## **HANDBOOK OF FOOD ANALYTICAL CHEMISTRY**

**WATER, PROTEINS, ENZYMES, LIPIDS, AND CARBOHYDRATES** 

## **HANDBOOK OF FOOD ANALYTICAL CH EM ISTRY**

**PIGMENTS, COLORANTS, FLAVORS, COMPONENTS TEXTURE, AND BIOACTIVE FOOD** 

# **HANDBOOK OF FOOD ANALYTICAL CHEMISTRY**

## **HANDBOOK OF FOOD ANALYTICAL CHEMISTRY**

## **WATER, PROTEINS, ENZYMES, LIPIDS, AND CARBOHYDRATES**

Edited by

Ronald E. Wrolstad Terry E. Acree Eric A. Decker Michael H. Penner David S. Reid Steven J. Schwartz Charles **F.** Shoemaker Denise Smith Peter Sporns



## **HANDBOOK OF FOOD ANALYTICAL CH EM ISTRY**

## **PIGMENTS, COLORANTS, FLAVORS, COMPONENTS TEXTURE, AND BIOACTIVE FOOD**

Edited by

Ronald E. Wrolstad Terry E. Acree Eric A. Decker Michael H. Penner David S. Reid Steven **J.** Schwartz Charles F. Shoemaker Denise Smith Peter Sporns



A JOHN WILEY & SONS, INC., PUBLICATION

Copyright  $\odot$  2000-2005 by John Wiley & Sons, Inc. All rights reserved.

Published by John Wiley & Sons, Inc., Hoboken, New Jersey. Published simultaneously in Canada.

No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording, scanning, or otherwise, except as permitted under Section 107 or 108 of the 1976 United States Copyright Act, without either the prior written permission of the Publisher, or authorization through payment of the appropriate per-copy fee to the Copyright Clearance Center, Inc., 222 Rosewood Drive, Danvers, MA 01923, 978-750-8400, fax 978-646-8600, or on the web at www.copyright.com. Requests to the Publisher for permission should be addressed to the Permissions Department, John Wiley & Sons, Inc., 11 **1** River Street, Hoboken, NJ 07030, (201) 748-601 I, fax (201) 748-6008.

Limit of Liability/Disclaimer of Warranty: While the publisher and author have used their best efforts in preparing this book, they make no representations or warranties with respect to the accuracy or completeness of the contents **of**  this book and specifically disclaim any implied warranties of merchantability or fitness for a particular purpose. No warranty may be created or extended by sales representatives or written sales materials. The advice and strategies contained herein may not be suitable for your situation. You should consult with a professional where appropriate. Neither the publisher nor author shall be liable for any loss of profit or any other commercial damages, including but not limited to special, incidental, consequential, or other damages.

For general information on our other products and services please contact our Customer Care Department within the US. at 877-762-2974, outside the **U.S.** at 317-572-3993 or fax 317-572-4002.

Wiley also publishes its books in a variety of electronic formats. Some content that appears in print, however, may not be available in electronic format.

### *Library of Congress Cataloging-in-Publication Data:*

Handbook of food analytical chemistry / edited By Ronald E. Wrolstad . . . [et al.]. Includes bibliographical references and index. ISBN 0-471-66378-6 Volume I (cloth), ISBN 0-47 1-7 18 17-3 Volume 2 (cloth) ISBN 0-471-72187-5 (set) p. cm.

**1.** Food--Analysis--Handbooks, manuals, etc. **I.** Wrolstad, Ronald E. TX545.H34 2005 664'.07--dc22

2004013225

Printed in the United States of America

10 9 8 7 6 *5* 4 *3* 2 **1** 

## **Contents**

- **ix Preface**
- **xi Foreword to Current Protocols in Food Analytical Chemistry**
- **xii Contributors**

### **A** WATER 1

### **A1 Gravimetric Measurements of Water** / **5**

- Al.l Gravimetric Determination of Water by Drying and Weighing *I* 7
- A1.2 Karl Fischer Titration *I* 13
- A 1.3 Application of Low-Resolution NMR for Simultaneous Moisture and Oil Determination in Food (Oilseeds) *I* 17
- Traditional Indirect Methods for Estimation of Water Content: Measurement of <sup>o</sup>Brix / 29 Al.4

