

Surinder Kumar Gupta *Editor*

# Biotechnology of Crucifers

 Springer

# Biotechnology of Crucifers



Surinder Kumar Gupta  
Editor

# Biotechnology of Crucifers

 Springer

*Editor*

Surinder Kumar Gupta

Division of Plant Breeding and Genetics

S.K. University of Agricultural Sciences and Technology

Chatha, India

ISBN 978-1-4614-7794-5

ISBN 978-1-4614-7795-2 (eBook)

DOI 10.1007/978-1-4614-7795-2

Springer New York Heidelberg Dordrecht London

Library of Congress Control Number: 2013941733

© Springer Science+Business Media, LLC 2013

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed. Exempted from this legal reservation are brief excerpts in connection with reviews or scholarly analysis or material supplied specifically for the purpose of being entered and executed on a computer system, for exclusive use by the purchaser of the work. Duplication of this publication or parts thereof is permitted only under the provisions of the Copyright Law of the Publisher's location, in its current version, and permission for use must always be obtained from Springer. Permissions for use may be obtained through RightsLink at the Copyright Clearance Center. Violations are liable to prosecution under the respective Copyright Law.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

While the advice and information in this book are believed to be true and accurate at the date of publication, neither the authors nor the editors nor the publisher can accept any legal responsibility for any errors or omissions that may be made. The publisher makes no warranty, express or implied, with respect to the material contained herein.

Printed on acid-free paper

Springer is part of Springer Science+Business Media ([www.springer.com](http://www.springer.com))

# Preface

Despite the recent advances made in the improvement of crucifer crops using conventional breeding techniques, the yield levels and the oil and meal quality that were expected could not be achieved. The understanding of genetic material (DNA/RNA) and its manipulation by scientists have provided the opportunity to improve crucifers by increasing their diversity beyond conventional genetic limitations. The application of biotechnological techniques will have two major benefits: first, it allows to choose from a number of techniques/methods for appropriate selection of favorable variants, and second, it gives an opportunity to utilize alien variation available in the crucifers to develop high-yielding varieties with good nutritional quality and resistance to insects, pests, and diseases.

Realizing the importance of biotechnology, there is an urgent need to update current techniques for enhancing crucifer crop production at the global level. The editor approached the leading scientists of the world for write-ups on the advances made in the area of crucifer biotechnology to be packaged into one volume for the benefit of students, nutritionists, and biotechnologists as well as researchers engaged in the improvement of Brassicas. The book consists of 12 chapters. Chapter 1 deals with the importance, origin, and evolution of Brassicas, while Chaps. 2 and 3 describe the major advances made in cytogenetics at the molecular level and the introgression of genes from wild species. Chapter 4 deals with microspore culture and double haploid technology, while Chap. 5 describes phytoremediation in crucifers. These are followed by chapters on genome analysis (Chap. 6) and genetic engineering of lipid biosynthesis in seeds (Chap. 7). Metabolism and detoxification of crucifer phytoalexins, the molecular basis of cytoplasmic male sterility, and self-incompatibility have been discussed in detail in Chaps. 8, 9, and 10. Chapters 11 and 12 provide brief accounts of the molecular basis of hybrid technology and genetic modifications for pest resistance.

I am highly indebted to Prof. D. K. Arora, Honorable Vice-Chancellor, Sher-e-Kashmir University of Agricultural Sciences and Technology of Jammu, India, for encouraging me to carry out oilseed research work with all required facilities for the same.

I am indeed grateful to Prof. W. J. Zhou, Institute of Crop Science, Hangzhou, China, for providing crucial inputs and critically reviewing some of the chapters. Help rendered by Prof. Y. Takahata, Iwate University, Japan; Prof. Graham King, Southern Cross University, Australia; Prof. Randall Weselake, University of Alberta, Canada; Prof. M. S. C. Pedras, University of Saskatchewan, Canada; Prof. Takeshi Nishio, Tohoku University, Japan; and Prof. M. H. Fulekar, Central University of Gujarat, India, is gratefully acknowledged.

Joni Fraser, Developmental Editor for Springer Science + Business Media, deserves special thanks for her hard work in bringing this book to life. I owe a lot to my wife, Dr. Neena Gupta, for her support and patience during the preparation of this manuscript.

Chatha, India

Surinder Kumar Gupta

# Contents

<b>1</b>	<b>The Importance, Origin, and Evolution</b> .....	1
	Surinder Kumar Gupta	
<b>2</b>	<b>Molecular Cytogenetics</b> .....	13
	Annaliese S. Mason	
<b>3</b>	<b>Distant Hybridization Involving Different In Vitro Techniques</b> .....	23
	Dan Liu, Ling Xu, Xinxin Geng, Yuanfei Zhou, Zhenchao Zhang, Bing Wang, and Weijun Zhou	
<b>4</b>	<b>Microspore Culture and Doubled Haploid Technology</b> .....	45
	Yoshihito Takahata, Yu Takahashi, and Ryo Tsuwamoto	
<b>5</b>	<b>Biotechnological Strategies for Enhancing Phytoremediation</b> .....	63
	Bhawana Pathak, Razia Khan, Jyoti Fulekar, and M.H. Fulekar	
<b>6</b>	<b>Genome Analysis</b> .....	91
	Graham J. King	
<b>7</b>	<b>Genetic Engineering of Lipid Biosynthesis in Seeds</b> .....	111
	Stacy D. Singer, Michael S. Greer, Elzbieta Mietkiewska, Xue Pan, and Randall J. Weselake	
<b>8</b>	<b>Metabolism and Detoxification of Phytoalexins from Crucifers and Application to the Control of Fungal Plant Pathogens</b> .....	151
	M. Soledade C. Pedras	
<b>9</b>	<b>Molecular Basis of Cytoplasmic Male Sterility</b> .....	173
	Jinghua Yang and Mingfang Zhang	
<b>10</b>	<b>Self-Incompatibility</b> .....	187
	Hiroyasu Kitasiba and Takeshi Nishio	



**11 Hybrid Technology in Cruciferous Vegetables** ..... 209  
Muhammad Awais Ghani, Langlang Zhang, Junxing Li,  
Bin Liu, and Liping Chen

**12 Genetic Modifications for Pest Resistance** ..... 221  
Hongbo Liu, Bizeng Mao, Peng Cui, Tian Tian,  
Changrong Huang, Xi Xu, and Weijun Zhou

