Leukocyte Trafficking

Molecular Mechanisms, Therapeutic Targets, and Methods

Edited by Alf Hamann and Britta Engelhardt



WILEY-VCH Verlag GmbH & Co. KGaA

Leukocyte Trafficking

Edited by Alf Hamann and Britta Engelhardt

Related Titles

R. A. Meyers (Ed.) Encyclopedia of Molecular Cell Biology and Molecular Medicine, 2nd Edition

2005 ISBN 3-527-30542-4 http://meyers-emcbmm.de

D. Wedlich (Ed.) Cell Migration in Development and Disease 2005 ISBN 3-527-30587-4

A. Ridley, M. Peckham, P. Clark (Eds.) Cell Motility: From Molecules to Organisms 2004 ISBN 0-470-84872-3

A. Steinkasserer, N. Romani, M. Lutz (Eds.) Handbook of Dendritic Cells – Biology, Diseases and Therapies 2005 ISBN 3-527-31109-2

S. H. E. Kaufmann (Ed.) Novel Vaccination Strategies

ISBN 3-527-30523-8

H. Kropshofer, A. Vogt (Eds.) Antigen Processing Cells – From Mechanisms to Drug Development 2005 ISBN 3-527-31108-4

A. Meager (Ed.) **The Interferons – Characterization and Application** 2005 ISBN 3-527-31180-7

K. M. Pollard (Ed.) Autoantibodies and Autoimmunity – From Mechanisms to Treatments 2005

ISBN 3-527-31141-6

Leukocyte Trafficking

Molecular Mechanisms, Therapeutic Targets, and Methods

Edited by Alf Hamann and Britta Engelhardt



WILEY-VCH Verlag GmbH & Co. KGaA

Editors

Prof. Dr. Alf Hamann

Charité University Medicine Berlin Experimental Rheumatology Medical Clinic for Rheumatology and Clinical Immunology c/o German Rheumatism Research Center Schumannstr. 21/22 10117 Berlin Germany

Prof. Dr. Britta Engelhardt

Immunobiology Theodor Kocher Institute University of Bern Freiestrasse 1 3012 Bern Switzerland All books published by Wiley-VCH are carefully produced. Nevertheless, authors, editors, and publisher do not warrant the information contained in these books, including this book, to be free of errors. Readers are advised to keep in mind that statements, data, illustrations, procedural details or other items may inadvertently be inaccurate.

Library of Congress Card No.: Applied for British Library Cataloging-in-Publication Data: A catalogue record for this book is available from the British Library.

Bibliographic information published by Die Deutsche Bibliothek

Die Deutsche Bibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data is available in the Internet at http://dnb.de.

© 2005 WILEY-VCH Verlag GmbH & Co. KGaA, Weinheim

All rights reserved (including those of translation in other languages). No part of this book may be reproduced in any form – nor transmitted or translated into machine language without written permission from the publishers. Registered names, trademarks, etc. used in this book, even when not specifically marked as such, are not to be considered unprotected by law.

Printed in the Federal Republic of Germany. Printed on acid-free paper.

Composition Asco Typesetters, Hong Kong Printing betz-druck GmbH, Darmstadt Bookbinding J. Schäffer GmbH i.G., Grünstadt

ISBN-13 978-3-527-31228-3 ISBN-10 3-527-31228-5

Contents

Preface XV

List of Authors XIX

Movies on the included CD XXIII

Color Plates XXV

- Part I Molecular Mechanisms 1
- 1
 The Multistep Model of Leukocyte Trafficking: A Personal Perspective from

 15 Years Later
 3

 Eugene C. Butcher
 3

 Acknowledgments
 9

 References
 9
- 2 Capture and Rolling: Selectins and Their Ligands 14 Claudine S. Bonder and Paul Kubes
- 2.1 Introduction 14
- 2.2 Selectins 15
- 2.2.1 L-Selectin 15
- 2.2.2 P-Selectin 18
- 2.2.3 E-Selectin 19
- 2.3 P-Selectin Glycoprotein Ligand 1 and Other Ligands of Selectins 20
- 2.4 Glycosyltransferases 24 References 27
- 3 Chemokines and Their Receptors: Biochemical, Structural and Biological Properties 36 Martin Oppermann and Reinhold Förster
- 3.1 Introduction 36
- 3.2 Chemokines 38
- 3.3 Chemokine Receptors 43

VI Contents

- 3.4 Role of Chemokines in Lymphocyte and Dendritic Cell Trafficking to and Within Primary and Secondary Lymphoid Organs 48
- 3.4.1 Primary Lymphoid Organs 48
- 3.4.1.1 Bone Marrow 48
- 3.4.1.2 Thymus 50
- 3.4.2 Secondary Lymphoid Organs 51
- 3.4.2.1 Spleen 52
- 3.4.2.2 Lymph Nodes 53
- 3.4.2.3 Mucosa-Associated Lymphoid Tissue 54 Acknowledgments 55 References 56

4 Mechanisms of Leukocyte Integrin Activation 68

Gabriela Constantin and Carlo Laudanna

- 4.1 Introduction 68
- 4.2 Modalities of Integrin Activation and the Role of Chemokines 69
- 4.3 Signaling Mechanisms Controlling Rapid Integrin Activation 71
- 4.4 Chemokines, Integrins and Concurrency in Leukocyte Recruitment 76
- 4.5 The Way Ahead 77 References 77
- 5 Mechanisms of Leukocyte Transmigration: Role of Immunoglobulin Superfamily Molecules 82

