Parvaiz Ahmad · M.M. Azooz M.N.V. Prasad *Editors* 

# Ecophysiology and Responses of Plants under Salt Stress



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## Foreword

Salinity imposes a serious problem to agriculture and biomass production. Several important crop species are sensitive to increased salinity which results in decreased photosynthetic performance and lower yield. Increased salinity of arable lands frequently is accompanying drought stress in arid and semiarid areas. On the other hand there are numerous species which not only tolerate increased salinity but they salty soils and waters as their natural environment. Therefore, the underneed standing molecular basis of salt stress and the mechanisms of its tolerance is of utmost importance. The volume entitled "Ecophysiology and Responses of Plants under Salt Stress" gives up-dated information about recent achievements and future trends in research on salt stress in plants. International team of authors contributed eighteen chapters on various aspects of salt stress in plants, dealing both with the ecological and physiological responses, biochemical mechanisms, processes of salt stress damage and factors contributing to its defense and tolerance. The volume edited by the group of competent Editors brings together recent results in the broad area of salt stress in plants and gives good overview of the recent literature. In my opinion the volume is well prepared and necessary on the market and it may be recommended to researchers and students working in the field of salt stress in plants.

Krakow, Poland

Kazimierz Strzałka

### Preface

Soil is the soul of life and is an invaluable natural resource. Healthy soil is essential for high-quality food production. Therefore, it is necessary to maintain soil fertility for sustainable crop production. Since the cultivable land is declining day by day because of urbanization, there is need to use the uncultivable land for food production to feed teeming population. Changing environment (biotic and abiotic stress) has a negative impact on the growth and development of the plants. Salinity stress is one of the major abiotic stresses that limit agricultural yield. Salinity is responsible for the induction of primary effects like ionic and osmotic stress, which in turn induce oxidative stress in plants.

In plants exposed to salt stress, all the major processes such as photosynthesis, protein synthesis, and energy and lipid metabolisms are affected. Reactive oxygen species (ROS) generated as a result of salt-imposed oxidative stress is highly deleterious for plants. ROS destroys the structure and functions of biomolecules such as membrane lipids, proteins and nucleic acids, and in higher concentrations causes death of the plant cells. Nonetheless, plants have mechanisms to counteract the deleterious effects of primary and secondary stresses through the generation of osmolytes or antioxidants.

This volume consists of 18 chapters which deal with the effect of salt stress on plants. Chapter 1 deals with the causes and types of salt stress and responses of plants. Chapter 2 describes how exogenous protectants help plants to withstand the negative effect of salt stress. Chapter 3 highlights the effect of salt stress on ion transport, water relations and oxidative damage in plants. Chapter 4 is about symbiotic coalition of lichens against salt stress. Chapter 5 deals with the changes in photosystem II under salt stress. Chapter 6 describes the effect of salt stress on the root system and its tolerance. Chapter 7 highlights the effect of salt stress on rice yield and the mechanism of salt tolerance in rice plants. Chapter 8 deals with aquaporins' activity, functions and their role in plant growth and development. Chapter 9 describes production of oil seed crops under salt stress and improving salt tolerance by different methods. Chapter 10 narrates the response of tomato plants to

salinity and the role of ABA in plants under salt stress. Chapter 11 deals with changes of phenolic compound content in various plants under salt stress. Chapters 12 and 13 highlight the role of polyamines in plants under salt stress. Chapter 14 describes the physiological functions of jasmonates in relation to environmental stress in plants. Chapter 15 deals with the role of nitric oxide in osmoregulation, ion homeostasis and signalling in plants under salt stress. Chapters 16 and 17 highlight the role and metabolism of nutrients under salt stress. Chapter 18 describes a case study of Neretva river valley about soil and water management for sustained agriculture in alluvial plains and flood plains exposed to salinity.

In this volume, we have tried to provide the readers a background for understanding salt stress and tolerance mechanisms in plants. We are thankful to all the authors for their valuable contributions. We are also thankful to several colleagues who helped us directly or indirectly in completing this volume. We appreciate Hanna Smith (Associate Editor, Springer) and Margaret Burns (Developmental Editor, Springer) for their prompt help, suggestions and punctuality in publication of this collective volume.