**Index** ..... 235

# Contributors

**Liping Chen, Ph.D.** Department of Horticulture, Zhejiang University, Hangzhou, Zhejiang Province, China

**Peng Cui, M.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Jyoti Fulekar, M.Phil.** School of Environment and Sustainable Development, Central University of Gujarat, Gandhinagar, Gujarat, India

**M.H. Fulekar, Ph.D.** School of Environment and Sustainable Development, Central University of Gujarat, Gandhinagar, Gujarat, India

**Xinxin Geng, B.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Muhammad Awais Ghani, M.S., M.D.** Department of Horticulture, Zhejiang University, Hangzhou, Zhejiang Province, China

**Michael S. Greer, M.Sc.** Agricultural, Food Nutritional Sciences, University of Alberta, Edmonton, Alberta, Canada

**Surinder Kumar Gupta, Ph.D.** Division of Plant Breeding and Genetics, S.K. University of Agricultural Science and Technology, Chatha, India

**Changrong Huang, M.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Razia Khan, M.Phil.** School of Environment and Sustainable Development, Central University of Gujarat, Gandhinagar, Gujarat, India

**Graham J. King, B.Sc., Ph.D.** Southern Cross Plant Science, Southern Cross University, Lismore, Australia

**Hiroyasu Kitasiba, Ph.D.** Graduate School of Agricultural Science, Tohoku University, Sendai, Miyagi, Japan

**Junxing Li, M.D.** Department of Horticulture, Zhejiang University, Hangzhou, Zhejiang Province, China

**Bin Liu, B.S.** Department of Horticulture, Zhejiang University, Hangzhou, Zhejiang Province, China

**Dan Liu, Ph.D.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

Institute of Tobacco Research, Chinese Academy of Agricultural Sciences, Qingdao, Shandong, China

**Hongbo Liu, Ph.D.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

College of Agricultural and Food Sciences, Zhejiang A&F University, Lin'an, Zhejiang, China

**Bizeng Mao, Ph.D.** Institute of Biotechnology, Zhejiang University, Hangzhou, China

**Annaliese S. Mason, Ph.D., B.Sc. (Hons)** School of Agriculture and Food Sciences, The University of Queensland, Brisbane, Australia

**Elzbieta Mietkiewska, Ph.D.** Agricultural, Food Nutritional Science, University of Alberta, Edmonton, Alberta, Canada

**Takeshi Nishio, Ph.D.** Graduate School of Agricultural Science, Tohoku University, Sendai, Miyagi, Japan

**Xue Pan, M.Sc.** Agricultural, Food Nutritional Science, University of Alberta, Edmonton, Alberta, Canada

**Bhawana Pathak, Ph.D.** School of Environment and Sustainable Development, Central University of Gujarat, Gandhinagar, Gujarat, India

**M. Soledade C. Pedras, Lic, Ph.D., D.Sc.** University of Saskatchewan, Saskatoon, Saskatchewan, Canada

**Stacy D. Singer, Ph.D.** Agricultural, Food Nutritional Science, University of Alberta, Edmonton, Alberta, Canada

**Yoshihito Takahata, Ph.D.** Iwate University, Ueda, Morioka, Iwate, Japan

**Yu Takahashi, Ph.D.** Iwate University, Ueda, Morioka, Iwate, Japan

**Tian Tian, B.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Ryo Tsuwamoto, Ph.D.** Misato Agricultural Extension Centre, Miyagi prefecture, Misato-machi, Toda-gun, Miyagi, Japan

**Bing Wang, M.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Randall J. Weselake, B.Sc., M.Sc., Ph.D.** Agricultural, Food Nutritional Science, University of Alberta, Edmonton, Alberta, Canada

**Ling Xu, Ph.D.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Xi Xu, B.S.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China

**Jinghua Yang, Ph.D.** Institute of Vegetable Science, Hangzhou, Zhejiang Province, China

**Langlang Zhang, B.S.** Department of Horticulture, Zhejiang University, Hangzhou, Zhejiang Province, China

**Mingfang Zhang, Ph.D.** Institute of Vegetable Science, Hangzhou, Zhejiang Province, China

**Zhenchao Zhang, Ph.D.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China  
Zhenjiang Agricultural Research Institute, Jurong; Jiangsu, China

**Weijun Zhou, Ph.D.** Institute of Crop Science and Zhejiang Key Laboratory of Crop Germplasm, Zhejiang University, Hangzhou, China  
Agricultural Experiment Station, Zhejiang University, Hangzhou, China

**Yuanfei Zhou, Ph.D.** Agricultural Experiment Station, Zhejiang University, Hangzhou, China

# Chapter 1

## The Importance, Origin, and Evolution

Surinder Kumar Gupta

**Abstract** The family Brassicaceae constitutes one of the world's most economically important group of plants which includes important vegetable oilseeds and condiment crops. Amongst the crucifer crops, rapeseed is the main source of fats and oil and shown an upward trend during the past 25 years (Kalia and Gupta 1997). Besides improvement in the nutritional profile of the Brassica oil and its meal, the conventional breeding as well modern biotechnological tools have led to the improvement of various agronomically important quantitative and qualitative characters. The nuclear restriction fragment length polymorphism technology has greatly aided in determining the degree of genetic variability among various Brassicas as well in studying their evolution pattern. The oldest references regarding origin and cultivation of rapeseed come from Asia, though the evolution of this crop took place in many countries throughout the globe. Lack of consistency in names, inclusion of too many forms in one species, and the entirely different forms of present day Brassicas from their ancestors make this genus a complex member of Brassicaceae and poses several taxonomic and classification problems. Still many attempts have been made to establish the origin of various Brassica species and their interrelationships through cytogenetic, chemotaxonomic, and molecular studies. The present chapter focuses on the importance origin and evolutionary developments in crucifers.

