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in Children VI

Volume 659

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Adam Finn · Nigel Curtis · Andrew J. Pollard
Editors

Hot Topics in Infection and Immunity in Children VI

 Springer

Editors

Adam Finn
Institute of Child Life and Health
UBHT Education Centre
University of Bristol
Upper Maudlin Street
Bristol
UK BS2 8AE
adam.finn@bristol.ac.uk
a.finn@sheffield.ac.uk

Nigel Curtis
Royal Children's Hospital
Department of Paediatrics
University of Melbourne
Parkville VIC 3052
Australia
nigel.curtis@rch.org.au

Andrew J. Pollard
University of Oxford
Level 4, John Radcliffe
Hospital
Oxford
UK OX3 9DU
andrew.pollard@paediatrics.ox.ac.uk

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Preface

Each of the chapters in this book is based on a lecture given at the sixth “Infection and Immunity in Children” course, held at the end of June 2008 at Keble College, Oxford.

Thus, it is the sixth book in a series that provides succinct and readable updates on just about every aspect of the discipline of Paediatric Infectious Diseases.

The seventh course (29th June–1st July 2009) has a new and topical programme delivered by top-class speakers, and a seventh edition of this book will duly follow.

As we send this edition off for publication, the news of fatal cases of H1N1 swine flu in Mexico with spread into other countries around the globe is coming through, amidst fears of a global pandemic. By the time the book is being read, the size and severity of the problem will have become much clearer. For now, the outbreak serves as yet another reminder of the enduring importance of infectious diseases in human health and global prosperity and the need to continue studying them with all the epidemiological, clinical, and laboratory tools at our disposal.

We hope this book will provide a further useful contribution to the materials available to trainees and practitioners in this important and rapidly developing field.

UK, Australia

Adam Finn,
Nigel Curtis,
Andrew J. Pollard

Acknowledgments

We thank all the contributors who have written chapters for this book, which are all based on lectures delivered at the 2008 Infection and Immunity in Children course. We are very grateful to the staff of Keble College, Oxford, UK where the course was held.

Sue Sheaf administers and runs the course. As is the rule for people who organize extremely well, she creates the illusion that it is easy and straightforward, but we know it really is not, and she deserves enormous thanks from the organizers as well as all the speakers and delegates who have attended the Oxford IIC over recent years. Once again, Lorraine Cantle has administered the production of this book, obtaining the chapters from the authors – some of whom need more persuasion and reminding than others – and working with the editors to generate the finalized product for publication. We thank her for this patient work and gratefully share with her the credit for the finished object.

We thank the European Society for Paediatric Infectious Diseases for continuing practical and financial support for this series of courses. In particular, we draw attention to the ESPID bursaries towards the costs of many young delegates' attendance. We also acknowledge the recognition given to the course by the Royal College of Paediatrics and Child Health.

Finally, we are grateful to the several pharmaceutical industry sponsors who continue generously to offer unrestricted educational grants towards the budget for the meeting.

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Contributors

Aditya H. Gaur Department of Infectious Diseases, St. Jude Children's Research Hospital, Memphis, Tennessee, USA

Alessandro R. Zanetti Department of Public Health-Microbiology-Virology, Faculty of Medicine, University of Milan, Milan, Italy

Alison J. Cody Department of Zoology, University of Oxford, South Parks Road, Oxford OX1 2PS, UK

Andrew Cant Paediatric Immunology & Infectious Diseases, Newcastle General Hospital, Westgate Road, Newcastle upon Tyne NE4 6BE, UK

Anne A. Gershon Department of Pediatrics, Columbia University College of Physicians and Surgeons, New York, NY 10032, USA

Aubrey Cunnington Immunology Unit, London School of Hygiene and Tropical Medicine, London, UK

B. Keith English The University of Tennessee Health Science Center, Memphis, Tennessee, USA

Cassandra Moran Department of Pediatrics, Duke University Medical Center, Durham, NC 27710, USA

Daniel Shouval Department of Medicine, Liver Unit, Hadassah-Hebrew University Hospital Jerusalem, Israel

Danny Benjamin Jr, MD Ph.D MPH Division of Quantitative Sciences, Duke Clinical Research Institute, Duke University Pediatrics, Durham, NC 27715, USA

