early work investigating the potential for combining an established method of thermal AFM with a mid-IR laser system also shows promise for detecting the response of the protein.

It was also the focus of this work to study the aggregation of $A\beta$ using dynamic light scattering (DLS), in order to confirm whether the technique could identify

differences between populations throughout the aggregation process. This was applied in conjunction with potential therapeutics which targets the early aggregates to prevent their accumulation, as well as block formation of fibrils.

Ultimately, this work aims to show with care to the initial protocols used, physical techniques such as AFM and DLS can be added to the existing methods of monitoring aggregation. Synergistic use of these techniques can generate a clearer overall picture of the effect of metal ions/developing therapeutics on A β aggregation and provide more detail than classical biological techniques alone.

Acknowledgments and Dedications

Acknowledgments

I would first like to thank Prof. David Allsop and Dr. Oleg Kolosov for their patience and guidance throughout the completion of this work. I would also like to extend my eternal gratitude to my colleagues in both the Department of Physics and Biomedical Life Sciences for their help and advice.

I am blessed to have made so many friends throughout this Ph.D., both within the university and elsewhere: Jenny Mayes and Alex Robson, thank you for brightening my morning with many a cup of coffee and chatter, in addition to all your support with my research; Riccardo Mazzocco, who will always be “my favourite Italian”; Kylie O’Shea and Ben Shreeve, without whom I would not have enjoyed nearly as much cake as I have currently; the wonderful Taylor Lura, Emily Smith and Christie Herd, who have made me laugh so hard I have cried; My “Bookend” Louise Walker, for fostering my madness and creating the candyfloss world with me. I would like to thank Dr. Christine Shirras for her guidance and support as my teaching mentor. Lastly, my friends in Canada: Ronnie and Liz Drolle, who I would never have had the joy of meeting if it were not for this Ph.D. All of these people, and more, have made this experience the happiest of my life. You have brought so much joy, sparkle and laughter as we have journeyed together. I’m so lucky to have you in my life.

My family’s support and belief in me has been paramount throughout my studies, and I thank them from the bottom of my heart of believing in me, comforting me and giving me the strength I have.

Most importantly, I want to thank my long-suffering husband, Richard. Without you I would not be me, and this would not have happened. We share this achievement as we do everything. I love you now, forever and always and cannot thank you enough for your support and love.

Dedications

The work in this thesis is dedicated to those that sadly did not get the opportunity to see me finish this journey, but offered their unconditional support and love throughout my life: Vera Pennington, who sadly lost her own battle with Alzheimer's disease inspired with her tireless work as a daughter, mother and grandmother; Rex Pennington, who went long before this journey began, thank you for the memories; Marjorie and Ernest Tinker, without whom I would not have been able to begin this journey, I love and miss you both so much and am eternally grateful for all that you did for me, and It breaks my heart you will not see me graduate.

Contents

1	Introduction	1
	References	4
2	Theoretical Concepts of Scanning Probe Microscopy and Dynamic Light Scattering and Their Relation to the Study of Peptide Nanostructures	7
2.1	Introduction	7
2.2	Scanning Probe Microscopy	8
	2.2.1 Tip-Surface Interactions	8
	2.2.2 AFM Detection Modes.	10
2.3	Dynamic Light Scattering (DLS)	21
2.4	Direct Imaging via AFM and Electron Microscopy Studies of A β 1:42 and Their Findings.	23
2.5	Conclusion	25
	References	26
3	Alzheimer's Disease and the Aggregation of Amyloid β	31
3.1	Introduction	31
3.2	Symptoms and Diagnosis	32
3.3	Pathology and Physiology.	33
3.4	Epidemiology of AD	34
3.5	Genetic Risk Factors Associated with AD.	35
3.6	The Amyloid Precursor Protein	36
3.7	The Amyloid Cascade Hypothesis	39
3.8	Oligomers of A β Are Likely to Be the Cause of AD Pathology	41
3.9	Counter Arguments of the Amyloid Cascade Hypothesis	43
3.10	Therapeutic Design for the Treatment of AD.	44
3.11	Amyloidosis as the Causative Factor in a Wide Range of Diseases	45
3.12	Conclusions	46
	References	47