### **A2 Vapor Pressure Measurements of Water** / **35**

- A2.1 Factors to Consider When Estimating Water Vapor Pressure *I* 37
- Dew-Point Method for the Determination of Water Activity *I* 41 A2.2
- A2.3 Measurement of Water Activity Using Isopiestic Method / 51
- A2.4 Direct Manometric Determination of Vapor Pressure / 61
- A2.5 Measurement of Water Activity by Electronic Sensors / 67

### **PROTEINS**  B

### 71

#### **B1 Measurement of Protein Content** / **73**

- B 1.1 The Colorimetric Detection and Quantitation of Total Protein *I* 77
- B1.2 Determination of Total Nitrogen / 105
- B 1.3 Spectrophotometric Determination of Protein Concentration *I* 115

### **B2 Biochemical Compositional Analyses of Proteins** / **123**

- B2.1 Analyses of Protein Quality / 125
- B2.2 Evaluation of the Progress of Protein Hydrolysis / 141

#### **B3 Characterization of Proteins** / **155**

- B3.1 Electrophoresis Analysis *I* 157
- B3.2 Electroblotting from Polyacrylamide Gels / 185
- B3.3 Detection of Proteins on Blot Membranes / 199
- B3.4 Immunoblot Detection / 207
- B3.5 Determining the CD Spectrum of a Protein *I* 219
- B3.6 Determining the Fluorescence Spectrum of a Protein / 245

### **B4 Purification of Proteins** / **267**

- B4.1 Overview of Protein Purification and Characterization / 269
- B4.2 Overview of Conventional Chromatography / 279

### **B5 Functionality of Proteins** / **289**

- B.5. 1 Measurement of Functional Properties: Overview of Protein Functionality Testing / 291
- B5.2 Measurement of Protein Hydrophobicity / 301
- B5.3 Water Retention Properties of Solid Foods / 315

### **C ENZYMES 325**

### **C1 Strategies for Enzyme Activity Measurements** / **329**

- C1.l Expression and Measurement of Enzyme Activity / 331
- C1.2 Detecting Enzyme Activity: A Case Study of Polygalacturonase / 335

### **C2 Proteolytic Enzymes** / **349**

- C2.1 Activity Measurements of Proteinases Using Synthetic Substrates / 351
- C2.2 Peptidase Activity Assays Using Protein Substrates / 359

### **C3 Lipolytic Enzymes** / **369**

C3.1 Lipase Assays / 371

### **C4 Oxidoreductases** / **385**

- C4.1 Polarographic and Spectrophotometric Assay of Diphenol Oxidases (Polyphenol Oxidase) / 387
- C4.2 Analysis of Lipoxygenase Activity and Products / 403

### D **LIPIDS 419**

### **Dl Lipid Composition** / **423**

- D1.l Extraction and Measurement of Total Lipids / 425
- D1.2 Analysis of Fatty Acids in Food Lipids / 437
- D1.3 Cholesterol / 453
- D1.4 Oil Quality Indices / 467
- D1.5 Analysis of Tocopherols and Tocotrienols / 479
- D1.6 Quantitation of Lipid Classes by Thin-Layer Chromatography with Flame Ionization Detection / 491
- D1.7 Infrared Spectroscopic Determination of Total *Trans* Fatty Acids / 505

### **D2 Lipid Oxidation/Stability** / **513**

- D2.1 Measurement of Primary Lipid Oxidation Products / *5* 1.5
- D2.2 Chromatographic Analysis of Secondary Lipid Oxidation Products / 531
- D2.3 Assessment of Oxidative Stability for Lipids / 541
- D2.4 Spectrophotometric Measurement of Secondary Lipid Oxidation Products / 547

### **D3 Physical Properties of Lipids** / **565**

- D3.1 Determination of Solid Fat Content by Nuclear Magnetic Resonance *I* 567
- D3.2 Lipid Crystal Characterization *I* 575
- D3.3 Emulsion Droplet Size Determination *I* 581
- D3.4 Emulsion Stability Determination *I* 591
- D3.5 Key Concepts of Interfacial Properties in Food Chemistry *I* 609
- D3.6 Static and Dynamic Interfacial Tension Analysis / 631

### E **CARBOHYDRATES 647**

#### **El Mono- and Oligosaccharides** / **651**

- E 1.1 Colorimetric Quantification of Carbohydrates *I* 653
- E1.2 HPLC of Mono- and Disaccharides Using Refractive Index Detection / 661