**Keywords** *Brassicaceae* • Rapeseed • *B. rapa* • *B. juncea* • *B. carinata* • Origin • Evolution • RFLP • Oilcrops

---

S.K. Gupta, Ph.D. (✉)  
Division of Plant Breeding and Genetics, S.K. University of Agricultural  
Science and Technology, Chatha 180009, India  
e-mail: guptaskpbg@rediffmail.com

## 1.1 Introduction

Brassicaceae are the world's third important source of vegetable oils after palm and soya bean (Beckman 2005) and contribute 14 % to the world's total vegetable oil pool. The production has shown a steady upward trend during the past 25 years. Brassica oilseed crops grow at relatively low temperature. In temperate regions of the world, oilseed rape (*B. napus*) and toria /turnip rape/Indian mustard are grown in subtropical parts of the Asia and is the main source of oil. The mode of reproduction varies from species to species. *B. napus*, *B. juncea* and *B. carinata* are predominantly self-pollinated, although they show some degree of cross-pollination ranging from 5 to 30 %, whereas, *B. rapa*, *B. oleracea* and *B. nigra* show cross-pollination due to sporophytic self-incompatibility. All the cultivated Brassica species are highly polymorphic including oilseed crops, root crops, and vegetables such as Chinese cabbage, Broccoli, and Brussels sprouts. These Brassica vegetables are dietary staple food in various parts of the world. However, our discussion in this chapter shall concentrate on the importance and origin of major species of Brassicaceae.

*B. napus* and *B. campestris* with both spring and winter type are grown in Canada and Europe. However, in countries like India and China, the production is also shared by other species, viz, *B. oleracea* and *B. juncea*. Rapeseed oil has gradually become important domestic and industrial oil in the western nations as a result of breeding for improved oil and meal quality and better processing techniques.

## 1.2 History

The family Brassicaceae contains over 338 genera and 3,709 species (Al-Shehbaz et al. 2006). The crop Brassicaceae have been very important as food crops in the form of vegetables, oilseeds, feed and fodder, green manure, and condiments and have played a great role in the human history by contributing a good share of food in one form or another. The Greek, Roman, and Chinese writings of 500–200 BC refer to rapiferous forms of *B. rapa* and also described their medicinal values (Downey and Robellen 1989).

Species grown as oilseed crops are *B. napus*, *B. juncea*, *B. rapa* and *B. carinata*. The vegetable Brassicaceae include *B. napus*, *B. rapa* (Chinese Cabbage, pak-choi, Chinese mustard, broccoli and kale); *B. oleracea* (cabbage, broccoli, cauliflower, Brussels sprouts, kale, etc.), *Raphanus sativus* and *Lepidium sativum*, *B. nigra* (black mustard), *B. juncea*, (brown mustard) and *Sinapis alba* are the main condiment of crops.

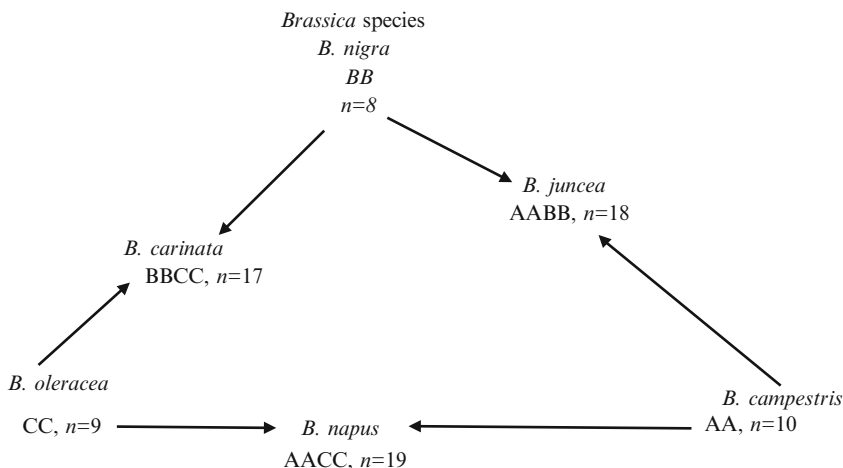
Early records indicate that Brassicaceae cultivated for several years in Asia. Seeds of *B. juncea* have been excavated from Chanhundaro, a site of Indus Valley civilization that existed in the plains of Punjab along the river of Indus ca 2300–1750 (Piggot 1950). Species from the genus Brassicaceae were in use and also in Gallia (Fussel 1955) and the seeds of the species had also been found in old German graves and Swiss constructions from the Bronze Age (Neuweiller 1905; Schiemann 1932;

Witmack 1904). In Dodoneus's "Herbalist" (1578), a mention has been made regarding the growing of *B. rapa* var. *rapifera* in 1470 as a winter crop. In his "Herball," Gerarde (1597) had very clearly differentiated between turnips (*B. rapa*) and navews (*B. napus*). Rape has been recorded as an oilseed crop in Europe at least since the Middle Ages., but it is still uncertain which species was cultivated (Appelquist and Ohlson 1972).

Domestication of rapeseed in Europe appears to have started in the early Middle Ages, although the true turnip was probably introduced by Romans since many other oil-yielding plants, particularly olive tree, were available in Southern Europe, *B. rapa* initially spread mainly as turnip rape crop within Europe. *B. rapa* had a wide distribution before the recorded history. Indian Sanskrit literature first mentions the plants about 1599 BC as Siddharth (Prakash 1961). Seed of both *B. rapa* and *juncea* were found in the archaeological excavation of ancient village Banpo, China, that existed in Neolithic times 6,000–7,000 years ago (Liu 1985). *B. nigra* (black mustard) is mentioned in Greek literature for its medicinal value. Ancient records indicated the cultivation rape seed was predominant during the thirteenth century. The rapeseed oil is used as major source of lamp oil and it was replaced by petroleum by the end of nineteenth century

A high quality of rapeseed named as Canola developed through genetic modification following the conventional plant breeding. Canola emerged in the 1970s as a viable oilseed, with high quality oil and meal for both human as well as livestock consumption (Shahidi 1990). Today, the fatty acid profile of Canola is considered as the most desirable, of all vegetable oil profiles by nutritionists (Stringam et al. 2003). The occurrence of two important components, glucosinolates and erucic acid were considered antinutritional for animal and humans, respectively. The high amount of glucosinolates in the meal still remained a major concern in the expansion of market of the vegetable oil derived from rapeseed. Prior to 1960, the erucic acid (a long chain fatty acid) content of rapeseed oil was not of particular interest while evaluating the oil use for edible purposes. The concern was felt by the European Economic Community (EEC) in 1960 with France, West Germany, Italy, the Netherland, Belgium, and Luxemburg as the founder members, for the development of low erucic acid varieties (less than 5 %). As a result, the traditional rapeseed oil started being considered as unsafe for human health. This led to the concentration of rapeseed breeding efforts toward the development of such varieties in late 1960s and early 1970. The application of gas liquid chromatography (Craig and Murphy 1959) led to the identification of low erucic acid plants in *B. napus* and *B. campestris* with the first low erucic acid plants in them identified in 1968 and the first *B. campestris* variety in 1971. In 1977, the cultivation of such varieties was made mandatory.