Federica M. Marelli-Berg and Sussan Nourshargh

- 5.1 Introduction 82
- 5.2 Leukocyte Migration Through Endothelial Cells 83
- 5.3 Endothelial Cell Junctional Molecules 83
- 5.4 Role of Immunoglobulin Superfamily Cell Adhesion Molecules in Leukocyte Transmigration 87
- 5.5 Intercellular Adhesion Molecules 87
- 5.5.1 Structure, Ligands, and Expression Profile 87
- 5.5.2 Role in Leukocyte Transmigration 88
- 5.5.3 Signaling by ICAM-1 and ICAM-2 89
- 5.6 Junctional Adhesion Molecules 90
- 5.6.1 Structure, Ligands, and Expression Profile 90
- 5.6.2 Role in Leukocyte Transmigration 91
- 5.6.3 Signaling by JAMs 92
- 5.7 PECAM-1 (CD31) 92
- 5.7.1 Structure, Ligands, and Expression Profile 92
- 5.7.2 Role in Leukocyte Transmigration 93
- 5.7.3 Signaling by PECAM-1 94
- 5.8 Role of Additional Molecules in Regulation of Leukocyte Transmigration 95
- 5.8.1 T Cell Receptor 95

- 5.8.2 CD99 97
- 5.9 Summary and Future Directions 98
 Acknowledgments 99
 References 99

6 The Endothelial Cell Basement Membrane and Its Role in Leukocyte Extravasation 109

- Lydia M. Sorokin
- 6.1 Introduction 109
- 6.2 Extracellular Matrix of Blood Vessels 110
- 6.2.1 Basement Membranes 110
- 6.2.2 Laminins 112
- 6.3 Function of Endothelial Cell Basement Membranes 116
- 6.3.1 Leukocyte Adhesion and Migration Studies 116
- 6.3.2 Methods of Investigation of Leukocyte Migration on Extracellular Matrix Substrates *118*
- 6.3.3 Murine Inflammatory Models 118
- 6.3.4 Role of Proteases 121
- 6.4 Conclusion 122 Acknowledgment 122 References 122

Part II Trafficking in vivo 129

- 7 Control of Homing Receptor Expression during Lymphocyte Differentiation, Activation, and Function 131 Daniel J. Campbell
- 7.1 Introduction 131
- 7.2 Developing Lymphocytes Undergo Programmed Changes in Homing Receptor Expression 132
- 7.3 Control of Homing Receptor Expression During Lymphocyte Activation and Effector Cell Differentiation 134
- 7.3.1 Cytokine Control of Homing Receptor Expression by Th1 and Th2 Cells 135
- 7.3.2 Function of Dendritic Cells in Directing T Cell Homing Receptor Expression 136
- 7.3.3 Generation of Central and Effector Memory Populations 139
- 7.4 Homing Receptor Expression by Effector/Memory Lymphocytes: Lineage or Lifestyle? 140
- 7.5 Selection vs. Instruction 140
- 7.6 Transcriptional Control of Homing Receptor Expression 141
- 7.7 Concluding Remarks 145 Acknowledgments 145 References 145

VIII Contents

Alf Hamann, Carrie N. Arnold, and Gudrun F. Debes8.1Major Lymphocyte Lineages8.1.1NK and NKT Cells and γδ T Cells1548.1.2Naïve T and B Cells1548.2Impact of Activation on Trafficking of T Cells155
8.1Major Lymphocyte Lineages 154 8.1.1NK and NKT Cells and $\gamma\delta$ T Cells 154 8.1.2Naïve T and B Cells 154 8.2Impact of Activation on Trafficking of T Cells 155
 8.1.1 NK and NKT Cells and γδ T Cells 154 8.1.2 Naïve T and B Cells 154 8.2 Impact of Activation on Trafficking of T Cells 155
8.1.2 Naïve T and B Cells 1548.2 Impact of Activation on Trafficking of T Cells 155
8.2 Impact of Activation on Trafficking of T Cells 155
± Ø
8.3 Trafficking of Effector/Memory T Cells 157
8.4 Specialized Effector/Memory T Cell Subsets Defined by their Expression
of Chemokine Receptors 159
8.4.1 CCR7 and Effector/Memory T Cells 159
8.4.2 Follicular Homing and Germinal Center CD4 ⁺ T Cells 161
8.5 Differential Trafficking of Functional Subsets: Th1, Th2, and Regulatory
T Cells 163
8.5.1 Th1 and Th2 Cells 163
8.5.2 Regulatory T Cells 164
8.6 Summary 165
Acknowledgments 166
References 166
9 Trafficking of B Cells 173
Rudolf A. Manz
9.1 Introduction 173
9.2 B ₁ Cells 173
9.3 B Cell Precursors and Immature B Cells 174
9.4 Peripheral B Cells 176
9.5 Germinal Center B Cells and Memory B Cells 178
9.6 Plasma Cells 179
References 182
10 Trafficking of Dendritic Cells 184
Nikolaus Romani, Sandra Holzmann, Christoph H. Tripp, Michael Sixt, and
Patrizia Stoitzner
10.1 Introduction 184
10.1.1 Dendritic Cells 184
10.1.2 Langerhans Cells: The Prototype of Trafficking Dendritic Cells 184
10.2 Pathways and Morphology of Dendritic Cell Trafficking 185
10.2.1 Life Path of a Dendritic Cell from Birth to Death 185
10.2.2 Trafficking from the Bone Marrow to the Tissue of Residence 185
10.2.3 Egress from the Tissue of Residence, Migration Through Connective
Tissue, and Entry into Lymph Vessels 186
10.2.4 Arrival at the Lymphatic Organs and Entry into the T Cell Area 189
10.2.5 Dendritic Cell Traffic "In the Fast Lane" 191
10.3 Regulation of Dendritic Cell Trafficking 191
10.3.1 Trafficking from the Bone Marrow to the Tissue of Residence 191
10.3.2 Egress from the Tissue of Residence 193
10.3.3 Relationship Between Migration and Maturation of Dendritic Cells 199