David A. Kaufman Division of Neonatology, Department of Pediatrics, University of Virginia Health System, Charlottesville, VA 22908, USA

Deirdre Kelly The Liver Unit, Birmingham Children's Hospital, Birmingham B4 6NH, UK

Elizabeth Molyneux Paediatric Department, College of Medicine, P/Bag 360, Blantyre, Malawi

Frances M. Colles Department of Zoology, University of Oxford, South Parks Road, Oxford OX1 2PS, UK

H.K. Brand Department of Pediatrics, Radboud University Nijmegen Medical Center, Nijmegen, The Netherlands

Hermione Lyall Imperial College Healthcare NHS Trust, St. Mary's Hospital, London W2 1NY, UK

Koen Van Herck Centre for the Evaluation of Vaccination, Vaccine & Infectious Disease Institute, Faculty of Medicine, University of Antwerp, Antwerp, Belgium; 4Research Foundation – Flanders (FWO), Brussels, Belgium

Kondwani Kawaza Paediatric Department, College of Medicine, P/Bag 360, Blantyre, Malawi

Marc Tebruegge Department of Paediatrics, The University of Melbourne; Infectious Diseases Unit, Department of General Medicine; Microbiology & Infectious Diseases Research Group, Murdoch Children's Research Institute: Royal Children's Hospital Melbourne, Parkville, VIC 3052, Australia

Martin C.J. Maiden Department of Zoology, University of Oxford, South Parks Road, Oxford OX1 2PS, UK

Nigel Curtis Department of Paediatrics, The University of Melbourne; Infectious Diseases Unit, Department of General Medicine; Microbiology & Infectious Diseases Research Group, Murdoch Children's Research Institute: Royal Children's Hospital Melbourne, Parkville, VIC 3052, Australia

P.W.M. Hermans Department of Pediatrics, Radboud University Nijmegen Medical Center, Nijmegen, The Netherlands

Pierre Van Damme Faculty of Medicine, Centre for the Evaluation of Vaccination, Vaccine & Infectious Disease Institute, University of Antwerp, Antwerp, Belgium

R. de Groot Department of Pediatrics, Radboud University Nijmegen Medical Center, Nijmegen, The Netherlands

Samuel K. Sheppard Department of Zoology, University of Oxford, South Parks Road, Oxford OX1 2PS, UK

Sanjay Patel Department of Paediatric Infectious Diseases, St Mary's Hospital, London W2 1NY, UK

Simon J. Glover College of Medicine, University of Malawi, Blantyre, Malawi; Malawi-Liverpool-Wellcome Trust Clinical Research Programme, Blantyre, Malawi

Stephen J. Rogerson Department of Medicine (RMH/WH), Post Office Royal Melbourne Hospital, Parkville, VIC 3050, Australia

Theresa Cole Paediatric Immunology & Infectious Diseases, Newcastle General Hospital, Westgate Road, Newcastle upon Tyne NE4 6BE, UK

Yamikani Chimalizeni Paediatric Department, College of Medicine, P/Bag 360, Blantyre, Malawi

Infections in the Immunocompromised

Andrew Cant and Theresa Cole

1 Introduction

Infections in the immunocompromised differ significantly from those in the immunocompetent. They can be more serious, more often life threatening, more difficult to diagnose and are caused by more unusual organisms. Children can be immunocompromised for a variety of reasons and the numbers, worldwide, are growing.

2 Types of Immunodeficiency and Infection

Globally, the most common causes for immunocompromise are acquired. Severe malnutrition and the spread of HIV continue to be major causes of immunocompromise in the developing world. The development of more intensive chemotherapy and immunosuppressive drugs for malignancy and inflammatory diseases continues and the number of patients undergoing solid organ transplant (SOT) or haemopoietic stem cell transplant (HSCT) is also rising. Primary immunodeficiencies (PIDs) are also being increasingly recognized. Defects in humoral immunity make up the largest group of PIDs, at around 50%, with 20–30% being combined humoral and cellular deficiencies.

Different types of immunodeficiency or immunosuppression affect different aspects of the immune system and so predispose to different types of infection: bacterial, viral, fungal or protozoal. Understanding which part of the immune system is compromised helps identify the most likely infecting agent.