#### **E2 Starch and Starch Derivatives** / **671**

- E2.1 Overview of Laboratory Isolation of Starch from Plant Materials *I* 673
- E2.2 Enzymatic Quantitation of Total Starch in Plant Products *I* 679
- E2.3 Determination of Total Amylose Content of Starch *I* 689

#### **E3 Cell Wall Polysaccharides** / **695**

- E3.1 Isolation of Plant Cell Walls and Fractionation of Cell Wall Polysaccharides *I* 697
- Determination of Neutral Sugars by Gas Chromatography of Their Alditol Acetates *I* 721 E3.2
- Determination of the Uronic Acid Content of Plant Cell Walls Using a Colorimetric Assay *I* 735 E3.3
- Determining the Degree of Methylation and Acetylation of Pectin *I* 739 E3.4
- E3.5 Quantitative Determination of P-Glucan Content *I* 745

### **Index** / **757**

## **Contents**

- **ix Preface**
- **xi Foreword to Current Protocols in Food Analytical Chemistry**
- **xii Contributors**

### **F PIGMENTS AND COLORANTS 1**

#### **F1 Anthocyanins** / **5**

- Fl.l Extraction, Isolation, and Purification of Anthocyanins *I* 7
- F1.2 Characterization and Measurement of Anthocyanins by UV-Visible Spectroscopy *I* 19
- F1.3 Separation and Characterization of Anthocyanins by HPLC *I* 33
- F1.4 Characterization of Anthocyanins by NMR *I* 47

#### **F2 Carotenoids** / **71**

- F2.1 Extraction, Isolation, and Purification of Carotenoids *I* 73
- F2.2 Detection and Measurement of Carotenoids by UV/VIS Spectrophotometry *I* 81
- F2.3 Chromatographic Techniques for Carotenoid Separation / 91
- F2.4 Mass Spectrometry of Carotenoids *I* 107

#### **F3 Miscellaneous Colorants** / **121**

- F3.1 Betalains *I* 123
- F3.2 Spectrophotometric and Reflectance Measurements of Pigments of Cooked and Cured Meats *I* 131
- F3.3 Measurement of Discoloration in Fresh Meat *I* 139

#### **F4 Chlorophylls** / **153**

- F4.1 Overview of Chlorophylls in Foods *I* 155
- F4.2 Extraction of Photosynthetic Tissues: Chlorophylls and Carotenoids / 165
- F4.3 Chlorophylls and Carotenoids: Measurement and Characterization by UV-VIS Spectroscopy *I* 171
- F4.4 Chromatographic Separation of Chlorophylls *I* 179
- F4.5 Mass Spectrometry of Chlorophylls *I* 191

### **F5 Strategies for Measurement of Colors and Pigments** / **201**

F5.1 Overview of Color Analysis *I* 203

### **G FLAVORS**

### **G1 Smell Chemicals** / **223**

- G1.l Direct Sampling *I* 225
- G1.2 Isolation and Concentration of Aroma Compounds *I*  235
- GI *.3* Identification and Quantitation of Aroma Compounds *I* 245
- **217**

- G1.4 Stereodifferentiation of Chiral Odorants Using High-Resolution Gas Chromatography / 257
- G1.5 Analysis of Citrus Oils / 277
- G1.6 Solid-Phase Microextraction for Flavor Analysis / 301
- G1.7 Simulation of Mouth Conditions for Flavor Analysis / 313
- G1.8 Gas Chromatography/Olfactometry / 329

### **G2 Acid Tastants** / **341**

- G2.1 Titratable Activity of Acid Tastants / 343
- G2.2 Liquid Chromatography of Nonvolatile Acids / 351

### **H TEXTURE/RHEOLOGY 363**

### **H1 Viscosity of Liquids, Solutions, and Fine Suspensions** / **367**

- H1.l Overview of Viscosity and Its Characterization / 369
- H1.2 Measuring the Viscosity of Non-Newtonian Fluids / 375
- HI .3 Viscosity Determination of Pure Liquids, Solutions, and Serums Using Capillary Viscometry / 385
- H1.4 Measuring Consistency of Juices and Pastes / 391

### **H2 Compressive Measurements of Solids and Semi-Solids** / **395**

- H2.1 General Compressive Measurements / 397
- H2.2 Textural Measurements with Special Fixtures / 405
- H2.3 Texture Profile Analysis / 417

#### **H3 Viscoelasticity of Suspensions and Gels** / **425**

- H3.1 Dynamic or Oscillatory Testing of Complex Fluids / 427
- H3.2 Measurement of Gel Rheology: Dynamic Tests / 439
- H3.3 Creep and Stress Relaxation: Step-Change Experiments / 449

