Asymmetric Synthesis with Chemical and Biological Methods

Edited by Dieter Enders and Karl-Erich Jaeger



WILEY-VCH Verlag GmbH & Co. KGaA

Asymmetric Synthesis with Chemical and Biological Methods

Edited by Dieter Enders and Karl-Erich Jaeger

1807–2007 Knowledge for Generations

Each generation has its unique needs and aspirations. When Charles Wiley first opened his small printing shop in lower Manhattan in 1807, it was a generation of boundless potential searching for an identity. And we were there, helping to define a new American literary tradition. Over half a century later, in the midst of the Second Industrial Revolution, it was a generation focused on building the future. Once again, we were there, supplying the critical scientific, technical, and engineering knowledge that helped frame the world. Throughout the 20th Century, and into the new millennium, nations began to reach out beyond their own borders and a new international community was born. Wiley was there, expanding its operations around the world to enable a global exchange of ideas, opinions, and know-how.

For 200 years, Wiley has been an integral part of each generation's journey, enabling the flow of information and understanding necessary to meet their needs and fulfill their aspirations. Today, bold new technologies are changing the way we live and learn. Wiley will be there, providing you the must-have knowledge you need to imagine new worlds, new possibilities, and new opportunities.

Generations come and go, but you can always count on Wiley to provide you the knowledge you need, when and where you need it!

William J. Resce

William J. Pesce President and Chief Executive Officer

The Broth Willy

Peter Booth Wiley Chairman of the Board

Asymmetric Synthesis with Chemical and Biological Methods

Edited by Dieter Enders and Karl-Erich Jaeger



WILEY-VCH Verlag GmbH & Co. KGaA

The Editors

Prof. Dr. Dieter Enders Institut für Organische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Prof. Dr. Karl-Erich Jaeger Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany All books published by Wiley-VCH are carefully produced. Nevertheless, authors, editors, and publisher do not warrant the information contained in these books, including this book, to be free of errors. Readers are advised to keep in mind that statements, data, illustrations, procedural details or other items may inadvertently be inaccurate.

Library of Congress Card No.: applied for

British Library Cataloguing-in-Publication Data

A catalogue record for this book is available from the British Library.

Bibliographic information published by the Deutsche Nationalbibliothek

Die Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available in the Internet at <http://dnb.d-nb.de>.

© 2007 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim

All rights reserved (including those of translation into other languages). No part of this book may be reproduced in any form – by photoprinting, microfilm, or any other means – nor transmitted or translated into a machine language without written permission from the publishers. Registered names, trademarks, etc. used in this book, even when not specifically marked as such, are not to be considered unprotected by law.

Typesetting SNP Best-set Typesetter Ltd., Hong Kong

 Printing
 betz-druck GmbH, Darmstadt

 Binding
 Litges & Dopf GmbH, Heppenheim

 Cover Design
 Adam Design, Weinheim

 Wiley Bicentennial Logo
 Richard J. Pacifico

 Printed in the Federal Republic of Germany

Printed on acid-free paper

ISBN: 978-3-527-31473-7

Foreword

Stereochemistry has been an important topic for more than a hundred years. Nevertheless, as far as the chemist's everyday life was concerned, it was mainly of interest to natural product chemists for most of the time. This changed in the 1950s when synthetic chemists, following the example of R. B. Woodward, G. Stork and others, began to boldly address complex natural product targets. At this time the racemic compound was targeted, i.e. it was diastereoselectivity that counted. In the 1970s it became increasingly clear that biological activities of enantiomers could differ to the extent that one member of a pair is toxic or generally harmful. In this respect, the Contergan disaster was a signal. Pharmacologic testing of both individual enantiomers rather than the racemic agent became a common practice.

v

Demand created interest in the development of new methods for syntheses of *enantiomerically pure compounds*, termed *EPC*-syntheses by Dieter Seebach. First auxiliary controlled, stoichiometric asymmetric synthesis began to flourish in the second half of the 1970s. Dieter Enders, the spiritus rector of this book, with his SAMP/RAMP method, was one of the pioneers of this field. At about the same time the potential of enzyme-catalyzed enantioselective reactions became more and more visible, not least through pioneering early work of the M. R. Kula/C. Wandrey team in Düsseldorf/Jülich and of Hans-Joachim Gais in Darmstadt, later Aachen. In the 1980s, very few people dared to address transition metal catalyzed asymmetric synthesis. This changed in the 1990s after work of Kagan, Knowles, Sharpless, Noyori and others had shown that results useful for organic synthesis can be obtained.

Thus, in the early 1990s the stage was set for an Aachen/Jülich group of chemists to launch a collaborative program in the field of EPC synthesis that led to a prestigious Collaborative Research Center (Sonderforschungsbereich, SFB) "Asymmetric Syntheses with Chemical and Biological Methods", which was to become operative for 12 years (1994–2005). In this book the main results, obtained by ca. 20 research groups, are reviewed. A very large and colorful landscape of methods and applications is presented.

The first part of the book is devoted to auxiliary controlled reactions using the SAMP/ RAMP method (D. Enders) and metallated allylsulfoximines (H.-J. Gais).

VI Foreword

Syntheses of an impressive array of natural products, including medicinally interesting alkaloids, underline the usefulness of these methods. The following part deals with enantioselective reactions catalyzed by transition metal complexes. Chiral ligands with a modular make-up are of crucial importance here and many new classes are described: phosphines containing an arenechromium-tricarbonyl moiety ("Daniphos" ligands, A. Salzer), phosphaferrocenes (C. Ganter), sulfoximine-based N,N- and P,N-ligands (C. Bolm), P,C- and N,O-ligands containing a [2,2]paracyclophane skeleton (C. Bolm, S. Bräse) and phosphines based on dihydroquinolines ("Quinaphos" ligands, W. Leitner). Catalyst immobilization on or in a zeolite matrix was much debated in the SFB; finally, W. F. Hoelderich's group has been able to obtain highly active, reusable hydrogenation as well as Jacobsen type epoxidation catalysts.