T lymphocyte defects predispose to infections such as life-threatening cytomegalovirus (CMV) pneumonitis or disseminated Epstein Barr virus (EBV) infection. These defects also present a significant risk of serious infection with

A. Cant (✉)

Paediatric Immunology & Infectious Diseases, Newcastle General Hospital, Westgate Road, Newcastle upon Tyne NE4 6BE, UK
e-mail: andrew.cant@nuth.nhs.uk

cryptosporidium and *Aspergillus* species. B lymphocyte defects can result in infections by bacteria such as *Streptococcus pneumoniae*, *Haemophilus influenzae* and *Staphylococcus aureus*, as well as echo virus and protozoa such as *Giardia*, whereas phagocytic defects will result in infections by *Staphylococcus*, *Pseudomonas* and *Aspergillus* species.

When considering infections in those undergoing HSCT, it is also important to note that there is a recognized sequence of risk for different infections at different times after HSCT, which equates with different aspects of the immune system compromise at these times (Fig. 1). Within the first 30 days, when the patient is neutropenic, there is a significant risk of infection by both Gram-negative and Gram-positive bacteria, along with herpes-simplex virus. Between 30 and 90 days after transplant, when T-cell immunity is still limited, there is a rise in the numbers of fungal and CMV infections. Later infections are more commonly caused by varicella zoster virus (VZV) or *S. pneumoniae*.

Week:	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	...	52	104	
HSV	■	■	■																							
Gram +ve	■	■	■	■																						
Gram -ve	■	■	■	■																						
Candida	■	■	■	■																						
Aspergillus												■	■	■	■	■										
Resp. virus	■	■	■	■	■																					
CMV								■	■	■	■	■	■	■	■											
VZV													■	■	■	■	■	■	■	■	■	■	■	■	■	■
PCP				■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■	■

HSV: herpes simplex virus, Gram +ve: Gram positive bacteria, Gram -ve: Gram negative bacteria, CMV: cytomegalovirus, VZV: varicella zoster virus, PCP: *Pneumocystis jiroveci* pneumonia

Fig. 1 Diagrammatic representation of timing of infections after HSCT

Whatever the cause of the immunocompromise, possible infection requires a different approach to investigation and management from those in an immunocompetent child.

3 Site of Infection

Although an understanding of the type of immunocompromise is helpful to predict the likely organism, infection in the immunocompromised also needs to be considered by the system affected.

Infections can occur in any system of the body, but the respiratory system and gastrointestinal tract are especially vulnerable, as they have large surface areas and their barrier defences are, of necessity, compromised by the need to transport

oxygen and nutrients, respectively. Disseminated viral and fungal infections are another important risk, whilst central venous catheter (CVC) infections also constitute a frequent problem in the immunocompromised. Each of these will be discussed in turn.

4 Respiratory Tract

The respiratory tract can be exposed to a wide variety of different organisms. *Pneumocystis jiroveci* pneumonia (PCP), CMV and *Aspergillus* are particularly important and well recognized sources of infection in the immunocompromised host; however, other significant pathogens have more recently been identified. These include respiratory syncytial virus (RSV); influenza; parainfluenza; adenovirus; picornaviruses; measles; human metapneumovirus; cocavirus; Coronaviruses NL63, and HKU1 and polyomaviruses WU and K1.

Pneumonitis and bronchiolitis are the most common presentations of respiratory infection, but lobar pneumonia may also occur. A defective immune/inflammatory response means that patients may have few respiratory symptoms, so there should be a low threshold for investigation. In one study where broncho-alveolar lavage (BAL) was performed in 69 children with immunodeficiency pre-HSCT, pathogens were isolated in 26 of these, six of whom were asymptomatic. PCP and bacteria were the most commonly identified organisms, followed by parainfluenza virus, CMV, RSV, influenza B and human herpes virus-6 (HHV6) (Slatter et al., 2007).

Accurate diagnosis depends on collecting the right samples and using appropriate diagnostic techniques. These include throat swabs, nasopharyngeal aspirates (NPA), BAL fluid and even lung biopsy, as deemed appropriate. Samples must be sent to look specifically for bacteria, fungi and viruses. Some respiratory pathogens will not be isolated from the upper respiratory tract; for example, PCP will not be identified on NPA, whilst other organisms found on NPA may not be found in the lower respiratory tract. This highlights the importance of BAL as a diagnostic procedure. Lung biopsy may be particularly important in the diagnosis of fungal infection, especially when there is a negative BAL in patients with persistent signs, symptoms or chest x-ray changes.