#### **I Bioactive Food Components**

**457** 

- I1 Polyphenolics / 461
- 11.1 Determination of Total Phenolics / 463
- 11.2 Extraction and Isolation of Polyphenolics / 471
- 11.3 HPLC Separation of Polyphenolics / 483
- 11.4 Proanthocyanidins: Extraction, Purification, and Determination of Subunit Composition by HPLC / 499
- 11.5 Identification of Flavonol Glycosides Using MALDI-MS / 511
- 11.6 Analysis of Isoflavones in Soy Foods / 519

### **APPENDICES AND INDEXES**

### **Appendices** / **537**

- A.l Abbreviations and Useful Data / 539
	- A Abbreviations Used in This Manual / 539
- A.2 Laboratory Stock Solutions, Equipment, and Guidelines *I* 543
	- A Common Buffers and Stock Solutions *I* 543
	- B Laboratory Safety *I* 551
	- C Standard Laboratory Equipment *I* 553
- A.3 Commonly Used Techniques *I 555* 
	- A Introduction *to* Mass Spectrometry for Food Chemistry *I 555*

**Suppliers Appendix** / **563** 

**Index** / **597** 

### **PREFACE**

ccurate and state-of-the-art analysis of food composition is of interest and concern to a divergent clientele including research workers in academic, government and industrial settings, regulatory scientists, analysts in private commercial laboratories, and quality control professionals in small and large companies. Some methods are empirical, some commodity specific, and many have been widely accepted as standard methods for years. Others are at the cutting edge of new analytical methodology and are rapidly changing. A common denominator within this diverse group of methods is the desire for detailed descriptions of how to carry out analytical procedures. **A** frustration of many authors and readers of peer-reviewed journals is the brevity of most Materials and Methods sections. There is editorial pressure to minimize description of experimental details and eliminate advisory comments. When one needs to undertake an analytical procedure with which one is unfamiliar, it is prudent to communicate firsthand with one experienced with the methodology. This may require a personal visit to another laboratory and/or electronic or phone communication with someone who has expertise in the procedure. An objective of *the Handbook of Food Analytical Chemistry* is to provide exactly this kind of detailed information which personal contact would provide. Authors are instructed to present the kind of details and advisory comments they would give to a graduate student or technician who has competent laboratory skills and who has come to them to learn how to carry out an analytical procedure for which the author has expertise.

Some basic food analytical methods such as determination of  $\delta$ brix, pH, titratable acidity, total proteins and total lipids are basic to food analysis and grounded in procedures which have had wide-spread acceptance for a long time. Others such as analysis of cell-wall polysaccharides. analysis of aroma volatiles, and compressive measurement of solids and semi-solids, require use of advanced chemical and physical methods and sophisticated instrumentation. In organizing *the Handbook* of *Food Analytical Chemistry* we chose to categorize on a disciplinary rather than a commodity basis. Included are chapters on water, proteins, enzymes, lipids, carbohydrates, colors, flavors texture/ rheology and bioactive food components. We have made an effort to select methods that are applicable to all commodities. However, it is impossible to address the unique and special criteria required for analysis of all commodities and all processed forms. There are several professional and trade organizations which focus on their specific commodities, e.g., cereals, wines, lipids, fisheries, and meats. Their methods manuals and professional journals should be consulted, particularly for specialized, commodity-specific analyses.

This two-volume handbook is derived from another John Wiley & Sons publication, *Current Protocol in Food Analytical Chemistry.* That manual was published from January 2001 -December 2003 in loose-leaf and CD-Rom format. That design permitted addition of new and revised units on a quarterly basis. The two-year compilation of these units makes for a very complete reference on food analytical methods.

### **FOREWORD TO CURRENT PROTOCOLS IN FOOD ANALYTICAL CHEMISTRY**

ccurate, precise, sensitive, and rapid analytical determinations are as essential in **A** food science and technology as in chemistry, biochemistry, and other physical and biological sciences. In many cases, the same methodologies are used. How does one, especially a young scientist, select the best methods to use? A review of original publications in a given field indicates that some methods are cited repeatedly by many noted researchers and analysts, but with some modifications adapting them to the specific material analyzed. Official analytical methods have been adopted by some professional societies, such as the Official Methods of Analysis (Association of Official Analytical Chemists), Official Methods and Recommendation Practices (American Oil Chemists' Society), and Official Methods of Analysis (American Association of Cereal Chemists).