Contents IX

- 10.3.4 Trafficking of Plasmacytoid Dendritic Cells 200
- 10.4 Functional Implications of Dendritic Cell Trafficking 200
- 10.4.1 Homeostasis of the Sentinel Cell Network 200
- 10.4.2 Initiation of Immunity 200
- 10.4.3 Maintenance of Peripheral Tolerance 201
- 10.4.4 Application-Oriented Considerations for Immunotherapy 202 Acknowledgments 203 References 203
- Part III Inflammation 217
- 11 Molecular and Cellular Contributions to Selectin-Dependent Leukocyte Adhesion Under Flow 219 Rodger P. McEver
- 11.1 Introduction 219
- 11.2 Structure of Selectins 219
- 11.3 Regulation of Expression of Selectins 221
- 11.4 Selectin Ligands 225
- 11.5 Regulation of Cell Rolling Under Flow 229
- 11.6 Signaling Through Selectins or Selectin Ligands 235 References 236
- 12Mechanisms of Inflammation: Neutrophils248

Markus Sperandio and Barbara Walzog

- 12.1 Formation and Differentiation of Neutrophils 248
- 12.2 Regulation of Neutrophil Homeostasis in the Circulation 250
- 12.3 Neutrophil Activation and Recruitment 251
- 12.4 Phagocytosis by Neutrophils 256
- 12.5 Neutrophil Apoptosis 258
- 12.6 Resolution of Inflammation 260 Movies Chapter 12 262 References 262

13 Chemokines Drive Inflammatory Leukocyte Recruitment 279

Stefan Floess and Antal Rot

- 13.1 Introduction 269
- 13.2 Inflammatory and Homeostatic Chemokines 269
- 13.3 Chemokine "Redundancy" 270
- 13.4 Chemokine Presentation by GAGs 271
- 13.5 Chemokine Interceptors 272
- 13.6 Chemokines on Blood–Tissue Interface 273
- 13.7 In Vivo Veritas 273
- 13.7.1 Association Studies 274
- 13.7.2 Experimental Administration of Chemokine/Receptor-Specific Therapeutic Substances 275

X Contents

13.7.3	Study of Animal Chemokine/Receptor Knockouts in Experimental
	Disease Models 275
13.8	CCR1 275
13.9	CCR2 277
13.10	CCR3 279
13.11	CCR4 279
13.12	CCR5 280
13.13	CCR6 281
13.14	CCR8 281
13.15	CCR9 282
13.16	CXCR2 282
13.17	CXCR3 283
13.18	CXCR6 284
13.19	CX ₃ CR1 284
13.20	Concluding Remarks 285
	Acknowledgment 285
	References 285
14	Mechanism of Inflammation: Activation of the Endothelium 300
	Matthias Clauss and Carolyn F. Patterson
14.1	Introduction 300
14.2	Effects of Endothelial Activation 300
14.2.1	Adhesion Molecules 300
14.2.2	Chemokines 302
14.2.3	Hemostasis 303
14.2.4	Vascular Permeability 303
14.2.5	Other Effects 304
14.3	Cell Activating Factors and Principles 304
14.3.1	Infection and Bacterial Products 304
14.3.2	Cytokines 305
14.3.3	Other Bioactive Proteins and Peptides 307
14.3.4	Bioactive Lipids 307
14.3.5	Mechanical Forces 308
14.3.6	Leukocyte-Endothelial Binding 308
14.4	Signaling of Endothelial Activation 309
14.4.1	Acute Stimulation and the MAPK Cascade 309
14.4.2	NF- κ B and AP-1 Families of Transcription Factors 312
14.5	Role of Reactive Oxygen Species in Endothelial Activation 313
14.5.1	Oxidants 313
14.5.2	Redox Signaling 314
14.5.3	ADPH Oxidase 314
14.6	Chronic Endothelial Cell Activation 315
14.6.1	Examples of Continuous Local Inflammation 315
14.6.2	tmTNF Transgenic Mice as a Model of Chronic Inflammation 317
	References 318

Part IV	Trafficking	Mechanisms as	s Therapeutic	Targets	337
---------	-------------	---------------	---------------	---------	-----