The oilseed Brassica has another important byproduct known as meal/cake. It is an excellent source of protein with a favourable balance of amino acids. However, its use was limited by its high glucosinolate content, which is a constituent of most of the plants of Brassicas. Traditional rapeseed varieties contained high levels of glucosinolates in the meal which when fed to livestock in sufficient quantities led to the problems related with nutrition, digestion, and thyroid. The development of fast



**Fig. 1.1** Evolution of cultivated Brassica species and its relatives (Nagaharu 1935)

and accurate chemical methods led to the identification of plants of the *B. napus* cultivar Bronowski from Poland, which was essentially free of the harmful glucosinolates normally found in rapeseed. The low glucosinolate genes were then incorporated in the well adapted and high yielding cultivars of *B. napus* and subsequently transferred to *B. campestris*.

The Brassicaceae family comprise of 25 tribes (Al-Shehbaz et al. 2006). The tribe Brassiceae contains genus Brassica and its wild relatives. It comprised of 48 genera and approximate 240 species (Warwick and Hall 2009). Schulz (1919, 1936) established the basic taxonomic classification and he recognized ten sub-tribes whereas Gómez-Campo (1999) recommended 9 subtribes. The subtribes Brassicinae, Moricandiinae and Raphaninae are of the great relevance to the scientists who are working on the Brassica species. The relationship among the species viz., *Brassica*, *Sinapis*, *Diplotaxis*, *Erucastrum*, *Herschfeldia*, *Eruca* and *Raphanus*. These sub tribes have been studied by Prakash and Hinata (1980) and Takahata and Hinata (1983, 1986).

Further more during the domestication, man has modified the entire plant and the present day Brassicas are entirely different from their ancestors. Also the occurrence of similar plant forms in more than one Brassica species resulted in considerable confusion and misclassification by early botanists (Downey and Robellen 1989). The cytogenetic relationships between the Brassica species as well as their closest allies were first explained systematically by Nagaharu (1935) about 70 years ago (Fig. 1.1) These relationships show that *B. campestris* ( $2n=20$ , AA), *B. nigra* ( $2n=16$ , BB) and *B. oleracea* ( $2n=18$ , CC) are the primary species and *B. napus* ( $2n=38$ , AACC), *B. carinata* ( $2n=34$ , BBCC) and *B. juncea* ( $2n=36$ , AABB) are the amphidiploids resulting from paired crossings between the primary species. Morinaga (1928, 1929a, b, 1934a, b) discussed that crop Brassicas include six cytodemes, three elementary ones with 16, 18 and 20 chromosomes as diploid and



three with higher chromosomes number of 34, 36 and 38 as tetraploid, the latter having evolved through interspecific hybridization in nature between any two of the elementary taxa. Herberd (1972, 1976) defined coeno species and on the basis of their chromosome number, they have been classified into 43 diploid and 13 tetraploid cytodesmes (Warwick and Black 1993).

Morinaga and his associates carried extensive cytogenetic studies in oilseed Brassicas and clarified the relationships between them (Prakash and Hinata 1980). According to the hypothesis of Morinaga and his student Nagaharu (1935), the three species with the higher chromosome number, *B. napus* L, *B. juncea* L, Czern and Coss, and *B. carinata* A. Braun, are amphidiploids combining in pairs the chromosome sets of the low chromosome number species *B. nigra*, *B. oleracea* and *B. rapa*. Nagaharu (1935) verified the hypothesis with successful resynthesis of *B. napus*. Resynthesis of *B. juncea* and *B. carinata* was accomplished by Frandsen (1943, 1947). Further verification of these species relationships were obtained from the studies on phenolic compound (Dass and Nybom 1967), protein pattern (Vaughan 1977), isozymes (Coulthart and Denford 1982; Chen et al. 1989), nuclear DNA and RFLP (Song et al. 1988a, b), molecular analysis of nuclear and chloroplast DNA and fluorescence in situ hybridization (Snowdon 2007; Warwick and Sauder 2005; Lysak et al. 2005). Robellen (1960) suggested that the low chromosome number species might have developed from the ancestral species, which could have even lower chromosome numbers. Also the chromosome analysis of the monogenomic species revealed that only six chromosomes were distinctly different, the remaining being homologous with one or another of the basic set of six.

Olsson (1954) suggested that all the 20 chromosome forms of leafy, *oleiferous*, and *rapiferous* Brassicas should be grouped into one species *B. campestris*. This was in support of the views of Howard (1940) that the name *B. campestris* should be reserved for the forms with  $2n=20$  and *napus* for the forms with  $2n=38$ . He proposed that the name sarson and toria should be *B. campestris* L. var. sarson and *B. campestris* L. var. toria, respectively. Singh (1958) considered yellow and brown sarson as varieties whereas Prakash (1973) considered them as the form of subspp. *oleifera*. Toxopens et al. (1984) suggested a classification and nomenclature of *B. campestris* should be changed to *B. rapa*.

### 1.2.1 *B. rapa*

The name *B. rapa* was mentioned as annual weed by Linnaeus (1753) in “Species Planatarum”. It was described as a plant with rough, stiff hairs when young, and just like *B. rapa* by DeCandolle et al. (1824). However, when it was realized that *B. campestris* and the turnip rape *B. rapa* have been classified as same species, a confusion was created in nomenclature and the wild type was often subordinated under *B. rapa* (Reiner et al. 1995). *B. rapa* subspp. *campestris* (formerly subspp. *sylvestris*), the wild form of *oleifera* is morphologically indistinguishable from the cultivated spring oilseed rapa. *B. rapa* subspp. *dichotoma* commonly referred as

toria, is an oilseed crop grown in Indian subcontinent. The yellow sarson and brown sarson (*B. rapa* subsp. *trilocularis*) are also grown in this continent.

