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The Lightest Metals Science and Technology from Lithium to Calcium

Encyclopedia of Inorganic and Bioinorganic Chemistry



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Figure 27 (a) ²⁷Al static and MAS CT NMR spectra of Al₃Ti. (b) Expansion of the MAS spectrum in (a) with accompanying simulation (shown below and above, respectively). (c) ²⁷Al static spectrum of Al₃Ti showing the overlapping satellite ($\pm 1/2 \leftrightarrow \pm 3/2$ and $\pm 3/2 \leftrightarrow \pm 5/2$) line shapes. Resonances corresponding to metallic aluminum are denoted with * and ± 10 (b) and (c), respectively

Figure 28 3Q-D-HETCOR (3Q = triple quantum; D = dipolar correlation) NMR spectrum (left) of aluminophosphate AlPO₄-14 showing the correlations of tetrahedral and five-coordinated aluminum sites with phosphorus. The ³¹P skyline projections are shown above the spectrum, which were observed (a) without and (b) with SPAM. The total acquisition time was 20 h. (c) ²⁷Al-²⁹Si D-HETCOR, and (d) ²⁷Al-²⁹Si SPAM-3Q-D-HETCOR NMR spectra of microcline. The 1D ²⁹SI MAS spectrum is shown for comparison on top of (c). The total acquisition times were 80 and 72 h for (c) and (d), respectively. All spectra were acquired at 9.4 T

Figure 29 ²⁷Al MQMAS NMR spectrum of steamed zeolite β, (H)β1 550(3d)w30. Peaks corresponding to three tetrahedral aluminums (Al(IV)_a, Al(IV)_b, and Al(IV)_c) and two octahedral aluminums (Al(VI)_a and Al(VI)_b) are visible. The ²⁷Al MAS NMR spectrum is presented on top of the MQMAS spectrum Figure 30 27 Al MAS NMR spectra of super-quenched glass samples showing peaks corresponding to 4-, 5-, and 6-coordinate Al sites, and spectra of nonquenched sample only showing peaks corresponding to 4- and 6-coordinate sites. The sample compositions are indicated for each spectrum in mol % with SiO₂ and Al₂O₃ denoted by S and A, respectively. All spectra were acquired with a spinning speed of around 15 kHz

<u>Figure 31 Frequency-stepped ²⁷Al CPMG NMR</u> <u>spectra of (a) AlMes₃ and (b) Al(NTMS₂)₃. The</u> <u>individual sub-spectra are shown on top with the</u> <u>corresponding total powder pattern shown on the</u> <u>bottom. Co-addition was used to form the total</u> <u>powder pattern. † and # indicate FM radio signal</u> <u>interference and impurities, respectively</u>

<u>Figure 32 Experimental (top) and simulated (bottom)</u> <u>static ²⁷Al powder patterns of [Cp*₂Al][AlCl₄]</u> <u>collected at 9.4 T. The patterns are dominated by the</u> <u>effects of the aluminum CSA</u>

<u>Figure 33 (a)</u> ³⁹<u>K NMR lineshapes for alkali metal C₆₀</u> <u>compounds, given in order of increasing lattice</u> <u>constant from top to bottom.</u> ³⁹<u>K NMR lineshapes at</u> <u>several temperatures for (b) K₃C₆₀, and (c) Rb₂KC₆₀</u>

<u>Figure 34 ³⁹K spin-echo MAS NMR spectra for the</u> <u>potassium aluminosilicates. All spectra were</u> <u>collected at 18.8 T using a spinning speed of ca. 15</u> <u>kHz. The dashed line at 0 ppm is included for</u> <u>reference</u>

<u>Figure 35 Experimental ³⁹K MAS NMR spectra</u> <u>collected at 19.6 T. A spinning speed of 10 kHz was</u> used for G1 and G2 and a spinning speed of 8 kHz was used for G3, K(2'-AMP) and K(5'-ADP)

Figure 36 Experimental ³⁹K 1D MAS and 2D MQMAS NMR spectra of (a) KPic, (b) KGP, and (c) G4 collected at 19.6 T. The spinning speed was 8 kHz in all experiments and pure absorptive 2D spectra were collected by combing SPAM with the shifted-echo method. The t_1 increment in the MQMAS experiment was rotor synchronized. The sharp signal at 0 ppm in (a) results from trace amounts of micro domains within the solid

<u>Figure 37 A comparison of (a) ¹H-³⁹K BRAIN-</u> <u>CP/WURST-CPMG and (b) direct excitation ³⁹K DFS-</u> <u>QCPMG NMR spectra of potassocene (CpK) acquired</u> <u>at 9.4 T. The former took 128 scans and 10 min total</u> <u>acquisition time. The latter took 18 000 scans and 25</u> <u>h total acquisition time (Schurko, 2015, unpublished</u> <u>data)</u>