- **15** Integrins as Therapeutic Targets for Inflammatory Disease 339 Michael J. Briskin
- 15.1 Introduction 339
- 15.2 Preclinical Studies 340
- 15.2.1 α₄ Integrins 340
- 15.2.2 CD11a Preclinical Studies 342
- 15.3 Humanization of Anti-integrin mAbs 344
- 15.4 Clinical Trials with Humanized Anti-α₄ Integrin mAbs 345
- 15.4.1 Studies with Natalizumab in MS 345
- 15.4.2 Trials in Inflammatory Bowel Disease: Humanized Anti- α_4 and Anti- $\alpha_4\beta_7$ Integrins 348
- 15.4.3 LFA-1 as a Target for Treatment of Psoriasis 352
- 15.5 Clinical Studies of Humanized Anti-CD11a 352
- 15.5.1 Small Molecule Integrin Antagonists 357
- 15.6 Other Approaches: Antisense Antagonists 361
- 15.7 Final Remarks 361 Acknowledgments 362 References 363

16 Chemokine Receptor Antagonists: From the Bench to the Clinic 371 Sofia Ribeiro and Richard Horuk

- 16.1 Introduction 371
- 16.2 Chemokines and Their Receptors 372
- 16.3 CC Chemokine Receptor Antagonists 372
- 16.3.1 CCR1 Antagonists 372
- 16.3.2 CCR2 Antagonists 376
- 16.3.3 CCR3 Antagonists 378
- 16.3.4 CCR4 Antagonists 381
- 16.3.5 CCR5 Antagonists 381
- 16.3.6 Other Members of the CC Family 385
- 16.4 CXC Chemokine Receptor Antagonists 387
- 16.4.1 CXCR1/CXCR2 Antagonists 387
- 16.4.2 CXCR3 Antagonists 389
- 16.4.3 CXCR4 Antagonists 391
- 16.5 Conclusion 393
 - Acknowledgments 393
 - References 394

XII Contents

Part V Methods to Study Cell Trafficking 403

17	Leukocyte-Endothelial Cell-Cell Interactions in Vitro: Static Assays and
	Adhesion under Shear Stress 405
	Markus Hammel, Olaf Zilles, and Rupert Hallmann
	with a contribution from Silke Jennrich, Kerstin Siegmund, and Alf Hamann
17.1	Introduction 405
17.2	Adhesion of Leukocytes to High Endothelial Venules: The HEV
	Assay 405
17.2.1	Materials 406
17.2.2	Procedure 406
17.2.2.1	Preparation of Frozen Lymphatic Tissue Blocks 406
17.2.2.2	HEV Assay 406
17.2.3	Expected Results 407
17.2.4	Troubleshooting 407
	References 408
17.3	Adhesion of Leukocytes to Cultured Endothelium, With or Without
	Shear 408
17.3.1	Materials 409
17.3.2	Procedure 410
17.3.3	Expected Results 411
17.3.4	Troubleshooting 412
17.3.5	Additional Comments and Hints 413
	Bibliography 413
	Relevant Webpages 414
17.4	Adhesion Assay under Static Conditions in Microtiter Plates 414
17.4.1	Materials 414
17.4.2	Procedure 415
17.4.2.1	Coating of the 96-Well Plate with Adhesion Molecules 415
17.4.2.2	Cell Preparation and CFDA-SE Labeling 415
17.4.2.3	Adhesion Assay 416
17.4.3	Additional Comments 416
	Reference 417
18	Chemotaxis Assay: Analysis of Migration of Lymphocyte Subsets 418
	Kerstin Siegmund, Gudrun F. Debes, and Alf Hamann
18.1	Introduction 418
18.2	Basic Protocol 419
18.2.1	Cell Isolation 419
18.2.2	Cell Preparation 419
18.2.3	Chemotaxis Assay 420
18.2.4	Quantification of Migrated Cells by Flow Cytometry 420
18.2.5	Analysis of Migration Rates of Minor Lymphocyte Subsets 422
	References 422

19	In Vitro Transendothelial Migration Assay 424
	Ruth Lyck and Britta Engelhardt
19.1	Introduction 424
19.2	Methods for the Investigation of Transendothelial Migration in Vitro 425
19.3	Experimental Details 428
19.4	Protocol 431
19.4.1	Equipment and Reagents 432
19.4.2	Basic Protocol 432
	References 434
20	Real Time in Vitro Assays for Studying Leukocyte Transendothelial Migration
	Under Physiological Flow Conditions 437
	Ronen Alon, Guy Cinamon, and Francis W. Luscinskas
20.1	Background 437
20.2	Introduction 437
20.3	Disadvantages of Analysis of Leukocyte TEM Without Provision for Shear Flow 438
20.4	An Alternative System for Real-Time Analysis of Leukocyte TEM 439
20.5	Spatial and Temporal Analysis of Leukocyte Adhesion Molecules and
	Endothelial Junctional Molecules During Leukocyte TEM 442
20.6	Drawbacks of Present Flow Chamber Technologies 444
20.7	Open Questions and Extended Tools for Studying Leukocyte TEM
	In Vitro 446
	Appendix: Experimental Procedures for a Standard Flow Chamber-Based
	TEM Analysis 447
	Materials 447
	Methods 448
	Notes 449
	Movies Chapter 20 450
	References 451
21	Intravital Microscopy and In Vitro Flow Chamber: Techniques to Study
	Leukocyte Adhesion Under Flow and in Real Time 455
	Jens V. Stein
21.1	Introduction 455
21.2	Hemodynamics 456
21.3	In Vitro Flow Chamber 459
21.4	Advantages and Limitations of FCAs 460
21.5	IVM: Past and Present 461
21.6	Practical IVM 461
21.7	Observing Endogenous Versus Exogenous Cells 464
21.8	Advantages and Limitations of IVM 465
21.9	Emerging Applications for FCA and IVM 466
21.10	Conclusion 467
	Acknowledgements 46/
	Movies Chapter 21 467
	References 468