The objective of *Current Protocols in Food Analytical Chemistry* is to provide the type of detailed instructions and comments that an expert would pass on to a competent technician or graduate student who needs to learn and use an unfamiliar analytical procedure, but one that is routine in the lab of an expert or in the field.

What factors can be used to predetermine the quality and utility of a method? An analyst must consider the following questions: Do I need a proximate analytical method that will determine all the protein, or carbohydrate, or lipid, or nucleic acid in a biological material? Or do I need to determine one specific chemical compound among the thousands of compounds found in a food? Do I need to determine one or more physical properties of a food? How do I obtain a representative sample? What size sample should I collect? How do I store my samples until analysis? What is the precision (reproducibility) and accuracy of the method or what other compounds and conditions could interfere with the analysis? How do I determine whether the results are correct, as well as the precision and accuracy of a method? How do I know that my standard curves are correct? What blanks, controls and internal standards must be used? How do I convert instrumental values (such as absorbance) to molar concentrations? How many times should I repeat the analysis? And how do I report my results with appropriate standard deviation and to the correct number of significant digits? Is a rate of change method (i.e., velocity as in enzymatic assays) or a static method (independent of time) needed?

*Current Protocols in Food Analytical Chemistry* will provide answers to these questions. Analytical instrumentation has evolved very rapidly during the last 20 years as physicists, chemists, and engineers have invented highly sensitive spectrophotometers, polarometers, balances, etc. Chemical analyses can now be made using milligram, microgram, nanogram, or picogram amounts of materials within a few minutes, rather than previously when grams or kilograms of materials were required by multistep methods requiring hours or days of preparation and analysis. *Current Protocols in Food Analytical Chemistry* provides state-of-the-art methods to take advantage of the major advances in sensitivity, precision, and accuracy of current instrumentation.

How do chemical analyses of foods differ from analyses used in chemistry, biochemistry and biology? The same methods and techniques are often used; only the purpose of the analysis may differ. But foods are to be used by people. Therefore, methodology to determine safety (presence of dangerous microbes, pesticides, and toxicants), acceptability (flavor, odor, color, texture), and nutritional quality (essential vitamins, minerals, amino acids, and lipids) are essential analyses. *Current Protocols in Food Analytical Chemistry* is designed to meet all these requirements.

> John Whitaker Davis, California

### **CONTRIBUTORS**

Terry E. Acree Cornell University Geneva. New York

Ozlem Akpinar Oregon State University Corvallis, Oregon

0yvind M. Andersen University of Bergen Bergen, Norway

Tom Berkelman Amersham Pharmacia Biotech San Francisco, California

Hugues Brevard Nestle Research Center Lausanne, Switzerland

Zvonko Burkus University of Alberta Edmonton, Canada

Claus Buschmann Universitaet Karlsruhe Karlsruhe, Germany

Pavinee Chinachoti University of Massachusetts Alberta, Canada Shinya Ikeda<br>Amherst, Massachusetts Shinya Ikeda

Mary G. Chisholm Behrend College, The Pennsylvania State University Erie, Pennsylvania

Utah State University Logan, Utah

John Coupland Tempe, Arizona Pennsylvania State University University Park, Pennsylvania

Neal E. Craft Craft Technologies Wilson, North Carolina

Susan L. Cuppett University of Nebraska-Lincoln Lincoln, Nebraska

Kathryn D. Deibler Cornell University Geneva, New York Robert W. Durst Oregon State University Corvallis, Oregon

Wayne Ellefson Covance Laboratories Madison, Wisconsin

Cameron Faustman University of Connecticut Storrs, Connecticut

Mario G. Feruzzi Ohio State University Columbus. Ohio

E. Allen Foegeding North Carolina State University Raleigh, North Carolina

Anthony **J.** Fontana Decagon Devices Pullman, Washington

Torgils Fossen St. John's, Canada University of Bergen Bergen, Norway

Madrid, Spain Eric Fournier University of Alberta Alberta, Canada

Jane E. Friedrich Osaka, Japan Cargill Incorporated Minneapolis, Minnesota

Suzanne Frison Corvallis, Oregon University of Alberta Daren Cornforth Edmonton, Alberta, Canada

Motorola, Inc.