The next part of the book deals with enzyme catalysis and bioorganic synthesis. An important aim of this research has been the preparation of enantiomerically pure small molecules that are useful in general organic synthesis and as intermediates in drug process synthesis. It is apparent that there has been fruitful and remarkably successful collaboration between ca. 10 groups, led by established as well as junior group leaders. The first three articles, with authors from the groups of K.-E. Jaeger, M.-R. Kula, M. Pohl, M. Müller and G. A. Sprenger, deal with applications of techniques of enzyme biochemistry, for example site-directed mutagenesis and directed evolution based on recombinant DNA technology. The following articles describe asymmetric syntheses of a large variety of chiral alcohols using *R*-specific alcohol dehydrogenases (W. Hummel), aldolases and related types of C-C bond forming enzymes (W.-D. Fessner) as well as sucrose synthase I (L. Elling). An article naming 17 authors on asymmetric synthesis of 1,3-diols and propargylic alcohols concludes the section.

An asset of the Aachen/Jülich bioorganic synthesis approach is technology transfer, which is testified by no less than five start-up companies. Scale-up requires stable and highly efficient enzymes as well as appropriate reaction technology. The development of membrane reactors has been a key to success. Reaction technology is outlined by C. Wandrey and co-workers in the final article.

Reading this book is worthwhile for anybody seeking an impression of the state of the art of the entire field of asymmetric synthesis. A lot of interesting material is offered to the expert from academia or industry as well as to the student looking for an interesting field of graduate research.

Günter Helmchen

Contents

Foreword V

Preface XVII

List of Contributors XIX

1 Stoichiometric Asymmetric Synthesis 1

- 1.1 Development of Novel Enantioselective Synthetic Methods 1 Dieter Enders and Wolfgang Bettray
- 1.1.1 Introduction 1
- 1.1.2 α-Silyl Ketone-Controlled Asymmetric Syntheses 1
- 1.1.2.1 Regio- and Enantioselective α -Fluorination of Ketones 2
- 1.1.2.2 α-Silyl Controlled Asymmetric Mannich Reactions 3
- 1.1.3 Asymmetric Hetero-Michael Additions 5
- 1.1.3.1 Asymmetric Aza-Michael Additions 5
- 1.1.3.2 Asymmetric Oxa-Michael Additions 10
- 1.1.3.3 Asymmetric Phospha-Michael-Additions 11
- 1.1.4 Asymmetric Syntheses with Lithiated α-Aminonitriles 14
- 1.1.4.1 Asymmetric Nucleophilic α-Aminoacylation 14
- 1.1.4.2 Asymmetric Nucleophilic Alkenoylation of Aldehydes 16
- 1.1.5 Asymmetric Electrophilic α-Substitution of Lactones and Lactams 18
- 1.1.6 Asymmetric Synthesis of α-Phosphino Ketones and 2-Phosphino Alcohols 22
- 1.1.7 Asymmetric Synthesis of 1,3-Diols and *anti*-1,3-Polyols 24
- Asymmetric Synthesis of α-Substituted Sulfonamides and Sulfonates 26
- 1.2 Asymmetric Synthesis of Natural Products Employing the SAMP/ RAMP Hydrazone Methodology 38 Dieter Enders and Wolfgang Bettray
- 1.2.1 Introduction 38
- 1.2.2 Stigmatellin A 38

- VIII Contents
 - 1.2.3 Callistatin A 41
 - 1.2.4 Dehydroiridodiol(dial) and Neonepetalactone 51
 - First Enantioselective Synthesis of Dendrobatid Alkaloids Indolizidine 209I and 223J 53
 - 1.2.6 Efficient Synthesis of (2S,12'R)-2-(12'-Aminotridecyl)pyrrolidine, a Defense Alkaloid of the Mexican Bean Beetle 57
 - 1.2.7 2-epi-Deoxoprosopinine 58
 - 1.2.8 Attenol A and B 62
 - 1.2.9 Asymmetric Synthesis of (+)- and (-)-Streptenol A 64
 - 1.2.10 Sordidin 66
 - 1.2.11 Prelactone B and V 69
 - 1.3 Asymmetric Synthesis Based on Sulfonimidoyl-Substituted Allyltitanium Complexes 75 Hans-Joachim Gais
 - 1.3.1 Introduction 75
 - 1.3.2 Hydroxyalkylation of Sulfonimidoyl-Substituted Allylltitanium Complexes 80
 - 1.3.2.1 Sulfonimidoyl-Substituted Bis(allyl)titanium Complexes 80
 - 1.3.2.2 Sulfonimidoyl-Substituted Mono(allyl)tris(diethylamino)titanium Complexes 82
 - 1.3.3 Aminoalkylation of Sulfonimidoyl-Substituted Allyltitanium Complexes 85
 - 1.3.3.1 Sulfonimidoyl-Substituted Bis(allyl)titanium Complexes 85
 - 1.3.3.2 Sulfonimidoyl-Substituted Mono(allyl)tris(diethylamino)titanium Complexes 86
 - 1.3.4 Structure and Reactivity of Sulfonimidoyl-Substituted Allyltitanium Complexes 88
 - 1.3.4.1 Sulfonimidoyl-Substituted Bis(allyl)titanium Complexes 88
 - 1.3.4.2 Sulfonimidoyl-Substituted Mono(allyl)titanium Complexes 91
 - 1.3.5 Asymmetric Synthesis of Homopropargyl Alcohols 95
 - 1.3.6 Asymmetric Synthesis of 2,3-Dihydrofurans 96
 - 1.3.7 Synthesis of Bicyclic Unsaturated Tetrahydrofurans 98
 - 1.3.8 Asymmetric Synthesis of Alkenyloxiranes 100
 - 1.3.9 Asymmetric Synthesis of Unsaturated Mono- and Bicyclic Prolines 102
 - 1.3.10 Asymmetric Synthesis of Bicyclic Amino Acids 105
 - 1.3.11 Asymmetric Synthesis of β-Amino Acids 108
 - 1.3.12 Conclusion 111
 - 1.4 The "Daniphos" Ligands: Synthesis and Catalytic Applications 115 Albrecht Salzer and Wolfgang Braun
 - 1.4.1 Introduction 115
 - 1.4.2 General Synthesis 116