22	Immune Processes in the Light of Two-Photon Microscopy 472
	Alexander Flügel and Naoto Kawakami
22.1	Introduction 472
22.2	Two-Photon Live Microscopy: Basic Principles 473
22.2.1	Confocal Versus Two-Photon Microscopy 473
22.2.2	Pros and Cons of TPM 476
22.2.3	Components of a Two-Photon Setting 477
22.2.4	Two-Photon Markers 478
22.2.5	Live Imaging Setting 479
22.3	Two-Photon Analyses of Immune Processes 480
22.3.1	T Cell Development in Thymic Aggregate Cultures 481
22.3.2	Immune Cell Motility and Antigen Encounter in Lymph Nodes 484
22.3.3	The Effector Phase of Autoreactive CD4 ⁺ T Cells 485
22.4	Conclusions 486
	Acknowledgments 487
	References 488
23	Use of Labeled Lymphocytes to Analyze Trafficking In Vivo 497
	Kerstin Siegmund and Alf Hamann
23.1	Introduction 497
23.1.1	Use of v-Emitting Isotopes for Lymphocyte Migration Studies 497
23.1.2	Use of Fluorescent Dyes for Lymphocyte Migration Studies 498
23.1.3	Use of Genetic Markers for Lymphocyte Migration Studies 499
23.2	Protocols for Labeling with Radioisotopes 501
23.2.1	Materials 501
23.2.2	Procedure for Labeling with Sodium [⁵¹ Cr]Chromate 501
23.2.3	Modifications of the Above Protocol: Labeling with [¹²⁵ I]Iodine
2324	Further Comments 502
23.2.1	Safety Considerations 503
23.2.5	Homing Procedure: Injection of Labeled Cells and Determination of
23.5	Radioactivity Recovered 503
23.3.1	Materials 503
23.3.2	Procedure: Cell Injection, Organ Removal, Counting, and Data
	Analyses 503
23.4	Use of Antibodies Against Adhesion Molecules in Homing
	Experiments 505
23.5	Protocols for Labeling with Fluorescent Dyes 506
23.5.1	Fluorescent Labels Used for Cell Tracking 506
23.5.2	Procedure: Labeling with Carboxyfluorescein Diacetate Succinimidyl
	Ester 506
23.5.3	Safety Considerations 507
23.5.4	General Comments on Labeling Procedures 507
	References 508

Index 509

Preface

A Mobile Society – The Constitutive Role of Cell Trafficking in the Organization of the Immune System

Among other evolutionary achievements of the vertebrates, the immune system stands out not only for its complexity, but also for its unique construction as a system composed of highly cooperative individual cells as basic elements, which are mobile and distributed all over the body. The various subpopulations of leukocytes resemble the members of a complex society, with functional specializations, numerous interactions, order – and chaos. It is a consistently fascinating fact that this society of migrating leukocytes allows the execution of functions as diverse as balancing self-tolerance against defense, systemic response to antigens with generation of effector mechanisms and specific memory, and tailored, local immune reactions. Molecular mechanisms allowing the targeted migration of cell types into distinct compartments are a central organizational feature of this system because they allow the precise topographical and temporal delivery of leukocyte subpopulations.

Directed migration of cellular elements of the immune system begins after development within the primary compartment, predominantly the bone marrow. Some cell types emigrate as precursors and complete their differentiation within another compartment, e.g., pre-T cells, which migrate into the thymus where a highly specialized environment provides the appropriate conditions for their maturation and selection. Also linked to specific differentiation phases is the changing location of B-cells after their maturation within the bone marrow (or, in birds, the bursa of Fabricius), from where they migrate into the spleen or the lymph nodes and, after antigen encounter and differentiation into plasma blasts, back into the bone marrow, into the lamina propria, or into some inflamed tissues (see Chapter 9).

Unidirectional migration is typical for cells of the innate immune system such as neutrophils. Generated within the bone marrow, they stay for a short time (less than a day) idle within the blood stream without extravasating into any tissue until they undergo apoptosis and are cleared away by the liver. Eventually, inflammation recruits them into a peripheral tissue site and calls up their effector mechanisms, as discussed in this book (Chapter 12).

Since the pioneering work of Sir James L. Gowans [1] we have known that

XVI Preface

lymphocytes behave differently: naïve lymphocytes, both T cells and B cells, recirculate continuously between blood and lymphoid tissues. This process allows them to percolate for most of their time through lymphoid tissues all over the body, where a few dendritic cells – the critical antigen-presenting cell fraction involved in T cell priming – might have arrived after taking up antigen in the peripheral sites (see Chapter 10). It may be imagined that this continuous cycling greatly enhances the chances that lymphocytes of a given antigen specificity will meet those dendritic cells that present the cognate antigen.