Fernando García-Carreño Corvallis, Oregon Centro de Investigaciones Biol6gicas (CIBNOR) La Paz, Mexico

Harold W. Gardner National Center for Agricultural Peoria, Illinois Utilization Research, ARS, USDA

Trevor Gentry Cornell University Ithaca, New York

M. M6nica Giusti University of Maryland College Park, Maryland

N. Guizani College of Agriculture, Sultan Muscat, Sultanate of Oman Qaboos University

Sandra Harper The Wistar Institute Philadelphia, Pennsylvania

R.W. Hartel University of Wisconsin Madison, Wisconsin

M.L. Herrera University of Wisconsin Madison, Wisconsin

R. Hoover Memorial University of Newfoundland

Montana Camara Hurtado Universidad Complutense de Madrid

Osaka City University

James A. Kennedy Oregon State University

Dae-Ok Kim Cornell University Sean Gallagher Geneva, New York

> Sasithorn Kongruang Oregon State University

> Magnus M. Kristjansson University of Iceland Reykjavik, Iceland

Randall **1.** Krohn Pierce Chemical Rockford, Illinois

Peter H. Krygsman Bruker Ltd. Milton, Canada

Theodore P. Labuza University of Minnesota St. Paul, Minnesota

Duane K. Larick North Carolina State University Raleigh, North Carolina

Chang Y. Lee Cornell University Geneva, New York

P.P. Lewicki Warsaw Agricultural University (SGGW) Warsaw, Poland

Yong Li Purdue University West Lafayette, Indiana

Hartmut K. Lichtenthaler Universitaet Karlsruhe Karlsruhe, Germany

Kevin Loughrey Gretag Macbeth South Deerfield. Massachusetts

George Lunn Baltimore, Maryland

D. Julian McClements University of Massachusetts Amherst. Massachusetts

Richard E. McDonald Food and Drug Administration College Park, Maryland

Laurence D. Melton University of Auckland Auckland. New Zealand

Christian Milo Nestle Research Center Lausanne. Switzerland

Yoshi Mochizuki University of California Davis. California

Magdi M. Mossoba Food and Drug Administration College Park, Maryland

Shuryo Nakai University of British Columbia Vancouver, British Columbia, Canada

M. Angeles Navarrete del Toro Centro de Investigaciones Biol6gicas (CIBNOR) La Paz, Mexico

Toshiaki Ohshima Tokyo University of Fisheries Tokyo, Japan

Roger H. Pain Jozef Stefan Institute Ljubljana, Slovenia Gustavo A. Rodriguez

James D. Parker North Carolina State University Raleigh, North Carolina

Kirk L. Parkin University of Wisconsin Madison, Wisconsin

Ronald B. Pegg University of Saskatchewan Saskatoon, Canada

Michael H. Penner Oregon State University Corvallis, Oregon

Amy Phillips University of Connecticut Storrs, Connecticut

Oscar **A.** Pike Brigham Young University Provo, Utah

Praphan Pinsirodom University of Wisconsin Madison. Wisconsin

M. Shafiur Rahman Sultan Qaboos University Muscat, Sultanate of Oman

Barbara Rasco Washington State University Pullman, Washington

W. **S.** Ratnayake Memorial University of Newfoundland St. John's, Canada

Joe M. Regenstein Cornell University Ithaca, New York

David **S.** Reid University of California at Davis Davis, California

Jody Renner-Nantz University of California Davis, Cailfornia

Khee C. Rhee Texas A&M University College Station, Texas

Deborah Roberts Nestle Research Center Lausanne, Switzerland

Prodemex Los Mochis, Mexico

Luis E. Rodriguez-Saona University of Maryland and Joint Institute for Food Safety and Applied Nutrition Washington, D. C.