Diagnosis may require culture of organisms (bacteria, mycobacteria or viruses), immunofluorescence (viruses), polymerase chain reaction (bacteria, viruses and fungi) or antigen testing (e.g. galactomannan for *Aspergillus*). Serological testing is often ineffective, as immunodeficient children may not mount an antibody response or may be receiving intravenous immunoglobulin (IVIG), which will make results impossible to interpret. It is important to know what tests are available in your local laboratory. Discussion with the local microbiologist or virologist is essential to ensure the right samples are sent for appropriate investigations, so as not to miss a serious infection.

High resolution computerized tomography (HRCT) of the chest is more sensitive than chest x-ray, aiming to classify a disease as interstitial, airway or involving airspace, which may aid diagnosis.

4.1 *Pneumocystis jiroveci* Pneumonia (PCP)

PCP has historically been associated with HIV but is also a significant cause of morbidity in other groups of immunocompromised patients, particularly those with haematological malignancies, brain tumours requiring prolonged courses of steroids, prolonged neutropaenia or lymphopaenia, and those undergoing HSCT. Therefore PCP prophylaxis is important, as recommended by a recent Cochrane review (Green et al., 2007). This treatment is generally in the form of cotrimoxazole given three times per week. In children that cannot tolerate cotrimoxazole, either dapsone or aerosolized pentamidine can be used.

P. jiroveci infection commonly presents with tachypnoea, non-productive cough and fever, but the severity can vary. There is usually a sub-acute diffuse pneumonitis and chest x-ray changes can be subtle. These often take the form of bilateral diffuse interstitial changes, although lobar, miliary or nodular changes can be seen. HRCT may show ground glass attenuation, consolidation, nodules, thickening of interlobular septa and thin walled cysts. Mortality ranges between 5 and 40%, if treated, but can reach nearly 100% if left untreated.

Identification of PCP can be difficult. Definitive diagnosis depends on identifying the organism in respiratory tract secretions or lung tissue, usually from tracheal secretions, bronchial secretions or from lung biopsy. More recently, PCR technology has been developed for identifying PCP from secretions. In a review of children diagnosed with severe combined immune deficiency (SCID) treated at a supra-regional center, 10 out of 50 were identified as having PCP. One was diagnosed on BAL prior to transfer to the supra-regional center, one was diagnosed on nasopharyngeal secretions and BAL, seven were diagnosed on BAL alone, and in one diagnosis was not made until lung biopsy was performed (Berrington et al., 2000).

Recommended first line PCP treatment is high dose cotrimoxazole. This can, however cause a number of adverse effects, for example, neutropenia, anaemia, renal dysfunction, rash, vomiting and diarrhea. Those that cannot tolerate cotrimoxazole or those that have not improved after 5–7 days of treatment should be changed to a different agent. Choices include pentamidine, atovaquone, clindamycin/primaquine or dapsone, but experience with these agents in children is limited.

Corticosteroids should be given as an adjunctive therapy in moderate and severe PCP. A number of studies have shown a reduction in acute respiratory failure, decreased need for ventilation and decreased mortality (Sleasman et al., 1993; Bye et al., 1994; McLaughlin et al., 1995). A recent Cochrane review supports the use of corticosteroids in HIV-infected patients with PCP, especially in those with substantial hypoxaemia (Briel et al., 2006).