<u>Figure 38 ²⁵Mg MAS NMR spectra of synthetic</u> <u>hydrotalcite acquired at 11.7 T. (a) unheated and</u> <u>heated to (b) 200 °C; (c) 300 °C; (d) 400 °C; (e) 600</u> <u>°C; and (f) 1200 °C</u>

<u>Figure 39 Natural-abundance ²⁵Mg SPAM-3QMAS</u> <u>NMR spectrum of MgAl-25-NO₃=. The dashed lines</u> <u>show the anisotropic slices of the three distinct Mg</u> <u>sites</u>

<u>Figure 40 Static ²⁵Mg SSNMR spectra of CPO-27-Mg</u> as a function of rehydration degree. All spectra were acquired under identical experimental conditions. The * indicates a small amount of impurity likely resulting from magnesium oxide <u>Figure 41 7QMAS ⁴³Ca NMR spectrum of CaSiO₃</u> <u>glass acquired at 21.8 T</u>

Figure 42 (a) Simulated and (b) experimental 43 Ca MAS SSNMR spectra of CaCrO₄ collected with a spinning speed of 4 kHz. (c) Simulated and (d) experimental 43 Ca static SSNMR spectra of CaCrO₄. Acquisition times of 18.9 and 5.2 h were required for (b) and (d), respectively, and all spectra were collected at 21.1 T

<u>Figure 43 ¹H{⁴³Ca} R³-HMQC spectra of HA and</u> <u>tHA. A spinning speed of 8 kHz was used. The ¹H</u> (black) and ⁴³Ca (red dashed) MAS NMR spectra are projected in the 2D spectra

Figure 44 ⁹Be MAS NMR spectra of (a) Cp_2Be , (b) $Cp*_2Be$, and (c) $(C_5Me_4H)_2Be$ at 9.4 T, along with accompanying spectral simulations

Cation-π Interactions

Figure 1 Prototypical cation–π interactions of alkali metal cations with benzene. Optimized distances above the ring center and gas-phase interaction energies (E_{int} , in kcal mol⁼¹) are based on highaccuracy gas-phase computations from Sherrill and coworkers¹

<u>Figure 2 Molecular electrostatic potential (MEP)</u> plots of benzene, toluene, benzonitrile, and triazine. In these plots, the electrostatic potential is shown on an electron density isosurface ($\rho = 0.001 \text{ e bohr}^{-3}$)

<u>Figure 3 DFT computed interaction energies, relative</u> to the unsubstituted case (X = H), for model <u>complexes of Na[±] above the center of</u> <u>monosubstituted benzenes, C₆H₅X, vs (a) the</u> electrostatic potential computed at the position of the Na^{\pm} above the ring and (b) the Hammett $\sigma_{\rm m}$ constant for the substituent

Figure 4 Competing models of substituent effects in cation–π interactions. (a) In conventional views, substituent effects arise from the modulation of the aromatic π-electron density by the substituents. (b) In the local dipole model of Wheeler and Houk,²⁹ substituent effects in cation–π interactions are dominated by the through-space electrostatic interaction of the local dipole associated with the substituent and the cation. Both models tend to provide similar predictions with regard to substituent effect trends

Figure 5 The electrostatic potential in the plane bisecting benzonitrile (left), as well as an additive approximation of this electrostatic potential constructed by adding the electrostatic potential in the plane bisecting benzene to the electrostatic potential of HCN

Figure 6 Accurate gas-phase interaction energy of <u>Na[±]</u> with benzene (gray line) and benzonitrile (black <u>line</u>) as a function of distance above the ring. Also <u>shown are a simple charge-dipole interaction (red</u> <u>line</u>) and the result of adding this charge-dipole term to the Na[±]...benzene interaction potential (red <u>dashed line</u>)

<u>Figure 7 Structure of the T1 lipase studied by</u> <u>Matsumura *et al.*,¹¹⁸ which exhibits a well-defined</u> <u>cation-π interaction between Na[±] and a nearby Phe</u> <u>residue (PDB: 2DSN)</u>

Ion Channels and Ionophores

<u>Figure x.1 Natural transmembrane transport</u> <u>mechanisms: (from left) protein-based ion channel,</u> <u>barrel stave, helical peptide dimerization, and</u> <u>ionophore</u>

<u>Figure 2 Artificial transmembrane transport</u> <u>mechanisms: (from left) amphiphilic ion pair, channel-</u> <u>forming peptide, membrane-spanning molecule,</u> <u>barrel-hoop channel, and ionophore</u>

<u>Figure 3 Examples of polyether ionophores: (a)</u> <u>monensin, (b) nonactin, (c) salinomycin, (d)</u> <u>calcimycin, (e) ionomycin, (f) nigericin, and (g)</u> <u>lasalocid</u>