Michael D.H. Rogers University of Guelph Guelph, Canada

Rennie P. Ruiz Hunt-Wesson, Inc. Fullerton, California

Shyam S. Sablani Sultan Qaboos University Muscat, Sultanate of Oman

Steven J. Schwartz Ohio State University Columbus, Ohio

R.K. Scopes LaTrobe University Bundoora. Australia

K. John Scott Institute of Food Research Colney, United Kingdom

Fereidoon Shahidi Memorial University of Newfoundland St. John's, Canada

Michael H. Simonian Beckman Coulter Fullerton, California

Bronwen *G.* Smith University of Auckland Auckland, New Zealand

Denise M. Smith University of Idaho Moscow, Idaho

David W. Speicher The Wistar Institute Philadelphia, Pennsylvania

Peter Sporns University of Alberta Edmonton, Alberta, Canada

Feral Temelli University of Alberta Edmonton, Canada

Marvin A. Tung University of Guelph Guelph, Canada

Richard B. van Breemen University of Illinois at Chicago Chicago, Illinois

Saskia van Ruth University College Cork Cork. Ireland

Thava Vasanthan University of Alberta Edmonton. Canada

Joachim H. von Elbe University of Wisconsin Madison, Wisconsin

John R. L. Walker University of Canterbury Christchurch, New Zealand

Andrew L. Waterhouse University of California, Davis Davis. California

Bruce **A.** Watkins Purdue University West Lafayette, Indiana

Jochen Weiss University of Tennessee Knoxville, Tennessee

Peter Whittingstall ConAgra Grocery Products Irvine, California

Alan Williams Amersham Pharmacia Biotech Piscataway, New Jersey

Ronald E. Wrolstad, Oregon State University Corvallis, Oregon

Yu Chu Zhang Ohio State University Columbus. Ohio

Shengying Zhou The Minute Maid Company Apopka, Florida

# <sup>I</sup>**A WATER**

### **INTRODUCTION**

### **A1 Gravimetric Measurements of Water**

- A1.1 Gravimetric Determination of Water by Drying and Weighing
- A 1.2 Karl Fischer Titration
- Al.3 Application of Low-Resolution NMR for Simultaneous Moisture and Oil Determination in Food (Oilseeds)
- A 1.4 Traditional Indirect Methods for Estimation of Water Content: Measurement of "Brix

### **A2 Vapor Pressure Measurements of Water**

- A2.1 Factors to Consider When Estimating Water Vapor Pressure
- A2.2 Dew-Point Method for the Determination of Water Activity
- A2.3 Measurement of Water Activity Using Isopiestic Method
- A2.4 Direct Manometric Determination of Vapor Pressure
- A2.5 Measurement of Water Activity by Electronic Sensors

**1** 

## **SECTION A Water**

### **INTRODUCTION**

 $\sum_{n=1}^{\infty}$  ater determination in foods is a deceptively simple theme. Defining the quantity to be measured identifies the inherent complexity. Three separate types of measure may be appropriate: (a) a gravimetric measure, (b) a measure related to vapor pressure, and (c) a measure of the mobilities of water molecules. The ubiquitous nature of water in our environment provides additional complexity in the challenge of preventing transfer of water between sample and environment. The earliest measures of amount of water were all gravimetric, determining the weight fraction of water in the food. These methods range from simple direct weighing, using a difference technique, to more complex methods where the amount of water is determined by spectroscopic methods or by chemical assay. **A** wide range of methods have been developed and are in daily use, since gravimetric water content is important for formulation and for labeling purposes. This measure, however, is of little value for the prediction of the stability of a food, even though water plays a critical role in determining the stability characteristics of foods.

For a measure of amount of water relevant to stability concerns, vapor pressure, or its related thermodynamic parameters, is more relevant. Determination of vapor pressure uses methods developed from thermodynamic roots, though if the product is not at true equilibrium, the measured quantity is not a thermodynamic descriptor of the product, although it is still a measure of a product characteristic. Water mobilities are often inferred from spectroscopic measurements of relaxational phenomena. Many workers attempt to identify different "classes" of water characteristic of different ranges of water content and water partial vapor pressure. Spectroscopic measurements, too, are often interpreted in terms of populations of water molecules with similar characteristics.

The objective of this section is to provide clear descriptions of the alternative methods for the determination of gravimetric water content (Chapter **Al)** and of the range of methods available for the estimation of vapor pressure or its related parameters (Chapter **A2).** 

### **GRAVIMETRIC MEASUREMENTS OF WATER**

*UNITA 1.1* describes the direct determination of gravimetric water by drying and weighing, surveying a range of well established techniques. *UNITAI.Z* describes the use of the Karl Fischer titration for the chemical determination of the amount of water contained in a sample. *UNITAI.3* describes a particular use of nuclear magnetic resonance spectroscopy (NMR) to determine the water content and the oil content of seeds. *UNITAI.4* provides an overview of some indirect methods for the estimation of water content. It considers the pros and cons of the measurement of physical characteristics (density and refractive index) that may be correlated with the water content of specific systems, and identifies the critical assumptions associated with the use of such indirect methods.