4.2 Viruses

A wide variety of respiratory viruses will also cause significant morbidity and mortality in the immunocompromised. Measles is an important respiratory pathogen in the immunocompromised host and it must be remembered that the typical rash may not develop. Mortality can be high, especially amongst patients with leukaemia and those undergoing HSCT. A prospective multi-center review of patients undergoing HSCT found direct RSV-associated mortality to be 17.4%, and mortality directly attributable to influenza A to be 15.3% (Ljungman et al., 2001). Respiratory viruses often present with non-specific symptoms but progress to a significant lower respiratory tract infection. Chest x-ray will often show a pneumonitis picture with diffuse interstitial changes. HRCT may show peri-bronchial thickening and ground glass attenuation without consolidation in a lobular distribution. Diagnosis requires identification of the organism from respiratory secretions. This may be possible on nasopharyngeal secretions or throat swab but may require more invasive testing, such as bronchoscopy and BAL. Laboratory techniques include immunofluorescence, PCR and viral culture. Treatment is mainly supportive, but specific treatment options are evolving, making rapid and accurate diagnosis increasingly important. Appropriate isolation and infection-control measures are essential to prevent transmission between immunocompromised patients, as these viruses can be easily spread. One UK study in a HSCT unit identified 10 cases of RSV over one winter season, and eight of the nine RSV strains that could be tested by molecular methods were found to be identical (Taylor et al., 2001).

Specific treatments for RSV infection include ribavirin and RSV monoclonal antibody (palivizumab). Ribavirin can be given orally, intravenously or via inhalation; however, the aerosolized route has been used most frequently for RSV infection. Historically, pooled hyperimmune RSV immunoglobulin has been proposed as an additional treatment, but this has been superseded by the anti-RSV monoclonal antibody, palivizumab. Combinations of inhaled ribavirin and intravenous palivizumab have shown encouraging results. Palivizumab has been shown to be safe and well tolerated in patients undergoing HSCT, with a suggestion of better outcome (improved survival) when compared to ribavirin alone (Boeckh et al., 2001; Chavez-Bueno et al., 2007).

There are two groups of drugs available for the treatment of influenza – namely the adamantanes (effective against influenza A, e.g. rimantadine) and the neuraminidase inhibitors (effective against both influenza A & B, e.g. oseltamivir). In recent years, there has developed increasing resistance to adamantanes. The neuraminidase inhibitors have been shown to reduce the duration of illness by one day when given to an immunocompetent host within 48 h of onset of symptoms. Although there are few data on the benefit of treating influenza in an immunocompromised patient with a neuraminidase inhibitor, their use appears sensible and safe.

There is, thus far, no specific treatment available for rhinovirus, coronavirus or human metapneumovirus. Ribavirin has been proposed as a treatment for parainfluenza virus infection but evidence, so far, of benefit is disappointing. Although

there is little clinical data on the use of ribavirin for measles pneumonitis, it does have in vitro activity and therefore, due to the high level of mortality with this condition, should be considered.

A review of respiratory viral infection in children with primary immune deficiencies in a HSCT unit found 22 of 73 patients admitted for HSCT had respiratory viral infection. Of these, 11 had paramyxoviruses (RSV or parainfluenza I–IV), and were treated with aerosolized ribavirin and IVIG. Five of these patients also received nebulized immunoglobulin and corticosteroid. Three of these five survived, compared to two out of the six who did not receive nebulized treatment. It was concluded that the nebulized treatment was well tolerated and could be a useful adjunctive therapy (Crooks et al., 2000).

In children who have undergone HSCT, infection and inflammation can become inextricably interwoven to generate pneumonitis. In this case, in addition to the need for anti-infective agents, immunomodulation will be required through agents such as steroids, IVIG and anti-tumour necrosis factor monoclonal antibodies.

5 Gut

The gastrointestinal tract is also exposed to a wide variety of organisms and viruses which are of particular concern in the immunocompromised child, notably enteroviruses, adenovirus, rotavirus, caliciviruses, but also protozoa, mainly *Cryptosporidium* and *Giardia*. Presentation is most commonly with diarrhea and vomiting, which may protracted. *Cryptosporidium* can also be responsible for ascending cholangitis and liver disease. In some cases, identification of the causative organism can be difficult. Culture may be required to identify some viruses. PCR can be useful, for example, for adenovirus and is more sensitive than microscopy alone in detecting *Cryptosporidium*.

Prevention of transmission between immunocompromised patients is essential. There must be strict adherence to infection-control policies to prevent hospital wards from becoming sources of infection. One study looking at the extent of gastroenteric virus contamination in a pediatric-primary immunodeficiency ward and a general pediatric ward found viruses on 17 and 19% of environmental swabs, respectively. Interestingly, these were contaminating objects used by parents rather than staff – for example the parents' room television, the parents' toilet tap and the microwave used by parents on the pediatric-primary immunodeficiency ward (Gallimore et al., 2008). This highlights the importance of ensuring that parents and visitors, as well as staff, comply with hand washing and infection control measures.