Naïve lymphocytes do not enter peripheral tissues, including inflamed sites; these compartments are reserved for the more mature members of the society, the memory and effector lymphocytes. Thus, the places where an immune response is induced – the lymphoid tissues – and the places where the actual defense reactions take place – the nonlymphoid tissues exposed to microbial attacks – are strictly separated. It only can be speculated that the evolutionary benefit of this organization is to keep cells concentrated in a few relevant places, and to provide separate environments for either priming and maturation of naïve lymphocytes or for activation and execution of effector functions.

Lymph node high endothelial venules provide an armamentarium of traffic signals for the rapid recruitment of naïve lymphocytes from the blood stream. Investigation of lymphocyte traffic across high endothelial venules in lymph nodes and Peyer's patches helped to define the multi-step cascade of lymphocyte–endothelial interaction as a universal scheme for how leukocytes in general can extravasate from the blood stream into any tissue (see Chapter 1). According to this model, adhesion molecules from different families – the selectin family (Chapter 2), the integrin family (Chapter 4), and the Ig supergene family (Chapter 5) – act in synergy with chemokines (Chapter 3) to govern the finely regulated process of leukocyte tethering, adhesion, and transmigration. Within the tissue, molecules of the extracellular matrix modulate the final step of leukocyte migration and entry into the target tissue (Chapter 6), whereas distinct chemokines produced by their respective cell populations help to facilitate the exact positioning of interacting partners.

In addition to recirculation, Gowans and others also detected 40 years ago the capacity of some lymphocyte populations to return selectively to the tissues from which they were isolated ("homing" in the narrower sense). It later became clear that this topographical memory is a property of activated or memory/effector cells. Memory of the site of priming is induced upon encounter with antigen and subsequent differentiation into effector/memory cells. The current dogma assumes that tissue-specific factors, most likely produced by local dendritic cells, shape the differentiation so as to generate lymphocytes that home back to the tissue of initial antigen encounter (see Chapter 7). The crucial question of how the expression of organ-specific homing receptors is regulated, what factors are involved, and whether permanent imprinting occurs, is a matter of recent research.

Tissue-specific homing has been interpreted as a means of focusing memory cells on the sites of initial priming, where the likelihood of later recurrence of the same infectious agent is highest. However, even after 40 years of research, it is unclear whether this concept applies to more compartments than the gastrointestinal tract together with its associated lymphoid structures, with perhaps the skin as another defined target. Lymphocytes are recruited to a variety of other tissues by more or less universal sets of receptors, or by inflammation-related migration pathways.

The mobile cell of the immune system needs anchor points to perceive the topographical information required for tailored, site-specific activities. Two cell populations have well-described functions in this context. First, the dendritic cell provides signals that shape the quality and direction of an immune response, in addition to presenting antigen. Although dendritic cells originate from bone marrow and arrive only in their final phase from different tissue in lymphoid organs, they seem to acquire a local flavor upon settling, with an ability to shape the homing behavior of T-cells upon interaction. Secondly, endothelial cells act as gatekeepers and selective catchers, guiding distinct cell populations into various compartments of the body. They provide the range of traffic signs recognized by the mobile leukocyte within the blood stream. In addition, the great variability in the composition of extracellular matrix and the multitude of chemokines produced by resident tissue cells or cells of the immune system constitute further road signs to guide traveling cells.

While the tissue-related determinants allow the assignment of specific subpopulations to site-specific duties, inflammation is a condition where adhesion molecules and chemokines become transiently upregulated and for a certain time window define a new target for leukocyte – especially effector cell – trafficking. Indeed, rapid infiltration of leukocytes into the affected tissue is a classical hallmark of inflammation. This feature allows the immune system to store large numbers of reactive cells in the circulation or in central depots such as spleen or lymph nodes from where they can be rapidly mobilized and redirected to the sites of immune reactions taking place at any place of the body.

A few adhesion molecules, notably the endothelial selectins (Chapter 11) and a series of chemokines and their receptors (Chapter 13), are exclusively induced under inflammatory conditions; other receptors might be upregulated in their expression levels. The functioning of endothelium is crucial in this process; its activation (Chapter 14) by inflammatory mediators such as cytokines produced within the tissue provides the key signals that attract different populations of leukocytes or lymphocyte differentiation stages out of the circulation into the inflamed area.

The inflammation-triggered accumulation of leukocytes at the site of an immune reaction represents a powerful adaptive response of the system. Not only does it allow the quantitative number of effector cells to be rapidly increased, but it also enables the character of the response to be shaped qualitatively. Increasing data suggest that the large number of toll-like and other innate receptors for pathogen-associated determinants affects the specific pattern of chemokines, cytokines, and adhesion molecules induced depending on the nature of the pathogen, so that customized reactions result.

Rapid inflammation-triggered recruitment into affected tissue sites is observed for cells of the innate immune system such as neutrophils (Chapter 12) in the same way as for the memory/effector stage of lymphocytes (Chapter 8), suggesting that trafficking and transmigration mechanisms evolved early in evolution and differentiated to allow finely tuned regulation of leukocyte positioning.