*B. rapa* is thought to have originated in the mountainous areas near the Mediterranean sea (Tsunoda 1980). The original progenitor of the Indian and European forms was the same and that the Indian brown sarson evolved in the northwest of the Indian subcontinent from the original stock as suggested by the Russian workers, (Sinskaia 1928; Vavilov and Bukinich 1929), who regarded India as one of the independent centers of origin. The species appears to have attained a wide distribution throughout the Europe, parts of Africa, Asia and the Indian subcontinent before the recorded history. As *B. rapa* was most intensively grown at that time, it can be concluded that this crop was the major source of producing large quantities of vegetable oils. Seeds of *B. rapa* were first recorded in Europe in 1620 by the Swiss botanist Casper Bahhin. However, Boswell (1949) was of the view that these existed much earlier than this. As per some anonymous authors, rapeseed was grown in Europe as early as in the thirteenth century. Prakash and Hinata (1980) also suggested that oleiferous *B. campestris* subspecies developed in two places giving rise to two different races, one European and other Asian. There is a lot of evidence that European oilseed type *B. rapa* must be very close to the turnip type *B. rapa* genetically because it was produced out of it only some 100 years ago. On other hand in China Lintao Caizi very well known to the world as *B. chinensis* (leafy type *B. rapa* n=10) is used as oilseed crop. This can interpreted as parallel to the evolution of the oilseed type of the turnip type *B. rapa* in Europe (Sun et al. 1991). Alam (1945) concluded that sarson and toria types of *B. rapa* grown as oilseed crops in India and Pakistan evolved in Afghan that Persian area and migrated South India and further East. Song et al. (1988a, b) studied the phylogenetic analysis of 17 cultivated and 5 wild population of *B. rapa*. All the 17 cultivated forms were designated into two distinct groups as European and East Asia group. The phylogenetic grouping seems to correspond with the respective geographic distribution of the cultivated and wild forms of Brassica.

### 1.2.2 *B. napus*

*B. napus* is an amphidiploid resulting from the cross between the plants of *B. oleracea* and *B. rapa* and is comparatively of recent origin (Olsson 1960). It is uncertain to maintain if *B. napus* is found wild or not, since wild forms of this crop are difficult to find (Hinata and Prakash 1984). However, if wild *napus* exists, it must be a European- Mediterranean species that originated in the area of overlap between *B. oleracea* and *B. campestris*. Olsson (1960) suggested that *B. napus* could have arisen several times by spontaneous hybridization between the different forms of *B. rapa* and *B. oleracea*. Song and Osborn (1992) on the basis of chloroplast and mitochondrial DNA analysis suggested that *B. montana* (n=9) might be closely related ancestral species of *B. rapa* and *B. montana* was the maternal donor. The parental origins of *B. napus* were also investigated using six microsatellite markers located in the chloroplast genome by Allender et al. (2005). Allender and king (2010) using chloroplast and nuclear markers concluded that it is highly unlikely

that *B. oleracea* or any of the C- genome species are closely related to the maternal progenitor of most *B. napus* accession. They also suggested that either of *B. rapa* and *montana* or a common ancestor could have been the maternal parent of *B. napus*. Though, they suggested that *B. oleracea* was not the parent of most of *B. napus* accessions, a small number of accessions shared *B. oleracea* haplotype. Similarly, the phylogenetic analysis based on nuclear RFLP data also suggests that *B. napus* has multiple origins (Song et al. 1990, 1993). The various cytoplasm types found in *B. napus* accessions correspond to the progenitor diploid species which provide a strong evidence for the multiple origins of this crop (Song et al. 1997).

In *B. napus* as well as *B. campestris*, a range of morphological forms are found both having annual and biennial types. Keeping this in view, Olsson (1960) suggested that *B. napus* could have arisen several times by spontaneous hybridization of different forms of *B. campestris* and *B. oleracea*. The majority of the cultivated *B. napus* accessions appear to have arisen by an interspecific cross in which a wild nine or ten chromosome species having the *B. montana* cytoplasm type.

Mizushima and Tsunoda (1967) inferred that *B. napus* was found in the coast of northern Europe because *B. oleracea* extended its territory up to northern Europe from the Irano Turanean regions with its high adaptability to low temperature. Sinskaia (1928) and Schiemann (1932) were also of the view that it might have originated in the Mediterranean region or in the western or northern Europe. In Europe, production of oleiferous, *B. napus* might have started during the middle Ages. In Asia, it was introduced during the nineteenth century. The Chinese and Japanese germplasm was developed by crossing European *B. napus* cultivars with indigenous *B. rapa* cultivars (Shiga 1970). Today most of the oilseed rape produced in China, Korea and Japan is harvested from *B. napus* cultivars. It is less adapted to the Indian sub continent due to the short days and warm growing conditions.

### 1.2.3 *B. juncea*

*B. juncea* is an amphidiploid and results from an interspecific cross between the plants of *B. rapa* and *B. nigra* and it has longer history than *B. napus*. A number of workers have suggested that China as the centre of origin where the maximum diversity is found (Prain 1898; Sinskaia 1928; Vavilov 1949). It came to India from China through a North Eastern route and its immigration to India has been independent of an Aryan incursion (Prain 1898). According to Sun (1970), *B. juncea* originated in Middle East from where it spread to Asia. Afghanistan is thought to be as secondary centre of origin (Olsson 1960; Mizushima and Tsunoda 1967; Tsunoda and Nishi 1968) from where it spread to secondary centre on the Indian sub continent as a major oilseed crops (Hemingway 1995; Prakash and Hinata 1980). The analysis of Fraction I protein data (Uchimiya and Wildman 1978) and chloroplast DNA established the fact that *B. rapa* served as female parent in the formation of the species (Erickson et al. 1983; Palmer et al. 1983; Palmer 1988; Song et al. 1988a, b; Warwick and Black 1991; Yang et al. 2002). Qi et al. (2007) reported that some phenotypes may have evolved with *B. nigra* as maternal parent as evidenced

from the investigation on nuclear Internal Transcribe Spacer (ITS) regions of ribosomal DNA from 15 different Chinese vegetables and one oilseed form. Wu et al. (2009) studied the relationship among 95 *B. juncea* accessions originated from China, India, Pakistan and Japan following the sequenced Related Amplified Polymorphisms (SRAPs). Although winter sown accessions exhibited more genetic diversity than the spring sown accessions yet, SRAP markers did not provide clear cut separation between Indian/Pak and China winter sown mustard. Data supporting the polyphyletic origin are parallel variation observed a nuclear RFLP pattern of *B. campestris* and *B. juncea* (Song et al. 1988a, b). Wu et al. (2009) and Qi et al. (2007) also supported the idea that vegetables and oilseed forms have polyphyletic origin and evolved separately during the course of evolution.