Srinagar, India Qena, Egypt Hyderabad, India Parvaiz Ahmad M.M. Azooz M.N.V. Prasad

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# **Chapter 1 Salt Stress: Causes, Types and Responses of Plants**

Saiema Rasool, Asiya Hameed, M.M. Azooz, Muneeb-u-Rehman, T.O. Siddiqi, and Parvaiz Ahmad

#### 1.1 Introduction

Abiotic factors like temperature, drought, salt stress, etc. results in the depletion of large number of food production in today's world and as a result these global changes have led to alarmist projections that seem to argue for additional strategies by which food supply can be guaranteed (Miflin 2000). Moreover, the increased productivity achieved in irrigated areas still do not benefit people because salinization following prolonged irrigation is unavoidable (Flowers and Yeo 1995; Postel 1999). These considerations have aroused strong interest in studying plant abiotic stress responses. Salinity has severely affected the agricultural productivity and the damaging effects of salt accumulation have influenced both ancient and modern civilizations. It is estimated that about 20% of the irrigated land in the present world is affected by salinity that is exclusively classified as arid and desert lands comprising

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25% of the total land of our planet (Yeo 1999). Saline soils with soluble salts affect plant growth at various stages leading to yield differences between crops and also the differences in their ion compositions at maturity. The loss of farmable land due to salinization directly affects the food requirement of the world population, which is projected to increase by 8.5 billion over the next 25 years. Since the cultivable land is declining day by day because of urbanization, there is need to use the uncultivable land for food production to feed teeming population. The maximum area of uncultivable land belongs to salt affected area. In developed and developing countries, the intensive use of agricultural practices has led to the degradations of farming land and water supplies. Preferably, use of crops that tolerate the high levels of salinity in the soils would be a practical contribution towards addressing the problem. Most crops tolerate salinity to a threshold level (Khan et al. 2006), a 'threshold' for a crop is defined as the value below which crop growth is generally not affected due to salinity.

According to Flowers et al. (1977) plants can be divided in to glycophytes and halophytes on the basis of their abilities to grow on different salt concentrations. Halophytes are the plants that grow and complete their life cycle on high concentration of salt, e.g. *Atriplex, Vesicaria*. Majority of the terrestrial plants including agricultural crops are glycophytic and cannot tolerate high concentration of salt. Plant growth and development is hampered due to salinity stress through: (1) low osmotic potential of soil solution (water stress), (2) nutritional imbalance, (3) specific ion effect (salt stress) or (4) a combination of these factors (Ashraf 1994). During the onset and development of salt stress within a plant, all the major processes such as photosynthesis, protein synthesis and energy and lipid metabolisms are affected.

Osmotic stress is caused due to the excess of Na<sup>+</sup> and Cl<sup>-</sup> in the environment that decrease the osmotic potential of the soil solution and hence water uptake by the plant root. During osmotic stress plant also accumulate low molecular mass compounds known as compatible solutes or osmolytes like, proline, protein, mannitol, sorbitol, glycine betaine, etc. Salt induced osmotic stress is responsible for the oxidative stress caused by reactive oxygen species (ROS).

ROS, such as singlet oxygen ( $_1O^2$ ), superoxide ions ( $O_2^-$ ) and peroxides, the most widely distributed being hydrogen peroxide ( $H_2O_2$ ) are toxic molecules (Apel and Hirt 2004; Triantaphylidès et al. 2008). ROS is capable of inducing damage to almost all cellular macromolecules including DNA (Jaleel et al. 2007a, b, c, 2008; Tuteja et al. 2009). ROS targets high-molecular mass molecules, such as membrane lipids or mitochondrial DNA, with the formation of lipid or nucleotide peroxides, especially at the level of thymine.

The harmful effect of ROS is primarily due to their ability to initiate a variety of autoxidative chain reactions on unsaturated fatty acids (Smirnoff 2000). Oxidative attack on proteins results in site specific amino acid modifications, fragmentation of the peptide chain, aggregation of cross linked reaction products and increased susceptibility to proteolysis (Ahmad et al. 2010b, c, 2011). ROS can also induce numerous lesions in DNA that cause deletions, mutations and other lethal genetic effects (Srivalli et al. 2003; Tuteja et al. 2009).

| Salinity class    | ECe range (dS/m) |  |  |
|-------------------|------------------|--|--|
| Non-saline        | 0–2              |  |  |
| Low salinity      | 2–4              |  |  |
| Moderate salinity | 4-8              |  |  |
| High salinity     | 8-16             |  |  |
| Severe salinity   | 16-32            |  |  |
| Extreme salinity  | >32              |  |  |

 Table 1.1
 Different classes of salinity

The toxic effects of ROS are counteracted by enzymatic as well as nonenzymatic antioxidative system such as: superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX), glutathione reductase (GR), ascorbic acid (AsA), tocopherol, glutathione and phenolic compounds, etc. (Shi and Zhu 2008; Sharma and Dietz 2009; Ashraf 2009; Ahmad et al. 2008a, b, 2010a, b, c, 2011, 2012b). Normally, each cellular compartment contains more than one enzymatic activity that detoxifies a particular ROS.