- 1.4.3 Applications in Stereoselective Catalysis 120
- 1.4.3.1 Enantioselective Hydrogenations 120
- 1.4.3.2 Diastereoselective Hydrogenation of Folic Acid Ester 122
- 1.4.3.3 Enantioselective Isomerization of Geranylamine to Citronellal 124
- 1.4.3.4 Nucleophilic Asymmetric Ring-Opening of Oxabenzonorbornadiene 124
- 1.4.3.5 Enantioselective Suzuki Coupling 126
- 1.4.3.6 Asymmetric Hydrovinylation 126
- 1.4.3.7 Allylic Sulfonation 128
- 1.4.4 Conclusion 129
- 1.5 New Chiral Ligands Based on Substituted Heterometallocenes 130 Christian Ganter
- 1.5.1 Introduction 130
- 1.5.2 General Properties of Phosphaferrocenes 131
- 1.5.3 Synthesis of Phosphaferrocenes 132
- 1.5.4 Preparation of Bidentate P,P and P,N Ligands 133
- 1.5.5 Modification of the Backbone Structure 136
- 1.5.6 Cp–Phosphaferrocene Hybrid Systems 139
- 1.5.7 Catalytic Applications 145
- 1.5.8 Conclusion 146

2 Catalytic Asymmetric Synthesis 149

- 2.1 Chemical Methods 149
- 2.1.1 Sulfoximines as Ligands in Asymmetric Metal Catalysis 149 Carsten Bolm
- 2.1.1.1 Introduction 149
- 2.1.1.2 Development of Methods for Sulfoximine Modification 150
- 2.1.1.3 Sulfoximines as Ligands in Asymmetric Metal Catalysis 162
- 2.1.1.4 Conclusions 170
- 2.1.2 Catalyzed Asymmetric Aryl Transfer Reactions 176 Carsten Bolm
- 2.1.2.1 Introduction 176
- 2.1.2.2 Catalyst Design 177
- 2.1.2.3 Catalyzed Aryl Transfer Reactions 180
- 2.1.3 Substituted [2.2]Paracyclophane Derivatives as Efficient Ligands for Asymmetric 1,2- and 1,4-Addition Reactions 196 Stefan Bräse

- Contents
 - 2.1.3.1 [2.2]Paracyclophanes as Chiral Ligands 196
 - 2.1.3.2 Synthesis of [2.2]Paracyclophane Ligands 199
 - 2.1.3.2.1 Preparation of FHPC-, AHPC-, and BHPC-Based Imines 199
 - Structural Information on AHPC-Based Imines 2.1.3.2 199
 - 2.1.3.3 Asymmetric 1,2-Addition Reactions to Aryl Aldehydes 200
 - 2.1.3.3.1 Initial Considerations 200
 - 2.1.3.3.2 Asymmetric Addition Reactions to Aromatic Aldehydes: Scope of Substrates 203
 - 2.1.3.4 Asymmetric Addition Reactions to Aliphatic Aldehydes 205
 - 2.1.3.5 Addition of Alkenylzinc Reagents to Aldehydes 206
 - 2.1.3.6 Asymmetric Conjugate Addition Reactions 208
 - 2.1.3.7 Asymmetric Addition Reactions to Imines 208
 - 2.1.3.8 Asymmetric Addition Reactions on Solid Supports 212
 - 2.1.3.8.1 Applications 213
 - 2.1.3.9 Conclusions and Future Perspective 213
 - 2.1.4 Palladium-Catalyzed Allylic Alkylation of Sulfur and Oxygen Nucleophiles - Asymmetric Synthesis, Kinetic Resolution and Dynamic Kinetic Resolution 215 Hans-Joachim Gais
 - 2.1.4.1 Introduction 215
 - 2.1.4.2 Asymmetric Synthesis of Allylic Sulfones and Allylic Sulfides and Kinetic Resolution of Allylic Esters 216
 - 2.1.4.2.1 Kinetic Resolution 216
 - 2.1.4.2.2 Selectivity 220
 - 2.1.4.2.3 Asymmetric Synthesis 220
 - 2.1.4.2.4 Synthesis of Enantiopure Allylic Alcohols 224
 - Asymmetric Rearrangment and Kinetic Resolution of Allylic 2.1.4.3 Sulfinates 225
 - 2.1.4.3.1 Introduction 225
 - 2.1.4.3.2 Synthesis of Racemic Allylic Sulfinates 225
 - 2.1.4.3.3 Pd-Catalyzed Rearrangement 226
 - 2.1.4.3.4 Kinetic Resolution 227
 - 2.1.4.3.5 Mechanistic Considerations 228
 - 2.1.4.4 Asymmetric Rearrangment of Allylic Thiocarbamates 229
 - 2.1.4.4.1 Introduction 229
 - 2.1.4.4.2 Synthesis of Racemic O-Allylic Thiocarbamates 229
 - 2.1.4.4.3 Acyclic Carbamates 229
 - 2.1.4.4.4 Cvclic Carbamates 231
 - 2.1.4.4.5 Mechanistic Considerations 232
 - 2.1.4.4.6 Synthesis of Allylic Sulfides 232
 - Asymmetric Synthesis of Allylic Thioesters and Kinetic Resolution of 2.1.4.5 Allylic Esters 233
 - 2.1.4.5.1 Introduction 233