### **VAPOR PRESSURE MEASUREMENTS OF WATER**

The initial unit in this chapter *(UNIT A2.1)* discusses the constraints which must be considered when attempting to estimate the vapor pressure above an aqueous system. These constraints are operative whichever technique may be utilized. The use of a dew-point cell to estimate vapor pressure and water activity is described in *UNITA2.2*. This unit clearly identifies the precautions which are essential to a good dew-point determination. The use of isopiestic techniques, in which a known atmospheric condition is produced and the sample is assumed to have equilibrated with this atmosphere, is the subject of *UNIT A2.3.* The techniques of this unit are frequently employed for the special purpose of determining moisture sorption isotherms. These describe the relationship between the gravimetric water content of a sample and the partial water vapor pressure sustained by the sample. Such relationships can be a useful tool for correlating/estimating moisture content from partial water vapor pressure measurements and vice versa. *UNITA2.4* describes the direct manometric measurement of water vapor pressure. The method is very demanding of good technique, which is why it is seldom used. All primary vapor pressure data result from the use of this type of apparatus. The primary vapor pressure data for pure water (used as the reference data for all of the indirect methods, including dew point, isopiestic, etc.) were produced by direct manometric evaluation. *UNIT A2.5* describes the use of electronic sensors for vapor pressure measurement. This method is considered the most simple method for measuring water activity. Advantages and limitations of different types of electronic sensors are discussed.

David **S.** Reid

**Introduction** 

## <sup>1</sup>**Gravimetric Measurements of Water**





**Contents** 

## **Gravimetric Determination of Water by Drying and Weighing**

Water (moisture) in a sample is measured gravimetrically by determining the weight loss in a sample after it has been placed in an appropriate oven (convection, vacuum, or microwave) for a given time. In addition, there are automatic moisture analyzers available that utilize infrared lamps as a heat source. These types of moisture analyzers are fast but many times are matrix dependent, which requires some trial-and-error testing to determine the correct settings (power and time). Water and moisture are used interchangeably in the description of these protocols. In addition, it is assumed in the gravimetric method that only water is removed in the drying process, when in fact there may be volatile loss in some samples.

Although the measurement of weight loss due to evaporation of water is frequently used to calculate moisture content, it should be pointed out that the value obtained may not be a true measure of water content. In some samples, only a proportion of the water present is lost at the drying temperature. The balance (bound water) is difficult to remove completely. In addition, the water lost may actually increase as the temperature is raised. Some samples with high fat content may exhibit volatile oil loss at drying temperatures of 100°C. Weight loss may also be dependent on such factors as particle size, weight of samples used, type of dish used, and temperature variations in the oven from shelf to shelf. Thus, it is important to compare results obtained using the same drying conditions.

This unit provides three protocols for which there are established procedures for various matrices. The Basic Protocol describes water removal and quantitation after a sample is placed in a convection oven. It is probably the method of choice when one does not know which method to choose when dealing with an unknown matrix, or when one looks at samples that foam excessively in the vacuum oven method or "react," such as popcorn under vacuum. Alternate Protocol **1** describes water removal and quantitation after a sample is placed in a vacuum oven. Because it is at reduced pressure, drying times are slightly reduced compared to the convection method. In addition, drying temperatures  $<$ 100 $\degree$ C are possible, which is important for samples that may decompose at higher drying temperatures. Alternate Protocol 2 describes water removal using a microwave source where such analyzers measure and calculate loss automatically.

### **MEASURING MOISTURE USING A CONVECTION OVEN**

Water is measured in a sample by determining the loss in weight for the sample after it has been dried in a convection oven. The method requires only a small amount of homogeneous sample and can measure an effective range of 0.01% to 99.99% water.

### *Materials*

Homogeneous sample Convection oven capable of maintaining a temperature of  $103^{\circ} \pm 2^{\circ}C$ Aluminum weighing dishes (with or without covers) Desiccator with desiccant Balance capable of measuring  $\pm 0.1$  mg

- 1. Set the temperature of **a** convection oven to 105°C.
- 2. Dry an aluminum weighing dish (and cover, if used)  $\geq 1$  hr at 105 $^{\circ}$ C. Cool and store dried dish in a desiccator. Cool 230 min before using.

*Covered weighing dishes are useful when analyzing samples that splatter: Weighing dishes without covers may otherwise be preferred, as they are disposable.* 

*BASIC PROTOCOL* 

**Gravimetric Measurements of Water**