#### **1.2 Salinity in India and World**

In the arid and semi-arid regions, low rainfall coupled with ambiguity of its occurrence has been the major limiting factors in crop production. This is mainly true for India because most of the agricultural productive regions lie in hyper-arid to subhumid regions where evaporation far exceeds the rainfall. The data collected for salt affected areas in different states by Soil Resource Maps has been published by the NBBS and LUP, Nagpur (Bhargava 2005). The NBSS and LUP (National Bureau of Soil Survey and Land Use Planning) had mapped saline soils into six classes on the basis of ECe (electrical conductivity of the saturation extract) viz., slight, moderate, moderately strong, strong, severe and very severe (Table 1.1). While compiling the figures for salt affected soils the values for ECe ranging between 2-4 dS/m were discarded because they constitute non-saline class as per norms evolved by (Richards 1954). Similarly, Bureau categorized the soil sodicity classes in three classes on the basis of ESP (exchangeable sodium percentage) namely high (< 5), moderate (5–15) and strong (>15). All the soils with less than 5 ESP have been considered non-sodic or non-alkali. But all black soils or vertisols with >5 ESP have been considered sodic or alkali. The Rann of Kutchchh (Bangalore), which has an area of 2.1507 million ha and comprises a saline marsh, has been separated in the Maps published by the Bureau. The figures so extracted have been presented in Table 1.2.

At global scale more than 77 mha of land are salt affected and about 43 mha are attributed to secondary salinization as reported by FAO (2007). It is estimated that about one-third of the irrigated land in the major countries with irrigated agriculture is badly affected by salinity or is likely to be salinized in the near future.

| State            | Saline  | Sodic   | Total    |
|------------------|---------|---------|----------|
| Punjab           | 10.2    | 190.9   | 201.1    |
| Haryana          | 175.2   | 255.7   | 430.9    |
| Rajasthan        | 490.4   | 9.5     | 499.9    |
| Dehli            | 21.2    | Nil     | 21.2     |
| Uttar Pradesh    | 128.9   | 3,394.1 | 3,523.0  |
| Bihar            | _       | 229.0   | 229.0    |
| West Bengal      | 377.7   | _       | 377.7    |
| Maharashtra      | 317.3   | 114.6   | 431.9    |
| Andhra Pradesh   | 312.7   | 217.2   | 529.9    |
| Madhya Pradesh   | 1,743.4 | 26.6    | 1,770.0  |
| Karnataka        | 100.0   | 10.0    | 110.0    |
| Orissa           | 74.5    | _       | 74.5     |
| Tamil Nadu       | 50.0    | 46.2    | 96.2     |
| Goa              | 1.0     | _       | 1.0      |
| A & N Islands    | 91.9    | _       | 91.9     |
| Karaikal         | 1.7     | 1.8     | 3.5      |
| Pondicherry      | 1.5     | 1.9     | 3.4      |
| Kerala           | 203.3   | _       | 203.0    |
| Jammu & Kashmir  | 40.0    | 20.2    | 60.0     |
| Gujarat          | 3,598.4 | 846.2   | 4,444.6  |
| Rann of Kutchchh | 2,150.7 | _       | 2,150.7  |
| Total            | 9,889.7 | 5,363.7 | 15,253.4 |

 Table 1.2
 Distribution of saline/sodic soils in India (,000 ha)

Source: (Bhargava 2005)

Current estimates of the salt-affected soils as percent of the irrigated lands for different countries are: 27% for India, 28% for Pakistan, 13% for Israel, 20% for Australia, 15% for China, 50% for Iraq, and 30% for Egypt (Stockle 2001). In only New South Wales (NSW) state of Australia, salinity is estimated to affect 15% of the irrigated land.