The relevance of these mechanisms for understanding the pathophysiology of acute and chronic inflammatory diseases and for the development of novel therapeutic options is obvious; accordingly, the field has attracted much attention in the last years. Indeed, some approaches to target either adhesion molecules (Chapter 15) or, to a lesser extent, chemokines (Chapter 16) for an anti-inflammatory treatment have yield promising results already being tested in initial clinical trials.

The chapters in this book, written by experts in their respective fields, cover many important aspects of the basic mechanisms, molecular pathways, cellular features, and possible therapeutic modulation of leukocyte trafficking mentioned above. Such a book can never be either complete or as up to date as a journal article. We hope, nevertheless, that it will help the reader to understand the central features of migration of cellular elements of the immune system. We hope that it will both serve as an introduction to novices in the field and provide the experienced researcher with some new insights that will complement his or her own work.

Last but not least, the series of chapters describing the armamentarium of available methods for studying leukocyte trafficking (Chapters 17–23) aims to help an understanding of how major findings were achieved in the field and to advise readers in the design of their own experiments or laboratory classes for immunology students.

Acknowledgements

Experimental work by A.H. was supported by the German Research Foundation (DFG; SFB366, SFB421, and SFB633); work by B.E. was supported by the Max Planck Society, the DFG (individual grants and program grants SFB293, SFB297, and SFB629), Astra Zeneca, Sweden, and GlaxoSmithKline, UK.

We thank the members of our research groups for their past and present scientific contributions to our research and their valuable help in editing this book.

Reference

GOWANS, J.L., and E.J. KNIGHT. 1964. The route of recirculation of lymphocytes in the rat. *Proc. Roy. Soc. Lond. B* 159:257–282.

List of Authors

Ronen Alon

The Weizmann Institute of Science Department of Immunology Rehovot 76100 Israel

Carrie N. Arnold

Stanford University School of Medicine and Veterans Affairs–Palo Alto Health Care System Department of Microbiology & Immunology 3801 Miranda Avenue Palo Alto, CA 94304 USA

Claudine S. Bonder

University of Calgary Immunology Research Group Department of Physiology and Biophysics Institute of Infection, Immunity and Inflammation 3330 Hospital Drive N.W. Calgary, Alberta, T2N 4N1 Canada

Michael J. Briskin

Director of Immunology Merrimack Pharmaceuticals, Inc. 101 Binney St. Cambridge, MA 02142 USA

Eugene C. Butcher

Stanford University School of Medicine Laboratory of Immunology and Vascular Biology Department of Pathology, L235 Stanford, CA 94305-5324 USA

Daniel J. Campbell

Benaroya Research Institute 1201 9th Ave. Seattle, WA 98101 USA

Guy Cinamon

University of California San Francisco Howard Hughes Medical Institute Department of Microbiology San Francisco, CA 94143–0414 USA

Matthias Clauss

Indiana University School of Medicine Cellular and Integrative Physiology 975 W. Walnut Str. IB 433 Indianapolis, IN 46202 USA

Gabriela Constantin

University of Verona Department of Pathology Division of General Pathology Strada Le Grazie 8 37134 Verona Italy

Gudrun F. Debes

Stanford University School of Medicine and Veterans Affairs–Palo Alto Health Care System Department of Pathology 3801 Miranda Avenue Palo Alto, CA 94304 USA

Britta Engelhardt

University of Bern Theodor Kocher Institute Freiestrasse 1 3012 Bern Switzerland

XX List of Authors

Stefan Floess

Charité University Medicine Berlin Experimental Rheumatology Medical Clinic for Rheumatology and Clinical Immunology c/o German Rheumatism Research Center Schumannstr. 21/22 10117 Berlin Germany

Alexander Flügel

Max Planck Institute of Neurobiology Department of Neuroimmunology Am Klopferspitz 18 82152 Martinsried Germany

Reinhold Förster

Hanover Medical School Institute of Immunology Feodor-Lynen-Str. 21 30625 Hannover Germany

Rupert Hallmann

University of Münster Institute for Physiological Chemistry and Pathobiochemistry Waldeyerstr. 15 48149 Münster Germany

Alf Hamann

Charité University Medicine Berlin Experimental Rheumatology Medical Clinic for Rheumatology and Clinical Immunology c/o German Rheumatism Research Center Schumannstr. 21/22 10117 Berlin Germany

Markus Hammel

Lund University Department of Experimental Pathology Soelvegatan 25 22362 Lund Sweden

Sandra Holzmann

Innsbruck Medical University Department of Dermatology and Venereology Anichstrasse 35 6020 Innsbruck Austria

Richard Horuk

Berlex Biosciences Department of Immunology 2600 Hilltop Drive Richmond, CA 94806 USA

Silke Jennrich

Charité University Medicine Berlin Experimental Rheumatology Medical Clinic for Rheumatology and Clinical Immunology c/o German Rheumatism Research Center Schumannstr. 21/22 10117 Berlin Germany

Naoto Kawakami

Max Planck Institute of Neurobiology Department of Neuroimmunology Am Klopferspitz 18 82152 Martinsried Germany