#### 1.2.4 *B. carinata*

*B. carinata* is commonly known as Abyssinian or Ethiopian mustard and it is indigenous Brassica oilseed and vegetable crop in Ethiopia. It is also an amphidiploid species derived from two parental species *B. nigra* as a female and *B. oleracea* a male parent (Uchimiya and Wildman 1978; Palmer et al. 1983; Song et al. 1988b; Erickson et al. 1983). Quiros et al. (1988) suggested on the basis DNA analysis that *B. carinata* is an amphidiploid of the recent origin and may have the multiple origin. Song et al. (1988a, b) also confirmed on the basis of RFLP study that *B. carinata* came from *B. nigra* and *B. oleracea*. This species is a new introduction to India however it is being bred for potential commercial production in Spain, Canada, India and Australia.

#### 1.2.5 *B. nigra*

*B. nigra* is an ancient crop which finds mention in the Sanskrit Upnisdas as a Sarshap (Prakash 1961). Hemingway (1995) placed it in Irano-Turanian, Saharo-Sindian region This species became wide spread in old world probably having its origin in Asia minor. The distribution in Europe, Mediterranean and Ethiopian plateau (Bailey 1930; Schulz 1919; Mizushima and Tsunoda 1967) suggest that *B. nigra* originated in central and Southern Europe.

### 1.3 Conclusion

Brassica have a range of morphotypes and accordingly vary in their origin, cultivation, use, and history. The evolution of each species of Brassica has witnessed a shift in their morphophysiological traits from their original form to present day cultivars. Canola is one of the examples in rapeseed. In *B. oleracea* present day cultivars have resulted from mutation followed by adaptation and selection.

## References

- Alam Z (1945) Nomenclature of oleiferous Brassicas cultivated in Punjab. *Indian J Agric Sci* 15:173–181
- Allender CJ, King GJ (2010) Origins of the amphiploid species *Brassica napus* L. investigated by chloroplast and nuclear molecular markers. *BMC Plant Biol* 10:54
- Allender C, Evered C, Lynn J, Graham K (2005) Tracing the origins of *Brassica napus* using chloroplast microsatellites. In: Proceedings of plant and animal genomes XIII conference, 15–19 Jan 2005. Town & country convention center, San Diego, p 411
- Al-Shehbaz LA, Beilstein MA, Kellog EA (2006) Systematics and phylogeny of the Brassicaceae (Cruciferae): an overview. *Plant Systemat Evol* 259:89–130
- Appelquist LA, Ohlson R (1972) Rapeseed: cultivation, composition, processing and utilization. Elsevier, Amsterdam/London/New York
- Bailey LH (1930) The cultivated Brassicas: second paper. *Gentes Herb* 2:211–267
- Beckman C (2005) Vegetable oils: competition in a changing market. *Bi-weekly Bulletin. Agriculture and Agri-Food Canada* 18(11). Available at [http://www.agr.gc.ca/mad-dam/e/bulletine/v18e/v18n11\\_e.htm](http://www.agr.gc.ca/mad-dam/e/bulletine/v18e/v18n11_e.htm)
- Boswell VR (1949) Our vegetable travelers. *Natl Geogr Mag* 96:145–217
- Chen BY, Heneen WK, Simonsen V (1989) Comparative and genetic studies of isozymes in resynthesized and cultivated *Brassica napus* L., *B. campestris* L. and *B. alboglabra* Bailey. *Theor Appl Genet* 77:673–679
- Coulthart MB, Denford KE (1982) Isozyme studies in *Brassica*. I lectrophoretic techniques for leaf enzymes and Comparison of *B. napus*, *B. campristris* and *B. oleracea* using phosphoglucomutase. *Can J Plant Sci* 62:621–630
- Craig BM, Murphy NL (1959) Quantitative fatty acid analysis of vegetable oil by gas liquid chromatography. *J Am Oil Chem Soc* 36:549–552
- Dass H, Nybom N (1967) The relationships between *Brassica nigra*, *B. campestris*, *B. oleracea*, and their amphidiploid hybrids studied by means of numerical chemotaxonomy. *Can J Genet Cytol* 9:880–890
- DeCandolle AP (1824) Translated into German by Berg, C.W.1824. Die verschiedenen Arten. Unterarten und Spielarten des Kohls und der Rettige, Welche in Europa gebauet warden, Leipzig
- Dodoneus R (1578) A nievve Herball. Antwerp. Translated by H. Lyte, London
- Downey RK, Robellen G (1989) Brassica species. In: Robellen G, Downey RK, Ashri A (eds) *Oil crops of the world*. McGraw Hill, New York, pp 339–362
- Erickson LR, Straus NA, Beversdorf WD (1983) Restriction patterns reveal origins of chloroplast genomes in *Brassica amphidiploids*. *Theor Appl Genet* 65:201–206
- Frandsen KJ (1943) The experimental formation of *B. juncea*. *Dansk Bot. Archiv* 11:1–17
- Frandsen KJ (1947) The experimental formation of *Brassica napus* L. va. *Oleifera* DC. and *Brassica carinata* Braun. *Dansk Bot. Arkiv* 12:1–16
- Fussel GE (1955) History of cole (*Brassica* sp). *Nature* 176:48–51
- Gerarde J (1597) Herball or generall historie of plantes. Norton, J, London
- Gómez-Campo C (1999) Taxonomy. In: Gómez-Campo C (ed) *Biology of Brassica coenospecies*. Elsevier, Amsterdam, pp 3–32
- Herberd DJ (1972) A contribution to the cytotaxonomy of Brassica (cruciferae) and its allies. *Bot J Linn Soc* 65:1–23
- Herberd DJ (1976) Cytotaxonomic studies of *Brassica* and related genera. In: Vaughan JG et al (eds) *The biology and chemistry of the cruciferae*. Academic, London, pp 47–68
- Hedge IC (1976) A systematic and geographical survey of the world cruciferae. In: Vaughan JG, McLeodand AJ, Jones BMG (eds) *The biology and chemistry of cruciferae*. Academic, New York, pp 1–45
- Hemingway JS (1995) Mustards: *Brassica spp.* and *Sinapis alba* (Cruciferae). In: Smartt J, Simmons NW (eds) *Evolution of crop plants*. Longman, London, pp 82–86