There has been a sequential increase in the level and strength of salinity that affects soil. A number of major irrigation schemes throughout the world have suffered to some extent from the effect of salinity and/or sodocity. Many once-productive areas have become salt affected wastelands. Salt related problems occur within the boundaries of at least 75 countries (Szabolcs 1994). An alphabetical list of the countries with serious salinity problems include Australia, China, Egypt, India, Iraq, Mexico, Pakistan, the Soviet Union, Syria, Turkey and the United States (Rhoades 1998). It has been reported by different international organizations like ICID (International Commission on Irrigation and Drainage), UNEP (United Nations Environment Programme) and FAO (Food and Agriculture Organization) that salinization and water logging of the soil in arid and semi-arid regions are highly responsible for the loss of agricultural productivity on irrigated land. Therefore, the focus should be on urgent measures to combat desertification via modification of farming

techniques and amelioration of salt affected and water logged soils with a resultant improvement in social and economic conditions of people dependent on agriculture (Rhoades 1998).

#### **1.3** Causes and Types of Soil Salinity

Salinity has also been categorized as primary and secondary on the basis of their source of cause. Former occurs as natural salt in the landscape like salt marshes, salt lakes, tidal swamps or natural salts clads. While as latter results due to human activity such as urbanization and agriculture (irrigated and dry land). Following are the factors responsible for soil salinity:

For Primary salinity:

- I. Weathering of rocks
- II. Capillary rise from shallow brackish groundwater
- III. Intrusion of sea water along the coast
- IV. Salt laden sand blown by sea winds
- V. Impeded drainage

Secondary salinization is due to human activities like:

- I. Introduction of irrigation without proper drainage system
- II. Industrial effluents
- III. Overuse of fertilizers
- IV. Removal of natural plant cover
- V. Flooding with salt rich waters
- VI. High water table and the use of poor quality ground water for irrigation

#### 1.3.1 Types of Salinity

There are two types of salt affected soils:

- (i) Sodic soils
- (ii) Saline soils

The main differences between these two lies in the nature of anions and the pH of the soil. Studies demonstrate that carbonate or bicarbonate ions constitute the sodic soils with pH above 8.5, whereas chloride or sulphate ion dominates the saline soils with pH below 8.5. Some plants grow well in salt affected coastal areas, shores of backwaters lakes and marshy lands. The plants that thrive well in high salt concentrations are called halophytes. However, some plants that cannot withstand even 10% of seawater are called glycophytes or non-halophytes (Gorham 1995; Cherian et al. 1999; Parida and Das 2005; Yadav et al. 2011; Mane et al. 2011).

#### **1.4 Reclamation and Management Strategies**

Reclamation is the process of restoring disturbed land into a cultivable soil. While as management is the sum total of all procedures to protect soil and increase its performance. This reclamation process of saline soil includes the following methods:

#### 1.4.1 Physical Method

Physical method of reclamation of saline soil includes the following processes:

#### 1.4.1.1 Scarping

Scraping is the temporary method for soil reclamation in which salt layer of soil surface is scrapped off by mechanical means and the lower layer with less salt content is used for cultivation. Since, with the lowering of ground level in relation to water table salt accumulates again, this method has resulted in limited success. Thus, it again intensifies the problem. This method is rarely used because it involves high cost (Gupta and Gupta 1987).

#### 1.4.1.2 Flushing

Another method of desalinization is to flush the soil with water. In this method accumulated salt on the surface is washed away by water which is associated with very low permeability and high salinity content in their surface layer. This method has little practical significance and shows high efficiency at the beginning but grad-ually decreases as saline concentration begins to fall.

#### 1.4.1.3 Leaching

In this method, the excess amount of salt can be removed by applying water onto the soil surface. The soluble salts dissolved and transported through the soil are consequently removed from the root zone through drainage. Nevertheless, the amount of liquid required for this process varies and depends on the various characteristics of the soil (Gupta and Gupta 1987). The quantity of soluble salt leached per unit volume of water applied describes the efficiency of leaching (Tanji 1990) and this depends on the (a) uptake of water (b) uniformity of distribution of water on the soil surface, and (c) sufficiency of drainage. The major factors like primary salt content, soil salinity requirement after leaching and depth of root zone becomes imperative to provide a reliable estimate of the quantity of water required for leaching. Leaching of the salts from the soil is usually done by two methods (a) continuous leaching (b) intermittent leaching.

*Continuous ponding* is the traditional method for leaching of salts from surface irrigated lands (Tanji 1990). In this method of leaching, the water flows in the macropores with salts in the micropores to diffuse to the mobile water. Being faster, this method is also extensively used when time becomes the limiting factor.