Paul Kubes

University of Calgary Immunology Research Group Department of Physiology and Biophysics Institute of Infection, Immunity and Inflammation 3330 Hospital Drive N.W. Calgary, Alberta, T2N 4N1 Canada

Carlo Laudanna

University of Verona Department of Pathology Division of General Pathology Strada Le Grazie 8 37134 Verona Italy

F.W. Luscinskas

Harvard Medical School Brigham and Women's Hospital Department of Pathology 77 Ave Louis Pasteur Boston, MA 02115 USA

Ruth Lyck

University of Bern Theodor Kocher Institute Freiestrasse 1 3012 Bern Switzerland

Rudolf A. Manz

German Rheumatism Research Center DRFZ Schumannstrasse 21/22 10117 Berlin Germany

Federica Marelli-Berg

Imperial College London Faculty of Medicine Hammersmith Hospital Campus Du Cane Road London W12 ONN UK

Rodger P. McEver

Cardiovascular Biology Research Program Oklahoma Medical Research Foundation 825 N.E. 13th Street Oklahoma City, OK 73104 USA

Sussan Nourshargh

Imperial College London Faculty of Medicine Hammersmith Hospital Campus Du Cane Road London W12 ONN UK

Martin Oppermann

Georg-August-University Göttingen Department of Immunology Kreuzbergring 57 37075 Göttingen Germany

Carolyn E. Patterson

Indiana University School of Medicine Roudebush VA Medical Center 1481 W. 10th St. VA111 P Indianapolis, IN 46202 USA

Sofia Ribeiro

Berlex Biosciences Department of Immunology 2600 Hilltop Drive Richmond, CA 94806 USA

Nikolaus Romani

Innsbruck Medical University Department of Dermatology and Venereology Anichstrasse 35 6020 Innsbruck Austria

Antal Rot

Novartis Institutes for Biomedical Research Brunner Str. 59 1235 Vienna Austria

Kerstin Siegmund

Charité University Medicine Berlin Experimental Rheumatology Medical Clinic for Rheumatology and Clinical Immunology c/o German Rheumatism Research Center Schumannstr. 21/22 10117 Berlin Germany

Michael Sixt

Max-Planck-Institute for Biochemistry Department of Molecular Medicine Am Klopferspitz 18 82152 Martinsried Germany

Lydia M. Sorokin

University of Münster Institute for Physiological Chemistry and Pathobiochemistry Waldeyerstr. 15 48149 Münster Germany

Markus Sperandio

University of Heidelberg Children's Hospital, Neonatal Unit Im Neuenheimer Feld 150 69120 Heidelberg Germany

Jens V. Stein

University of Bern Theodor Kocher Institute Freiestrasse 1 3012 Bern Switzerland

Patrizia Stoitzner

Department of Dermatology and Venereology Innsbruck Medical University Anichstrasse 35 6020 Innsbruck Austria

Christoph Tripp

Innsbruck Medical University Department of Dermatology and Venereology Anichstrasse 35 6020 Innsbruck Austria

XXII List of Authors

Barbara Walzog Ludwig-Maximilians-University Department of Physiology Schillerstr. 44 80336 Munich Germany

Olaf Zilles

Lund University Department of Experimental Pathology Soelvegatan 25 22362 Lund Sweden [Text not available in this electronic edition.]

xxiv

[Text not available in this electronic edition.]

Color Plates



Fig. 2.1. Structural organization of selectins. The N-terminal domain of each selectin is homologous to C-type lectins and binds to carbohydrate groups on their respective ligands. Following this is an epidermal growth factor-like domain and then a variable number of short consensus repeats homologous to complement regulatory proteins. The arrow indicates the cleavage site of L-selectin. (This figure also appears on page 15.)



Fig. 2.2. Shedding of L-selectin by leukocytes. In resting leukocytes, calmodulin, α -actinin, and ERM proteins are associated with the cytoplasmic tail of L-selectin as well as actin filaments. Upon cell activation by TNF α , IL-1, PMA, etc., calmodulin is released and L- selectin sheddase cleaves the 69-kDa extracellular domain whilst α -actinin and ERM proteins retain their contact and are involved in microvilli localization as well as leukocyte tethering. (This figure also appears on page 18.)



Fig. 2.3. Multiple ligands for L-, P-, and E-selectin have been detected on endothelial cells and leukocytes. (This figure also appears on page 23.)



Fig. 5.1. Interaction of key endothelial cell molecules implicated in the process of transendothelial cell migration with their respective leukocyte ligands. Key signaling pathways implicated with specific molecules/ molecular interactions are also shown. It should be noted that with respect to the JAM molecules, although homophilic interactions have been shown in other systems (e.g., in endothelial cell-endothelial cell interactions; see text for details), this has not always been demonstrated in the context of leukocyte/

endothelial cell interaction, though it is clearly a possibility. FAK, focal adhesion kinase; T cell receptor (TCR); ERM, ezrin-radixin-moesin; PKC, protein kinase C; ADAP, adhesion and degranulation promoting adaptor protein; SKAP-55, src kinase-associated phosphoprotein of 55 kDa; MAPK, mitogen-activated protein kinase; SHP-2, Src homology domain protein-2; ITIM, immunoreceptor tyrosine-based inhibitory motif; MHC, major histocompatibility complex. (This figure also appears on page 85.)