- 2.1.4.5.2 Asymmetric Synthesis of Allylic Thioesters 234
- 2.1.4.5.3 Kinetic Resolution of Allylic Esters 235
- 2.1.4.5.4 Memory Effect and Dynamic Kinetic Resolution of the Five-Membered Cyclic Acetate 238
- 2.1.4.5.5 Asymmetric Synthesis of Cyclopentenyl Thioacetate 242
- 2.1.4.6 Kinetic and Dynamic Kinetic Resolution of Allylic Alcohols 242
- 2.1.4.6.1 Introduction 242
- 2.1.4.6.2 Asymmetric Synthesis of Symmetrical Allylic Alcohols 242
- 2.1.4.6.3 Asymmetric Synthesis of Unsymmetrical Allylic Alcohols 244
- 2.1.4.6.4 Asymmetric Synthesis of a Prostaglandin Building Block 245
- 2.1.4.6.5 Investigation of an Unsaturated Analogue of BPA 245
- 2.1.4.7 Conclusions 246
- 2.1.5 The QUINAPHOS Ligand Family and its Application in Asymmetric Catalysis 250
 Giancarlo Franciò, Felice Faraone, and Walter Leitner
- 2.1.5.1 Introduction 250
- 2.1.5.2 Synthetic Strategy 252
- 2.1.5.3 Stereochemistry and Coordination Properties 254
- 2.1.5.3.1 Free Ligands 254
- 2.1.5.3.2 Complexes 256
- 2.1.5.4 Catalytic Applications 261
- 2.1.5.4.1 Rhodium-Catalyzed Asymmetric Hydroformylation of Styrene 261
- 2.1.5.4.2 Rhodium-Catalyzed Asymmetric Hydrogenation of Functionalized Alkenes 263
- 2.1.5.4.3 Ruthenium-Catalyzed Asymmetric Hydrogenation of Aromatic Ketones 265
- 2.1.5.4.4 Copper-Catalyzed Enantioselective Conjugate Addition of Diethylzinc to Enones 267
- 2.1.5.4.5 Nickel-Catalyzed Asymmetric Hydrovinylation 268
- 2.1.5.4.6 Nickel-Catalyzed Cycloisomerization of 1,6-Dienes 270
- 2.1.5.5 Conclusions 273
- 2.1.6 Immobilization of Transition Metal Complexes and Their Application to Enantioselective Catalysis 277 Adrian Crosman, Carmen Schuster, Hans-Hermann Wagner, Melinda Batorfi, Jairo Cubillos, and Wolfgang Hölderich
- 2.1.6.1 Introduction 277
- 2.1.6.2 Immobilized Rh Diphosphino Complexes as Catalysts for Asymmetric Hydrogenation 278
- 2.1.6.2.1 Preparation and Characterization of the Immobilized Rh–Diphosphine Complexes 279

- XII Contents
 - 2.1.6.2.2 Enantioselective Hydrogenation over Immobilized Rhodium Diphosphine Complexes 282
 - 2.1.6.3 Heterogeneous Asymmetric Epoxidation of Olefins over Jacobsen's Catalyst Immobilized in Inorganic Porous Materials 284
 - 2.1.6.3.1 Preparation and Characterization of Immobilized Jacobsen's Catalysts 285
 - 2.1.6.3.2 Epoxidation of Olefins over Immobilized Jacobsen Catalysts 287
 - 2.1.6.4 Novel Heterogenized Catalysts for Asymmetric Ring-Opening Reactions of Epoxides 291
 - 2.1.6.4.1 Synthesis and Characterization of the Heterogenized Catalysts 291
 - 2.1.6.4.2 Asymmetric Ring Opening of Epoxides over New Heterogenized Catalysts 293
 - 2.1.6.5 Conclusions 295
 - 2.2 Biological Methods 298
 - 2.2.1 Directed Evolution to Increase the Substrate Range of Benzoylformate Decarboxylase from Pseudomonas putida 298 Marion Wendorff, Thorsten Eggert, Martina Pohl, Carola Dresen, Michael Müller, and Karl-Erich Jaeger
 - 2.2.1.1 Introduction 298
 - 2.2.1.2 Materials and Methods 300
 - 2.2.1.2.1 Reagents 300
 - 2.2.1.2.2 Construction of Strains for Heterologous Expression of BFD and BAL 300
 - 2.2.1.2.3 Polymerase Chain Reactions 301
 - 2.2.1.2.4 Generation of a BFD Variant Library by Random Mutagenesis 302
 - 2.2.1.2.5 High-Throughput Screening for Carboligation Activity with the Substrates Benzaldehyde and Dimethoxyacetaldehyde 303
 - 2.2.1.2.6 Expression and Purification of BFD Variants 303
 - 2.2.1.2.7 Protein Analysis Methods 304
 - 2.2.1.2.8 Enzyme Activity Assays 304
 - 2.2.1.3 Results and Discussion 304
 - 2.2.1.3.1 Overexpression of BFD in Escherichia coli 304
 - 2.2.1.3.2 Random Mutagenesis of BFD Variant L476Q 305
 - 2.2.1.3.3 Development of a High-Throughput Screening Assay for Carboligase Activity 305
 - 2.2.1.3.4 Identification of a BFD Variant with an Optimized Acceptor Aldehyde Spectrum 306
 - 2.2.1.3.5 Biochemical Characterization of the BFD Variants 308
 - 2.2.1.3.6 Decreased Benzoyl Formate Decarboxylation Activity of Variant 55E4 308

- 2.2.1.3.7 Formation of 2-Hydroxy-3,3-dimethoxypropiophenone and Benzoin 308
- 2.2.1.3.8 Enantioselectivity of the Carboligation Reaction 310
- 2.2.1.4 Conclusions 311
- 2.2.2 C–C-Bonding Microbial Enzymes: Thiamine Diphosphate-Dependent Enzymes and Class I Aldolases 312 Georg A. Sprenger, Melanie Schürmann, Martin Schürmann, Sandra Johnen, Gerda Sprenger, Hermann Sahm, Tomoyuki Inoue, and Ulrich Schörken
- 2.2.2.1 Introduction 312
- 2.2.2.2 Thiamine Diphosphate (ThDP)-Dependent Enzymes 312
- 2.2.2.1 Transketolase (TKT) 313
- 2.2.2.2.2 1-Deoxy-D-xylulose 5-Phosphate Synthase (DXS) 317
- 2.2.2.2.3 Phosphonopyruvate Decarboxylase (PPD) from *Streptomyces* viridochromogenes 318
- 2.2.2.3 Class I Aldolases 318
- 2.2.2.3.1 Transaldolase (TAL) 320
- 2.2.2.3.2 Fructose 6-Phosphate Aldolase (FSA) 321
- 2.2.2.4 Summary and Outlook 321
- 2.2.3 Enzymes for Carboligation 2-Ketoacid Decarboxylases and Hydroxynitrile Lyases 327 Martina Pohl, Holger Breittaupt, Bettina Frölich, Petra Heim, Hans Iding, Bettina Juchem, Petra Siegert, and Maria-Regina Kula
- 2.2.3.1 Introduction 327
- 2.2.3.2 2-Ketoacid Decarboxylases 327
- 2.2.3.2.1 Comparative Biochemical Characterization of Wild-Type PDC and BFD 328
- 2.2.3.2.2 Identification of Amino Acid Residues Relevant to Substrate Specificity and Enantioselectivity 330
- 2.2.3.2.3 Optimization of the Substrate Range of BFD by Site-Directed Mutagenesis 330
- 2.2.3.2.4 Optimization of Stability and Substrate Range of BFD by Directed Evolution 330
- 2.2.3.3 Hydroxynitrile Lyases 332
- 2.2.3.3.1 HNL from Sorghum bicolor 333
- 2.2.3.3.2 HNL from Linum usitatissimum 337
- 2.2.4 Preparative Syntheses of Chiral Alcohols using (R)-Specific Alcohol Dehydrogenases from Lactobacillus Strains 341 Andrea Weckbecker, Michael Müller, and Werner Hummel
- 2.2.4.1 Introduction 341
- 2.2.4.2 (R)-Specific Alcohol Dehydrogenase from Lactobacillus kefir 341