- Hinata K, Prakash S (1984) Ethnobotany and evolutionary origin of Indian oleiferous Brassicaceae. *Indian J Genet* 44:102–112
- Holzner W (1981) Acker-Unkraut-Bestimmung, Verbreitung, Biologie und Ökologie. Leopold Stocker Verlag, Graz/Stuttgart
- Howard HW (1940) Nomenclature of Brassica species. *Curr Sci* 9:494–495
- Kalia HR, Gupta SK (1997) Importance, nomenclature and origin. In: Kalia HR, Gupta SK (eds) Recent advances in oilseed Brassicas. Kalyani, New Delhi, pp 1–11
- Linnaeus C (1973) *Species plantarum*. Holmiae (Stockholm) (Reprint London, 1957)
- Liu H (1985) Genetics and breeding of rapeseed. *Shanghai Sci. and Tech.*, Shanghai, p 592
- Lysak MA, Koch MA, Pecinka A, Schubert I (2005) Chromosome triplication found across the tribe Brassicaceae. *Genome Res* 15:516–525
- Mizushima U, Tsunoda S (1967) A plant exploration in Brassica and Allied Genera. *Tohoku J Agr Res* 17:249–276
- Morinaga T (1928) Preliminary note on interspecific hybridization in Brassica. *Proc Imper Acad Tokyo* 4:620–622
- Morinaga T (1929a) Interspecific hybridization in Brassica I. The cytology of F1 hybrids of *B. nepella* and various other species with 10 chromosomes. *Cytologia* 1:16–27
- Morinaga T (1929b) Interspecific hybridization in Brassica II. The cytology of F1 hybrids *B. cerna* and various other species with 10 chromosomes. *Jpn J Bot* 4:277–280
- Morinaga T (1934a) Interspecific hybridization in Brassica VI. The cytology of F1 hybrids of *B. juncea* and *B. nigra*. *Cytologia* 6:62–67
- Morinaga T (1934b) On the chromosome number of *Brassica juncea* and *Brassica napus* on the hybrid between these two and on cv spring of the hybrid. *Jpn J Genet* 9:161–163
- Nagaharu U (1935) Genome analysis in Brassica with special reference to the experimental formation of *Brassica napus* and peculiar mode of fertilization. *Jpn J Bot* 7:389–452
- Neuweiller E (1905) Die prahistorische pflanzenreste mitteleuropas. Albert Raustein, Zurich
- Olsson G (1954) Crosses within the campestris group of the genus Brassica. *Hereditas* 40:398–418
- Olsson G (1960) Species crosses within the genus Brassica II. Artificial *Brassica napus* L. *Hereditas* 46:351–386
- Palmer JD (1988) Intraspecific variation and multicircularity in *Brassica* mitochondrial DNAs. *Genetics* 118:341–351
- Palmer JD, Shields CR, Cohen DB, Orton TJ (1983) Chloroplast DNA evolution and the origin of amphiploid *Brassica* species. *Theor Appl Genet* 65:181–189
- Piggot S (1950) Prehistoric India to 1000 BC. Penguin Books, Harmondsworth
- Prain D (1898) The mustards cultivated in Bengal. *Agr Ledger* 5:1–80
- Prakash S (1961) Food and drinks in ancient India. Mitra R, Delhi, pp 265–266
- Prakash S (1973) Artificial Brassica juncea Coss. *Genetica* 44:249–263
- Prakash S, Hinata K (1980) Taxonomy, cytogenetics and origin of crop Brassica, a review. *Opera Botanica* 55:11–57
- Qi X, Zhang MF, Yang JH (2007) Molecular phylogeny of Chinese vegetable mustard (*Brassica juncea*) based on the Internal Transcribed Spacers (ITS) of nuclear ribosomal DNA. *Genet Res Crop Evol* 54:1709–1716
- Quiros CF, Ochoa O, Douches DS (1988) Exploring the role of  $x=7$  species in Brassica evolution: hybridization with *B. nigra* and *B. oleracea*. *J Hered* 79:351–358
- Reiner H, Holzner W, Ebermann R (1995) The development of turnip type and oilseed type Brassica rapa crops from the wild type in Europe—an overview of the botanical, historical and linguistic facts: rapeseed today and tomorrow. In: Proceedings of ninth international rapeseed congress, vol 4. Cambridge, 4–7 July 1995, pp 1066–1069
- Robellen G (1960) Beiträge zur analyse des Brassica-genoms. *Chromosoma* 11:205–228
- Schiemann E (1932) Entstehung der kulturpflanzen Handlab. *Verehrbis Lfg* 15
- Schulz OE (1919) IV. 105 Cruciferae-Brassicaceae. Part 1. Subtribes I. Brassicinae and II. Raphaninae. In: Engler A (ed) Das Pflanzenreich, Heft 68–70. Wilhelm Engelmann, Leipzig, pp 1–290