Figure 4 X-ray structures of polyether ionophore cation complexes with solvent molecules removed for clarity: (a) (Li·monensin) (CSD ID: DOPVEJ), (b) (Na·monensin) (CSD ID: BELDAX), (c) (K·monensin) (CSD ID: FECROU10), (d) (Na·nonactin) (CSD ID: NONACS), (e) (K·nonactin) (CSD ID: NONKCS), (f) (Ca·nonactin) (CSD ID: CAXHEO), (g) (Na·salinomycin) (CSD ID: GIZDIC), (h) (Mg·(calcimycin)₂) (CSD ID: DUJWOU), (i) (Ca·ionomycin) (CSD ID: IONCAH01), (j) (Na·nigericin) (CSD ID: NIGERI), (k) (Na·lasalocid) (CSD ID: NALASC), (l) (Ba·(lasalocid)₂) (CSD ID: KAFZIA); CSD, Cambridge Structural Database

<u>Figure 5 Cyclopeptide ionophores: (a) valinomycin,</u> (b) enniatin A, and (c) beauvericin

Figure 6 X-ray structures of cyclopeptide ionophore cation complexes with counterions removed for clarity: (a) [K·valinomycin][±] (CSD ID: TEFBAH), (b) [K·(enniatin B)₂][±] (CSD ID: PEKFEQ), and (c) [(Ba·beauvericin)₂]^{4±} (CSD ID: BEAVBA); CSD, Cambridge Structural Database Figure 7 Examples of artificial ionophores

Figure 8 Top and side views of natural transmembrane channel-forming proteins (not to scale): (a) ASIC (PDB ID: 2QTS), (b) KcsA (PDB ID: 1BL8), (c) CavAb (PDB ID: 4MS2) (d) NavAb (PDB ID: 3RVY), and (e) TmCorA (PDB ID: 4EED); PDB = Brookhaven Protein Data Bank

<u>Figure 9 Examples of channel-forming biomolecules:</u> (from top) amphotericin, nystatin, alamethicin, and magainin

<u>Figure 10 Peptide-based artificial transmembrane</u> <u>channel-forming molecules</u>

<u>Figure 11 Examples of nonpeptide-based artificial</u> <u>channel-forming molecules</u>

Aryllithiums and Hetaryllithiums: Generation and Reactivity

<u>Figure 1 *n*-BuLi/anisole complexation illustrating</u> <u>electron withdrawal</u>

<u>Figure 2 Early transition state depiction for ortho-</u> <u>metalation of anisole (metal, M = Li)²⁵</u>

Figure 3 Ether (e)-stabilized structures of *n*-BuLi, for the tetramer (1), for the dimer (2), and for the bischelating amine-stabilized structure of the dimer (3)

Figure 4 Bis-chelating amines used to accelerate metalation reactions: diazobicyclo[2,2,2]octane (DABCO), tetramethylethylenediamine (TMEDA), pentamethyldiethylenetriamine (PMDTA), and hexamethylphosphorotriamide (HMPTA)

Figure 5 Proposed intermediate for ortho-lithiation⁶⁴

<u>Figure 6 The "pair of tweezers" representation for</u> <u>the cooperative 2-metalation of 1,3-DMB⁸⁵</u> <u>Figure 7 Preferred sites of lithiation of five-</u> <u>membered ring heterocycles (h are the heteroatoms</u> <u>O, S, or NMe)</u>

<u>Figure 8 Nearly linear structure for the X/Li</u> <u>intermediate</u>

Magnesium and Calcium Complexes in Homogeneous Catalysis

<u>Figure 1 Increasing popularity of molecular alkaline</u> <u>earth catalysis. (a) Number of publications per year;</u> (b) Number of citations per year

<u>Scheme 1 General stoichiometric reactivity of</u> <u>alkaline earth complexes</u>

<u>Figure 2 Standard catalytic cycles observed in Group</u> <u>2-mediated reactions</u>

<u>Scheme 2 Homoleptic and heteroleptic benzylcalcium</u> <u>precatalysts for styrene polymerization</u>

<u>Scheme 3 Dissociative mechanism of carbanionic</u> <u>polymer chain end inversion during calcium-catalyzed</u> <u>styrene polymerization</u>

<u>Figure 3 Possible microstructure outcomes for lactide</u> <u>ROP</u>

Scheme 4 Mechanisms of lactide ROP

<u>Figure 4 Selection of highly efficient heteroleptic</u> <u>magnesium and calcium precatalysts for lactide ROP</u>

<u>Scheme 5 Influence of chelate-resting state on</u> <u>catalytic activity toward LA and e-CL</u>

<u>Figure 5 Tischenko reaction with homoleptic and</u> <u>heteroleptic Group 2 amides and proposed</u> <u>mechanism</u>