- 2.2.4.3 Comparison of (R)-Specific ADHs from L. kefir and L. brevis 342
- 2.2.4.4 Preparative Applications of ADHs from L. kefir and L. brevis 345
- 2.2.4.4.1 Synthesis of (R,R)-Diols 346
- 2.2.4.4.2 Synthesis of Enantiopure 1-Phenylpropane-1,2-diols 346
- 2.2.4.4.3 Synthesis of Enantiopure Propargylic Alcohols 346
- 2.2.4.4.4 Regioselective Reduction of *t*-Butyl 6-chloro-3,5-dioxohexanoate to the Corresponding Enantiopure (*S*)-5-Hydroxy Compound 346
- 2.2.4.5 Coenzyme Regeneration and the Construction and Use of "Designer Cells" 347
- 2.2.4.6 Discussion 349
- 2.2.5 Biocatalytic C–C Bond Formation in Asymmetric Synthesis 351 Wolf-Dieter Fessner
- 2.2.5.1 Introduction 351
- 2.2.5.2 Enzyme Mechanisms 352
- 2.2.5.2.1 Class II Aldolases 352
- 2.2.5.2.2 Class I Fructose 1,6-Bisphosphate Aldolase 355
- 2.2.5.2.3 Sialic Acid Synthase 355
- 2.2.5.2.4 Rhamnose Isomerase 356
- 2.2.5.3 New Synthetic Strategies 357
- 2.2.5.3.1 Sugar Phosphonates 357
- 2.2.5.3.2 Xylulose 5-Phosphate 359
- 2.2.5.3.3 RhuA Stereoselectivity 359
- 2.2.5.3.4 Aldolase Screening Assay 361
- 2.2.5.3.5 Aldose Synthesis 361
- 2.2.5.3.6 Tandem Chain Extension–Isomerization–Chain Extension 362
- 2.2.5.3.7 Tandem Bidirectional Chain Extensions 363
- 2.2.5.3.8 Non-Natural Sialoconjugates 369
- 2.2.5.4 Summary and Outlook 373
- 2.2.6 Exploring and Broadening the Biocatalytic Properties of Recombinant Sucrose Synthase 1 for the Synthesis of Sucrose Analogues 376 Lothar Elling
- 2.2.6.1 Introduction 376
- 2.2.6.2 Characteristics of Recombinant Sucrose Synthase 1 (SuSy1) Expressed in Saccharomyces cerevisiae 377
- 2.2.6.2.1 Expression and Purification of SuSy1 from Yeast 377
- 2.2.6.2.2 The Substrate Spectrum of SuSy1 from Yeast 378
- 2.2.6.3 Characteristics of Recombinant Sucrose Synthase 1 (SuSy1) Expressed in *Escherichia coli* 381
- 2.2.6.3.1 Expression and Purification of SuSy1 from E. coli 381
- 2.2.6.3.2 The Substrate Spectrum of SuSy1 from E. coli 382
- 2.2.6.4 Sucrose Synthase 1 Mutants Expressed in S. cerevisiae and E. coli 383
- 2.2.6.5 Outlook 384

- 2.2.7 Flexible Asymmetric Redox Reactions and C–C Bond Formation by Bioorganic Synthetic Strategies 386 Michael Müller, Michael Wolberg, Silke Bode, Ralf Feldmann, Petra Geilenkirchen, Thomas Schubert, Lydia Walter, Werner Hummel, Thomas Dünnwald, Ayhan S. Demir, Doris Kolter-Jung, Adam Nitsche, Pascal Dünkelmann, Annabel Cosp, Martina Pohl, Bettina Lingen, and Maria-Regina Kula
- 2.2.7.1 Introduction 386
- 2.2.7.2 Diversity-Oriented Access to 1,3-Diols Through Regio- and Enantioselective Reduction of 3,5-Dioxocarboxylates 386
- 2.2.7.2.1 Regio- and Enantioselective Enzymatic Reduction 387
- 2.2.7.2.2 Dynamic Kinetic Resolution 388
- 2.2.7.2.3 Stereoselective Access to 1,3-Diols by Diastereoselective Reduction 389
- 2.2.7.2.4 Nucleophilic Substitution of Chlorine 390
- 2.2.7.2.5 Application in Natural Product Syntheses 391
- 2.2.7.2.6 Discussion and Outlook 392
- 2.2.7.3 Chemo- and Enantioselective Reduction of Propargylic Ketones 395
- 2.2.7.3.1 Enantioselective Reduction of Aryl Alkynones 395
- 2.2.7.3.2 Synthesis of Enantiopure 3-Butyn-2-ol 396
- 2.2.7.3.3 Enzymatic Reduction of α-Halogenated Propargylic Ketones 397
- 2.2.7.3.4 Modification of α-Halogenated Propargylic Alcohols 398
- 2.2.7.3.5 Olefinic Substrates 399
- 2.2.7.3.6 Discussion and Outlook 401
- 2.2.7.4 Thiamine Diphosphate-Dependent Enzymes: Multi-purpose Catalysts in Asymmetric Synthesis 401
- 2.2.7.4.1 Formation of Chiral 2-Hydroxy Ketones Through BFD-Catalyzed Reactions 402
- 2.2.7.4.2 BAL as a Versatile Catalyst for C–C Bond Formation and Cleavage Reactions 405
- 2.2.7.4.3 Asymmetric Cross-Benzoin Condensation 407
- 2.2.7.4.4 Discussion and Outlook 408
- 2.2.7.5 Summary 409