5.1 Viruses

Rotavirus infection, which is usually relatively mild and self-limiting in the immunocompetent, can lead to persistent vomiting and diarrhea and, if untreated, severe malnutrition, in the immunocompromised. It can be identified in stool by

using enzyme immunoassay and may also be identified on electron microscopy. There is no specific treatment. Fluid and electrolyte management is important. Orally administered immunoglobulin has been used in some cases.

Caliciviruses, namely Noroviruses and Sapoviruses can also cause significant problems in the immunocompromised. Symptomatic infection and virus shedding can be prolonged; for example, one case report of a child undergoing HSCT for cartilage hair hypoplasia demonstrated Norovirus shedding for 156 days following transplant, during the period of immune reconstitution. The child was symptomatic throughout this time (Gallimore et al., 2004). Again, there is no specific treatment but meticulous management of fluids, electrolytes and nutritional support is essential, allowing time for immune reconstitution and consequent viral clearance.

Adenovirus will be discussed in more detail in the section on disseminated infection in the immunocompromised.

5.2 *Cryptosporidium*

Cryptosporidium species are oocyst-forming protozoa that cause watery diarrhea which can result in severe dehydration and even death, if not treated. Disease is normally confined to the gastrointestinal tract, but there is a risk of biliary tree, pulmonary or even disseminated disease in the immunocompromised. Infection may be diagnosed on identification of oocysts by microscopy. Enzyme immunoassays have also been used and PCR, too, can be helpful.

Treatment of *Cryptosporidium* infection can be difficult and a number of agents have been proposed, including nitazoxanide, paromomycin, rifabutin and the macrolides. Evidence is limited but a recent review has indicated that nitazoxanide may reduce parasite load and therefore be useful (Abubakar et al., 2007). In the authors' experience, azithromycin and nitazoxanide are safer options in post-HSCT patients, as paromomycin has been associated with significant hearing loss, particularly when given with ciclosporin. Supportive care remains essential. In those with HIV, anti-retroviral therapy, with its associated improvement in CD4 count, can result in improvement in the *Cryptosporidium* infection.

5.3 *Giardia*

Giardia intestinalis is a flagellate protozoan that exists in trophozoite or cyst forms. The cysts are the infective form. Children with humoral immunodeficiencies are particularly at risk of chronic symptomatic infection, with foul-smelling stool, abdominal distension and anorexia. Cysts may be identified on stool microscopy or by using immunofluorescent antibody testing. Treatment is with metronidazole, tinidazole or nitazoxanide. It may be necessary to use combination therapy in the immunocompromised if they have failed to respond to single-agent treatment.

6 Disseminated Infection

Disseminated viral infection in the immunocompromised is of particular concern. The most significant culprits are adenovirus and members of the human herpes virus: CMV, EBV, HHV6, HSV and VZV. These can affect the lungs, gastrointestinal tract and brain, resulting in a variety of symptoms. Reactivation of latent herpes viral infection is more common than primary infection after SOT or HSCT. Investigation using PCR techniques allows early diagnosis and quantification of viral load, and is now possible for adenovirus, CMV, EBV and HHV6. Prophylaxis to prevent CMV and HSV reactivation is used for children undergoing HSCT and many SOTs. Surveillance in high-risk patients enables pre-emptive treatment to be given before damaging disease occurs. Treatment will depend on the causative virus.