4	Experimental Methodology	53
4.1	Introduction	53
4.2	Materials and General Reagents	53
4.3	Nanomechanical Methods of SPM	54
	4.3.1 Nanomechanical Mapping of Peptides and Proteins via Scanning Probe Microscopy	54
4.4	Spectroscopic Methods of SPM	60
4.5	SPM Image Processing and Tip Convolution	65
4.6	Identification of Amyloid β by Classical Biomedical Techniques	65
	4.6.1 Aggregation Conditions of Amyloid Peptides	65
	4.6.2 Monitoring of Aggregation State Using Thioflavin T Assay	66
4.7	Dynamic Light Scattering	67
	4.7.1 Calibration of DLS System Using Gold Nanoparticles	69
	4.7.2 Experimental Setup for Temperature Dependent Measurements of LCST Compounds	69
	4.7.3 Characterization of Peptide Inhibitor NanoParticle Liposomes Using DLS	69
	4.7.4 Detection of A β 1:40 and 1:42 Using DLS	69
4.8	Substrate Modification for the Attachment of Amyloid Proteins	70
	4.8.1 Confirmation of A β Attachment to the Substrate	70
4.9	Conclusion	71
	References	72
5	Substrate Development of the Imaging of Amyloid Proteins with SPM Methods	73
5.1	Introduction	73
5.2	Muscovite Mica as a Standard SPM Substrate	74
5.3	Incubation of A β 1:42 in Volatile Buffers	75
5.4	Chemical Modification of the Mica Substrate	76
	5.4.1 Incubation of Cleaved Mica with Divalent Ions	77
	5.4.2 Poly Prep Slides	77
	5.4.3 PLL-Mica	78
5.5	Confirmation that PLL Does not Interfere with Fibre Morphology	80
5.6	Confirmation of Attachment of A β 1:42 to PLL Coated Mica	81
5.7	Conclusion	84
	References	85
6	Scanning Probe Microscopy Methods of Imaging Amyloid Peptides During the Aggregation Process	87
6.1	Introduction	87
6.2	Tapping Mode Imaging of A β 1:42—Detection of Metal Ions Induced Alterations in Morphology	88

6.3	UFM of A β 1:42	91
6.4	Determination of Fine Structural Details of A β 1–42 with UFM.	93
6.5	Reducing Friction Forces and Sample Damage Artefacts via UFM	95
6.6	Application of UFM Underliquid.	97
6.7	Tip Only Ultrasonic Excitation—Waveguide-UFM; Further Enhancement of the Technique.	99
6.8	Conclusion	101
	References	102
7	Spectroscopy and Thermal SPM Methods of Studying Aβ1:42	107
7.1	Introduction	107
7.2	Scanning Thermal Microscopy Nanoscale Mapping of Thermal Conductivity of A β 1:42.	108
7.3	SThM-IR Imaging at Fixed Wavelength of A β 1:42	110
7.4	Measurements of A β on Anasys “Nano-IR” System.	114
7.5	Conclusion	115
	References	117
8	The Application of Biophysical Techniques to the Study of the Inhibition of Aggregation of Aβ Using PINPs Liposomes	121
8.1	Introduction	121
8.2	Development of the RI-OR2-TAT PINP Inhibitor	122
8.3	Use of TM-AFM to Confirm the Inhibition of A β 1:42 Using RI-TAT.	123
8.4	Test Study of Non-biological Samples of Well-Defined Behaviours Using DLS.	124
8.5	Characterisation of the Morphology and Sizes of PINPs Liposomes	127
8.6	AFM of A β 1:42 Exposed to RI-OR2-TAT PINPs	129
8.7	Monitoring Aggregation of A β 1:42 Using DLS	129
8.8	Inhibition of A β 1:42 Aggregation Using RI-OR2-TAT PINPs as Detected by DLS.	132
8.9	Conclusion	135
	References	135
9	Conclusion and Future Perspectives	139
9.1	Conclusions	139
9.2	Future Perspectives	144
	References	147