3 Reaction Technology in Asymmetric Synthesis 415

- 3.1 Reaction Engineering in Asymmetric Synthesis 415 Stephan Lütz, Udo Kragl, Andreas Liese, and Christian Wandrey
- 3.1.1 Introduction 415
- 3.1.2 Membrane Reactors with Chemical Catalysts 418
- 3.1.3 Membrane Reactors with Biological Catalysts 420
- 3.1.3.1 Membrane Reactors with Whole Cells 420
- 3.1.3.2 Membrane Reactors with Isolated Enzymes 421
- 3.1.4 Two-Phase Systems 422
- 3.1.5 Conclusions 425

XVI Contents

- 3.2 Biocatalyzed Asymmetric Syntheses Using Gel-Stabilized Aqueous– Organic Two-Phase Systems 427 Marion B. Ansorge-Schumacher
- 3.2.1 Gel-Stabilized Two-Phase Systems 428
- 3.2.2 Benzoin Condensation with Entrapped Benzaldehyde Lyase 430
- 3.2.3 Reduction of Ketones with Entrapped Alcohol Dehydrogenase 432
- 3.2.4 Conclusion *433*

Index 435

Name Index 443

Preface

After the pioneering work of Louis Pasteur and Emil Fischer in the middle and at the end of the nineteenth century, respectively, it still took more than fifty years before chemists started to discuss transition state models together with polar and steric effects to gain more insight into the phenomenon of asymmetric induction. Even first observations in organic synthesis of enantioselectivities comparable to those of enzymes in the late fifties and sixties of the 20th century did not convince the chemical community and the term "asymmetric synthesis" was regarded a mechanistic curiosity rather than a practical way to synthesize compounds of high enantiomeric purity.

In the mid-seventies, with the development of generally applicable stoichiometric asymmetric syntheses, especially the Meyers oxazoline methodology as the first one, the scientific community began to believe that asymmetric synthesis really worked resulting in an explosive growth of this new field. Later on, and mainly driven by the fact that the biological activity of enantiomers is usually different, dozens of new chemical companies were founded all over the world in a newly created area called "chirotechnology".

Around that time and after intensive discussions several professors of the RWTH Aachen University and the nearby Jülich Research Center decided to apply at the German Research Council for a so-called Collaborative Research Center on the topic of asymmetric synthesis. Looking back, it was truly a seminal event when the Professors D. Enders, W. Keim, M.-R. Kula, H. Sahm and C. Wandrey stopped their cars at the highway station Köln-Frechen and nailed down the proposed research topic as "Asymmetric Synthesis with Chemical and Biological Methods". After Professor E. Winterfeldt, as an advisor, saw this new initiative "under a good star", indeed the new "Sonderforschungsbereich 380" was funded and started in 1994.

From the very beginning of this long term research endeavor, the aim has been to cover *all* aspects of the *entire* field of asymmetric synthesis including stoichiometric and catalytic asymmetric syntheses with chemical and biological methods as well as the development of new reaction technologies. The interdisciplinary cooperation among the areas of classical organic and inorganic chemistry as well as technical chemistry (RWTH Aachen University) and the various fields of enzyme technology and biotechnology (Research Center Jülich, HHU Düsseldorf) resulted in efficient asymmetric syntheses of synthetic building blocks, fine chemicals, natural products and biologically active compounds in general. Mechanistic and theoretical aspects, organic synthesis, organometallic chemistry, homogeneous and heterogeneous transition metal catalysis, microbiology, enzyme- and biotechnology were all employed and used for stereoselective C-H-, C-C-, and C-heteroatom bond formations.

Besides the scientific success of this Collaborative Research Center as measured in publications, patents and foundation of start-up companies, it should be mentioned that a high percentage of the younger scientific members received and accepted calls for full professorships including D. Vogt (Eindhoven), W.-D. Fessner (Darmstadt), U. Kragl (Rostock), A. Liese (Hamburg), S. Bräse (Karlsruhe), G. Sprenger (Stuttgart) and M. Müller (Freiburg) and also associate professorships as C. Ganter (Düsseldorf), L. Elling (Aachen), M. Ansorge-Schumacher (Berlin) and M. Pohl (Privatdozent, Düsseldorf). A highlight during the twelve years of funding was the "Deutsche Zukunftspreis" awarded by the Federal President of Germany to Prof. Kula and Dr. Pohl and presented in a spectacular nationwide television show broadcasted from Berlin in 2002. Professor Maria-Regina Kula, herself being a chemist, was always aware of the necessity to combine biological and chemical catalytic methods. As her 70th birthday coincides with the appearance of this book, the editors would like to express their warm congratulations and best wishes for her future.

We thank the German Research Council ("Deutsche Forschungsgemeinschaft") for the generous financial support of the Collaborative Research Center "Sonderforschungsbereich, SFB 380" over a period of twelve years. In particular, we are thankful to Dr. H. H. Lindner and Dr. A. Pollex-Krüger as well as Dr. W. Rohe, Dr. P. Schmitz-Möller and Dr. H. Schruff for their organizational help during the course of the priority programme. In addition, on behalf of all participants of the Collaborative Research Center, we would like to thank the scientific referees, the Professors M. Ballauff (Bayreuth), J. E. Bäckvall (Stockholm), A. Böck (München), H. Brunner (Regensburg), H. Buchholz (Erlangen-Nürnberg), W. Buckel (Marburg), G. Dziuk (Freiburg), F. Effenberger (Stuttgart), H. Eschrig (Dresden), H. Fischer (Konstanz), W. Francke (Hamburg), G. Gottschalk (Göttingen), H. Griengl (Graz), G. Helmchen (Heidelberg), U. Kazmaier (Saarbrücken), H. Kessler (München), H. Kunz (Mainz), E. P. Kündig (Genf), J. Mulzer (Wien), H.-U. Reißig (Berlin), K. Sandhoff (Bonn), G. Schulz-Eckloff (Bremen), H. Simon (München), W. Spiess (Mainz), J. Thiem (Hamburg), H. Tschesche (Bielefeld), H. Vahrenkamp (Freiburg), and H. Waldmann (Dortmund) for their help, advice and the many fruitful discussions.