6.1 Adenovirus

Adenovirus is usually responsible for relatively minor upper respiratory tract or gastrointestinal infection but can result in life-threatening pneumonia, meningitis, encephalitis and disseminated disease in the immunocompromised. Those most at risk are patients who receive allogeneic bone marrow transplant, those with active graft versus host disease and those who receive total body irradiation. There are a number of different species of adenovirus, and these are divided into serotypes, some of which are primarily associated with the respiratory tract, while others have a predilection for the gastrointestinal tract. Young children are particularly vulnerable, as they often carry adenovirus in their gastrointestinal tract, predisposing them to reactivation and dissemination when they become immunocompromised. In view of this, screening can be important in the immunocompromised and adenovirus is usually identified in urine, stool, or sometimes respiratory secretions prior to being identified in blood. A study of 132 patients undergoing HSCT were screened for adenovirus in stool, urine, on throat swab and in peripheral blood during the post transplant period. 27% had a positive adenoviral PCR on at least one screening test, but this was not associated with clinical signs unless it was detected in peripheral blood and, even then, there was a median delay of 3 weeks from first detection of adenovirus until the patient demonstrated clinical signs. In one study, mortality was as high as 82% in those with adenovirus detected on peripheral blood. This highlights the importance of early recognition and consideration of pre-emptive use of antivirals (Lion et al., 2003).

Successful treatment of adenovirus infection has so far been limited. The most widely used agents are cidofovir or ribavirin, which may be given together with IVIG. Although cidofovir has potent nephrotoxic effects, these can be greatly reduced by the concurrent use of intravenous hyperhydration and probenecid. Cidofovir has been shown to be more effective in adenovirus and is now considered the best first-line treatment. Data on the clinical effectiveness of ribavirin in adenoviral infections are more conflicting. In vitro data suggest that ribavirin

alone has activity against subgenus C serotypes. In a post-HSCT patient with adenoviral infection, immune suppression should be reduced as much as possible, as T-cell immune reconstitution is very important for viral elimination.

6.2 Cytomegalovirus

CMV infection is often asymptomatic in the immunocompetent; however, in the immunocompromised it can lead to pneumonia, colitis and retinitis. CMV persists in a latent form after primary infection and can result in reactivation in someone who later becomes immunosuppressed – for example, when undergoing HSCT. CMV can be identified from respiratory secretions, urine and blood. As with adenovirus, PCR screening may be useful in identifying the virus before a child becomes symptomatic, especially in cases where reactivation is likely with immunosuppression. Treatment is usually with intravenous ganciclovir, with foscarnet or cidofovir as second-line treatment. Oral valganciclovir is very well absorbed and is also now an option for treatment. Foscarnet has also been used in cases of children undergoing HSCT to avoid the myelosuppressive effects of ganciclovir. IVIG should be used alongside antiviral therapy. There has been one case report of ganciclovir- and foscarnet-resistant CMV being successfully treated with artesunate (Shapira et al., 2008). There is also interest in the new antiviral agent maribavir for resistant CMV.

6.3 Epstein Barr Virus

EBV is associated with lymphoproliferative disorders in the immunocompromised. Replication of EBV in B cells is usually inhibited by natural killer cells, antibody-dependent cell cytotoxicity and T-cell cytotoxic responses. Therefore, children with cellular immune deficiencies are at risk of uncontrolled lymphoproliferation. Those at particular risk are children who are transplant recipients, both SOT or HSCT, and those with HIV. EBV can be detected in blood by PCR and viral load can be monitored. Alongside monitoring of the virus, it is important to monitor for signs of lymphoproliferation, both clinically and biochemically. Biopsy of suspicious lesions is often needed to make a diagnosis.

EBV infection requires treatment if it causes B lymphoproliferation or post-transplant lymphoproliferative disease (PTLD). This may take the form of the anti-CD20 monoclonal antibody rituximab, chemotherapy or radiotherapy. Decreasing immunosuppression whenever possible in a post-transplant patient is very important. More recently there have been encouraging results from work with cytotoxic T-cell therapy in PTLD. This involves the infusion of EBV-specific cytotoxic T lymphocytes (CTLs) generated from EBV sero-positive blood donors. In one recent multi-center study, 33 patients who had failed conventional therapy were recruited and monitored for response: 14 patients achieved complete remission while three showed a partial response (Haque et al., 2007).

6.4 Human Herpes Virus 6

Primary HHV6 infection in the immunocompetent host leads to the typical clinical picture of roseola or a non-specific febrile illness. The virus remains latent after primary infection and therefore, similar to CMV, can reactivate in immunocompromised states. The importance of HHV6 as a pathogen in the immunocompromised is probably underestimated, and many labs do not screen for infection; thus, many infections may not be recognized. HHV6 can cause fever, rash, hepatitis, pneumonia and encephalitis, as well as bone marrow suppression. HHV6 also appears to have synergistic effects and interactions with other infectious agents, such as CMV, adenovirus and fungi. It can be identified and quantified on blood samples by PCR. Treatment, where necessary, is with intravenous ganciclovir or foscarnet.