- Schulz OE (1936) Cruciferae. In: Engler A, Harms H (eds) Die Natürlichen Pflanzenfamilien, 2nd edn. 17B, Verlag von Wilhelm Engelmann, Leipzig, pp 227–658
- Shahidi F (1990) Rapeseed and canola: global production and distribution. In: Shahidi F (ed) Canola and rapeseed: production, chemistry, nutrition and processing technology. Van Nostrand Reinhold, New York, p 13
- Shiga T (1970) Rape breeding by interspecific crossing between *Brassica napus* and *Brassica campestris* in Japan. Jpn Agr Res Q 5:5–10
- Singh D (1958) Rape and mustard. The Indian Central Oilseeds Committee, Bombay
- Sinskaia EN (1928) The oleiferous plants and roots of the family cruciferae. Bull Appl Bot Genet Plant Breeding 10:1–648
- Snowdon RJ (2007) Cytogenetics and genome analysis in *Brassica* crops. Chromosome Res 15:85–95
- Song K, Osborn TC (1992) Polyphyletic origins of *Brassica napus*: new evidence based on organelle and nuclear RFLP analyses. Genome 35:992–1001
- Song KM, Osborn TC, Williams PH (1988a) Brassica taxonomy based on nuclear restriction fragment length polymorphisms (RFLPs). 1. Genome evolution of diploid and amphidiploid species. Theor Appl Genet 75:784–794
- Song KM, Osborn TC, Williams PH (1988b) Brassica taxonomy based on nuclear restriction length polymorphisms (RFLPs) 2. Preliminary analysis of subspecies with *B. rapa* (syn. *campestris*) and *B. oleracea*. Theor Appl Genet 76:593–600
- Song KM, Osborn TC, Williams PH (1990) Brassica taxonomy based on nuclear restriction length polymorphisms (RFLPs) 3. Genome relationships in Brassica and related genera and the origin of *B. oleracea* and *B. rapa*. Theor Appl Genet 79:497–506
- Song KM, Tang KL, Osborn TC (1993) Development of synthetic Brassica amphidiploids by reciprocal hybridization and comparison to natural amphidiploids. Theor Appl Genet 86:811–821
- Song KM, Osborn TC, Williams PH (1997) Taxonomy based on nuclear RFLP analysis. In: Kalia HR, Gupta SK (eds) Recent advances in oilseed brassicas. Kalyani, New Delhi, pp 12–24
- Stringam GR, Ripley VL, Love HK, Mitchell A (2003) Transgenic herbicide tolerant canola. The Canadian experience. Crop Sci 43:1590–1593
- Sun VG (1970) Breeding plants of Brassica. J Agr Assoc China 71:41–52
- Sun WC, Pan QY, An XH, Yang YP (1991) Brassica and Brassica related oilseed crops in Gansu, China. In: McGregor DI (ed) Proceedings GCIRC, eighth international rapeseed congress, vol 4. Saskatoon, pp 1130–1135
- Takahata Y, Hinata K (1983) Studies on cytodemes in subtribe Brassicinae (Cruciferae). Tohoku J Agri Res 33:111–124
- Takahata Y, Hinata K (1986) Consideration of the species relationships in subtribe Brassicinae (Cruciferae) in view of cluster analysis of morphological characters. Plant Species Biol 1:79–88
- Toxopous H, Oost EH, Reuling G (1984) Current aspects of the taxonomy of cultivated Brassica species. The use of *B. rapa* L. versus *B. campestris* L. and a proposal for a new intraspecific classification of *B. rapa* L. Cruciferae Newsl 9:55–58
- Tsunoda S (1980) Eco-physiology of wild and cultivated forms in *Brassica* and allied genera. In: Tsunoda S et al (eds) Brassica crops and wild allies. Japan Scientific Societies Press, Tokyo, pp 109–120
- Tsunoda S, Nishi S (1968) Origin, differentiation and breeding of cultivated *Brassica*. Proc XII Int Congr Genet 2:77–88
- Uchimiya H, Wildman SG (1978) Evolution of fraction I protein in relation to origin of amphidiploid Brassica species and other members of the Cruciferae. J Hered 69:299–303
- Vaughan JG (1977) A multidisciplinary study of the taxonomy and origin of Brassica crops. Bioscience 27:35–40
- Vavilov NI (1949) The origin, variation, immunity and breeding of cultivated plants. Chron Bot 13:1–364
- Vavilov NI, Bukinich DD (1929) Agriculture in Afghanistan. Bull Appl Bot Genet Plant Breeding 33:378–382

- Warwick SI, Black LD (1993) Molecular relationships in the subtribe Brassicinae (Cruciferae, Tribe, Brassiceae). *Can J Bot* 71:906–918
- Warwick SI, Black LD (1991) Molecular systematics of *Brassica* and allied Genera (subtribe Brassicinae, Brassiceae) – chloroplast genome and cytodeme Congruence, *Theor Appl Genet* 82:839–850
- Warwick SI, Hall JC (2009) Phylogeny of *Brassica* and wild relatives. In: Gupta SK (ed) *Biology and breeding of crucifers*. CRC, Boca Raton, pp 19–36
- Warwick SI, Sauder CA (2005) Phylogeny of tribe *Brassicaceae* (Brassicaceae) based on chloroplast restriction site polymorphisms and nuclear ribosomal internal transcribed spacer and chloroplast *trnL* intron sequences. *Can J Bot* 83:467–483
- Warwick SI, Black LD, Aguinalalde I (1992) Molecular systematics of Brassicaceae allied genera (subtribe Brassicinae, Brassiceae) – chloroplast DNA variation in the genus *diplotaxis*. *Theor Appl Genet* 83:839–850
- Witmack L (1904) Über die in Pompej gefundenen Pflanzenreste. *Englers Bot. Jahrb. Bd 33*
- Wu X, Chen B, Lu G, Wang H, Xu K, Guizhan G, Song Y (2009) Genetic diversity in oil and vegetable mustard (*Brassica juncea*) landraces by SRAP markers. *Genet Resour Crop Evol* 56:1011–1022
- Yang YW, Tai PY, Chen Y, Li WH (2002) A study of the phylogeny of *Brassica rapa*, *B. nigra*, *Raphanus sativa* and their related genera using noncoding regions of chloroplast DNA. *Mol Phylogenet Evol* 23:268–275



# Chapter 2

## Molecular Cytogenetics

Annaliese S. Mason

**Abstract** Cytogenetics has played a key role in the history of scientific research in the Brassicaceae since the start of the last century. The discovery of the *Brassica* “U’s Triangle” species, elucidation of phylogenetic relationships and investigations of chromosome evolution all contributed to building up the basic genomic understanding of the Brassicaceae we have today. The advent of molecular cytogenetics in this family in the last 20 years has led to a progressively greater understanding of the factors underlying chromosome dynamics and organisation, meiotic and mitotic mechanisms and cell division processes. In addition, linking molecular cytogenetics with other molecular techniques, such as marker studies, DNA sequencing and protein expression analysis, has bridged the gap between chromosomes and linkage groups, resulting in a wealth of new information in this family. Future prospects for molecular cytogenetics in the Brassicaceae are bright. The recent and imminent release of additional Brassicaceae genomes will greatly facilitate development of probes for fluorescent in situ hybridisation as well as a comprehensive understanding of gene expression and protein interactions during cell division.

**Keywords** Cytogenetics • Fluorescent in situ hybridisation • Chromosomes • *Brassica* • *Arabidopsis*

### 2.1 Introduction

Cytogenetics, literally “cell genetics”, traditionally refers to the study of chromosomes. Cytogenetics conventionally encompasses studies of chromosome number, structure and organisation, chromosomal aberrations and chromosome behaviour

---

A.S. Mason, Ph.D., B.Sc. (Hons) (✉)  
School of Agriculture and Food Sciences, The University of Queensland 4072,  
Brisbane, Australia  
e-mail: Annaliese.mason@uq.edu.au