We hope that this book will be useful and a source of inspiration for all those interested in the chemical, biological and technical aspects of asymmetric synthesis in general and will stimulate new ideas and research activities among the young scientists in this rapidly growing field.

Aachen / Jülich, December 2006

Dieter Enders Karl-Erich Jaeger

List of Contributors

Prof. Dr. Marion-B.

Ansorge-Schumacher Technische Universität Berlin Institut für Chemie / Enzymtechnologie Straße des 17. Juni 124 10623 Berlin Germany

Melinda Batorfi

Institut für Brennstoffchemie und Physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Dr. Wolfgang Bettray

Institut für Organische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Silke Bode

Lehrstuhl für Pharmazeutische und Medizinische Chemie Institut für Pharmazeutische Wissenschaften Albert-Ludwigs-Universität Freiburg Albertstr. 25 79104 Freiburg Germany

Prof. Dr. Carsten Bolm

Institut für Organische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Prof. Dr. Stefan Bräse

Institut für Organische Chemie Universität Karlsruhe (TH) Fritz-Haber-Weg 6 76131 Karlsruhe Germany

Dr. Wolfgang Braun

Institut für Anorganische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

XX List of Contributors

Dr. Holger Breithaupt

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Annabel Cosp

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Adrian Crosman

Institut für Brennstoffchemie und Physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Dr. Jairo Cubillos

Institut für Brennstoffchemie und Physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Prof. Dr. Ayhan S. Demir

Department of Chemistry Middle East Technical University 06531 Ankara Türkei

Carola Dresen

Lehrstuhl für Pharmazeutische und Medizinische Chemie Institut für Pharmazeutische Wissenschaften Albert-Ludwigs-Universität Freiburg Albertstr. 25 79104 Freiburg Germany

Dr. Pascal Dünkelmann

Lehrstuhl für Pharmazeutische und Medizinische Chemie Institut für Pharmazeutische Wissenschaften Albert-Ludwigs-Universität Freiburg Albertstr. 25 79104 Freiburg Germany

Dr. Thomas Dünnwald

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Dr. Thorsten Eggert

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Prof. Dr. Lothar Elling

Lehr- und Forschungsgebiet Biomaterialien Institut für Biotechnologie und Helmholtz-Institut für Biomedizinische Technik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Prof. Dr. Dieter Enders

Institut für Organische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Prof. Dr. Felice Faraone

Dipartimento di Chimica Inorganica, Chimica Analitica e Chimica Fisica Università degli studi di Messina Salita Sperone 31 (vill. S. Agata) 98166 Messina, Italy

Ralf Feldmann

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Prof. Dr. Wolf-Dieter Fessner

Institut für Organische Chemie und Biochemie TU Darmstadt Petersenstr. 22 64287 Darmstadt Germany

Dr. Giancarlo Franciò

Institut für Technische und Makromolekulare Chemie RWTH Aachen Worringerweg 1 52074 Aachen Germany

Prof. Dr. Hans-Joachim Gais

Institut für Organische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Prof. Dr. Christian Ganter

Institut für Anorganische Chemie der HHU Düsseldorf Universitätsstr. 1 40225 Düsseldorf Germany

Petra Geilenkirchen

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Dr. Petra Heim

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Prof. Dr. Wolfgang Hölderich

Institut für Brennstoffchemie und physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Prof. Dr. Werner Hummel

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Hans Iding

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

XXII List of Contributors

Dr. Tomoyuki Inoue

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Prof. Dr. Karl-Erich Jaeger

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Sandra Johnen

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Bettina Juchem

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Doris Kolter-Jung

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Prof. Dr. Udo Kragl

Institut für Chemie Universität Rostock Albert-Einstein-Str. 3a 18059 Rostock Germany

Prof. em. Dr. Maria-Regina Kula

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Prof. Dr. Walter Leitner

Institut für Technische und Makromolekulare Chemie RWTH Aachen Worringerweg 1 52074 Aachen Germany

Prof. Dr. Andreas Liese

Institut für Technische Biokatalyse Technische Universität Hamburg-Harburg Denickestr. 15 21073 Hamburg Germany

Dr. Bettina Lingen

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Stephan Lütz

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Prof. Dr. Michael Müller

Lehrstuhl für Pharmazeutische und Medizinische Chemie Institut für Pharmazeutische Wissenschaften Albert-Ludwigs-Universität Freiburg Albertstr. 25 79104 Freiburg Germany

Adam Nitsche

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Priv. Doz. Dr. Martina Pohl

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Prof. Dr. Hermann Sahm

Institut für Biotechnologie I Forschungszentrum Jülich 52425 Jülich Germany

Prof. Dr. Albrecht Salzer

Institut für Anorganische Chemie RWTH Aachen Landoltweg 1 52074 Aachen Germany

Dr. Ulrich Schörken

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Dr. Thomas Schubert

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Dr. Martin Schürmann

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Dr. Melanie Schürmann

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Dr. Carmen Schuster

Institut für Brennstoffchemie und Physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

Dr. Petra Siegert

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Prof. Dr. Georg A. Sprenger

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Gerda Sprenger

Institut für Mikrobiologie Universität Stuttgart Allmandring 31 70550 Stuttgart Germany

Dr. Hans-Hermann Wagner

Institut für Brennstoffchemie und Physikalisch-chemische Verfahrenstechnik RWTH Aachen Worringerweg 1 52074 Aachen Germany

XXIV List of Contributors

Lydia Walter

Lehrstuhl für Pharmazeutische und Medizinische Chemie Institut für Pharmazeutische Wissenschaften Albert-Ludwigs-Universität Freiburg Albertstr. 25 79104 Freiburg Germany

Prof. Dr. Christian Wandrey

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

Dr. Andrea Weckbecker

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Marion Wendorff

Institut für Molekulare Enzymtechnologie der HHU Düsseldorf im FZ Jülich 52426 Jülich Germany

Dr. Michael Wolberg

Institut für Biotechnologie II Forschungszentrum Jülich 52425 Jülich Germany

1 Stoichiometric Asymmetric Synthesis

1.1 Development of Novel Enantioselective Synthetic Methods *Dieter Enders and Wolfgang Bettray*

1.1.1 Introduction

Since the pioneering times of the mid-1970s, when the first practical and generally applicable methods in asymmetric synthesis [1] were developed, such as the oxazoline method of Meyers [2] and the SAMP/RAMP hydrazone method [3], there has been a tremendous growth in this research field. One major driving force for this rapid development is of course the different biological activities of enantiomers and thus the need for enantiopure compounds. In this chapter we describe the development of some efficient synthetic methods for asymmetric carbon–carbon and carbon–heteroatom bond formation, which have been carried out within the frame of the "Sonderforschungsbereich 380" (1994–2005) and employing the concept of stoichiometric asymmetric synthesis.