6.5 Varicella Zoster Virus

Primary varicella infection results in chickenpox, a common and generally self-limiting childhood illness. In the immunocompromised, there is a significant risk of both primary or reactivated disease becoming disseminated. This is particularly associated with T lymphocyte defects. VZV is the second most common cause of viral pneumonitis in children with AIDS. It should be remembered that fatal VZV infection has been reported in cases where the only immunosuppressant medication has been corticosteroids at a dose of 1 mg/kg/day of prednisolone for 2 weeks. The virus can be identified from vesicular fluid. Treatment is usually in the form of intravenous aciclovir, but, oral valaciclovir is a useful alternative in older children.

An important area to consider in relation to VZV infection is that of post-exposure prophylaxis. Although long-term prophylaxis for VZV is not usually recommended, post-exposure prophylaxis in non-immune immunocompromised children is important. Two options are available. The most widely used is varicella zoster immunoglobulin (VZIG). However, due to a shortage of VZIG a few years ago, oral aciclovir was reconsidered and has been shown to be effective. It must be remembered, however, that aciclovir has low bioavailability when given orally and requires multiple daily dosing. It may be more appropriate to consider the oral pro-drug valaciclovir, which has been shown to be effective and well tolerated (Nadal et al., 2002). Further work to clarify the best prophylactic and pre-emptive treatment regimens is needed.

7 Fungal Infections

Fungal infections must be considered in specific circumstances; for example, in those who are neutropenic (where risk increases exponentially with duration of neutropenia), those on steroids and those with graft versus host disease. *Candida* and *Aspergillus* are of particular interest in children who have undergone HSCT.

Symptoms that should raise the suspicion of fungal infection are persistent fevers unresponsive to antibiotics, skin nodules, chest pain and radiological evidence of infection crossing tissue planes. *Candida* is most commonly associated with CVC infection but can also cause disseminated disease. *Aspergillus* infection can have an insidious onset, frequently affecting the respiratory tract but then spreading to involve other areas such as the spine and intracranial cavity. Investigation and diagnosis remain difficult and may require antigen testing, PCR, cross-sectional imaging and biopsy of suspicious lesions/areas.

7.1 Candidiasis

Persistent mucocutaneous candidiasis is seen in patients with defects in T-cell function and may be a presenting feature for HIV infection or primary immune deficiency. Disseminated infection can involve almost any organ or any anatomical site and can be rapidly fatal. It is a particular concern in patients with CVC, especially those receiving multiple infusions and/or parenteral nutrition. There are a number of different *Candida* species that can result in disseminated infection. *Candia albicans* is the most common but *C. parapsilosis*, *C. glabrata*, *C. tropicalis* and *C. krusei* are increasingly common (Fig. 2). Diagnosis may be difficult, as blood cultures are not always positive. However, identification can be made by microscopy of biopsy specimens. Suspicious lesions, which are often found in organs such as the liver, kidney, spleen and brain, are best identified by cross-section imaging. PCR techniques have been developed, as well as detection of antigen from the fungal cell wall (Mannan). However, these techniques are not as yet wholly reliable. There are a number of agents available for treatment, including amphotericin B, caspofugin or an azole, such as voriconazole. Prolonged treatment is usually required and if there is a CVC

Species	Potential resistance
<i>C. albicans</i>	Resistance remains rare
<i>c. glabrata</i>	Fluconazole resistance in up to 20% cases, decreased susceptibility to other azoles; some cases of amphotericin B resistance
<i>C. parapsilosis</i>	Decreased susceptibility to caspofungin
<i>C. tropicalis</i>	
<i>C. krusei</i>	Resistant to ketoconazole and fluconazole; decreased susceptibility to itraconazole and amphotericin B
<i>C. guilliermondi</i>	Decreased suscepability to fluconazole
<i>C. lusitanae</i>	Generally resistant to amphotericin B
<i>C. rugosa</i>	Resistant to fluconazole in up to 60% cases; resistant to voriconazole

Fig. 2 *Candida* species and potential resistance to antifungal agents