1

1.1.2 α -Silyl Ketone-Controlled Asymmetric Syntheses

Electrophilic substitutions with carbon and hetero electrophiles α to the carbonyl group of aldehydes and ketones are among the most important synthetic operations. Such regio-, diastereo-, and enantioselective substitutions can be carried out efficiently with the SAMP/RAMP hydrazone methodology [3]. For cases where virtually complete asymmetric inductions could not be attained, an alternative approach based on α -silylated ketones **2** was developed [4]. They can be prepared easily from ketones **1** in high enantiomeric purity (*ee* > 98%) by asymmetric carbon silylation employing the SAMP/RAMP hydrazone method (Fig. 1.1.1). After the introduction of various electrophiles via classical enolate chemistry with excellent asymmetric inductions, the desired product ketones **3**



Fig. 1.1.1 α -Silyl-controlled asymmetric synthesis (the "silyl trick").



Fig. 1.1.2 NF-reagents for electrophilic fluorination.

are obtained by removal of the "traceless" silyl directing group with various sources of fluoride.

1.1.2.1 Regio- and Enantioselective α-Fluorination of Ketones

Due to the unique properties of organofluorine compounds and their rapidly increasing practical usage in plant protection, medicine, and many other areas, the scientific and economic interest in organofluorine coumpounds has grown immensely over recent decades. With the availability of user-friendly NF reagents such as 4 (NFSI, Accufluor[®]), 5 (NFOBS), 6 (Davis et al.) and 7 (Selectfluor[®]) (Fig. 1.1.2), for electrophilic fluorination [5], the efficient synthesis of α -fluorinated ketones, aldehydes, and esters has become possible. However, the asymmetric inductions in enantioselective α -fluorinations of ketones reached no practical values (*ee* = 10–75%) until the mid-1990s. We were therefore pleased to see that our α -silyl ketone-controlled approach led for the first time to the target α -fluoro ketones in high yields, few steps, and very good enantiomeric excesses [6].

As shown in Scheme 1.1.1, symmetric and unsymmetric ketones (control of regioselectivity) as well as cyclic and acyclic ketones **8** were first converted to the corresponding virtually enantiopure α -silyl ketones **2** (*ee* > 98%) employing the SAMP/RAMP hydrazone methodology. Metallation with LDA and treatment of the enolates with the *N*-fluorosulfonamide **4** (NFSI) afforded the α -fluoro- α '-silylated ketones **9** with moderate to excellent diastereomeric excesses. Finally, the racemization-free removal of the sterically demanding silyl directing group was carried out with fluoride sources in almost quantitative yields, leading to the desired α -fluoroketones **10** (*ee* 55 to >96%). Especially in the case of cyclic ketones almost complete asymmetric inductions could be achieved. As the epimeric



Scheme 1.1.1 Asymmetric synthesis of α -fluoroketones.

fluorinated silyl ketones **9** can be separated easily by flash column chromatography, various enantiopure α -fluoroketones **10** could be obtained in this way.

Although efficient organocatalytic methods for the electrophilic α -fluorination of aldehydes and ketones have recently been developed [7], high enantiomeric excesses can only be reached with aldehydes so far. The asymmetric inductions in the case of ketone fluorinations have remained low ($ee \leq 36\%$) [7a]. Thus, the α -silyl ketone-controlled stoichiometric asymmetric synthesis of α -fluoroketones **10** (Scheme 1.1.1) still constitutes a practical method.

1.1.2.2 α-Silyl Controlled Asymmetric Mannich Reactions

The Mannich reaction, in which an aminomethyl group is introduced in the α position of the carbonyl function, has been the subject of investigations since the early 20th century [8]. In 1985 our research group, in close cooperation with Steglich and coworkers, developed a first asymmetric Mannich reaction [9]. Some ten years later, with the enantiopure α -silylated ketones **2** in hand, we reported a first practical procedure for the regio- and enantioselective α -aminomethylation of ketones taking advantage of the excellent asymmetric inductions with the help of the "traceless" silyl control group [10].

As depicted in Scheme 1.1.2, the silyl ketones (*S*)-**11** of high enantiomeric purity were converted into the *Z*-configured silyl enol ethers (*S*)-**12**, which were used in the aminomethylation step by treatment with dibenzyl(methoxymethyl)amine in the presence of a Lewis acid. The silylated Mannich bases (*S*,*R*)-**13** were obtained in excellent yields and diastereomeric excesses (de = 92-96%). Finally,



Scheme 1.1.2 Enantioselective synthesis of β -dibenzylamino ketones.

the silvl directing group was removed tracelessly by employing a fluoride source. In this way, the α -substituted β -amino ketones (*R*)-14 were obtained in three steps with superb overall yields of 90–95% and, most importantly, with very high enantiomeric excesses *ee* of 91–97%. To explain the almost complete diastereofacial selectivity of the Mannich key step, two transition states can be discussed: a closed one along the lines of the Zimmerman–Traxler model, and an open one with the iminium ion formed in situ, explaining in both cases the *R* configuration at the newly generated stereocenter (Scheme 1.1.3).

After the successful asymmetric synthesis of α -substituted β -amino ketones (*R*)-14, we envisaged the diastereo- and enantioselective synthesis of α , β -disubstituted Mannich bases. As shown in Scheme 1.1.4, we were able to use benzaldehyde-*N*-phenylimine [11] as well as α -alkoxycarbonylaminosulfones [12] as Mannich electrophiles to synthesize in good overall yields and high *de*- and *ee*-values the *anti*-configured β -amino ketones (*R*,*S*)-15 and (*R*,*S*)-16, respectively [13].