*Intermittent ponding* is most effective method for leaching when water is the limiting factor. Less than 30–35% of water is needed for intermitted ponding than continuous ponding, but the main disadvantage of this method is that it is much slower than continuous ponding method (Gupta and Gupta 1987).

#### 1.4.2 Chemical Method

Gypsum, sulfuric acid and farm yard manure amendments are applied especially in sodic or saline sodic soils. Sodic and saline sodic soil reclamation needs a different approach than saline soils which might be more costly. Furthermore an increase in the infiltration rates is to be required in sodic soils for reclamation that can be achieved by mechanical and chemical measures. Gypsum is considered as one of the most helpful soil amendment to leach out cations (Na<sup>+</sup>) from the soil. Gypsum is a slight soluble salt of calcium and sulphate and hence it will react slowly with the soil. The gypsum amount required will vary widely depending on the percentage of exchangeable sodium and soil texture.

It has been investigated that chemical amendment-based technology has been established to reclaim the alkali/sodic soils for proper regulation of salts in root zone (Singh 2009). This includes field leveling, bunding, soil sampling to know the sodicity status for working out amendment dose application of gypsum/pyrite as per the need of the soil followed by rice-wheat rotation for 3–4 years including sesbania as a green manure crop after wheat harvest in April.

#### 1.4.3 Biological Method

Leaching and water quality appears to be limited for the knowledge of soil amendments. By biological reclamation the use of saline wastelands can be made possible through effective management strategies by using salt tolerant plants. To reclaim salt affected soils of unproductive agricultural lands by biological means may be a feasible choice. Agricultural land fed with rain water, lack of irrigation water, shallow and brackish groundwater suggests that the salt affected lands may better be cultivated with crops tolerant to low to moderate saline conditions. Several varieties of salt tolerant crops like rice, wheat and mustard have been developed with a good economic yield both in high pH alkali soils and in saline soils (Singh and Sharma 2006). Among these rice varieties like CSR10, CSR13, CSR19, CSR23, CSR27, and CSR30 can be cultivated in soils with pH ranging from 9.4 to 9.8 and EC values of 6–11 dSm<sup>-1</sup>. Salt tolerant varieties with their level of tolerance to soil salinity and alkalinity has been shown in Table 1.3.

|            |  | Adaptability  |                                     |
|------------|--|---------------|-------------------------------------|
| Crop       | Tolerant varieties   | Sodic with pH | Saline with EC (dSm <sup>-1</sup> ) |
| Rice       | CSR 10 <sup>a</sup> , CSR 11, CSR 12, CSR 13 <sup>a</sup>                      | 9.8-10.2      | 6–11                                |
|            | CSR19, CSR23 <sup>a</sup> , CSR27 <sup>a</sup> , CSR30 <sup>a</sup> ,<br>CSR1, | 9.4–9.8       | 6–11                                |
|            | CSR2, CSR3, CSR4 <sup>a</sup> , CST7-1 <sup>a</sup> ,                          | 9.4–9.8       | 6–9                                 |
| Wheat      | KRL 1–4 <sup>a</sup> , WH157   | <9.3          | 6–10                                |
|            | Raj3077, KRL19ª  | <9.3          | 6–10                                |
| Barley     | DL200, Ratna, BH97, DL348  | 8.8–9.3       |                                     |
| Indian     | Pusa Bold, Varuna  | 8.8-9.2       | 6–8                                 |
| Mustard    | Kranti, CS52ª, CSTR330-1,  | 8.8–9.3       | 6–9                                 |
|            | CST609-B 10, CS54 <sup>a</sup>   | 8.8–9.3       | 6–9                                 |
| Gram       | Karnal Chana 1   | <9.0          | <6.0                                |
| Sugar beet | Ramonskaaya 06, Maribo<br>Resistapoly  | 9.5–10        | <6.5                                |
| Sugarcane  | Co453, Co1341  | <9.0          | ECe-10                              |

Table 1.3 Suggested salt tolerant varieties

<sup>a</sup>Institute varieties released by central varietal released committee. *Source*: (Singh 2009)

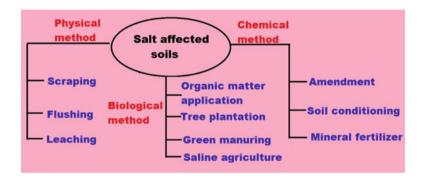


Fig. 1.1 Integrated management practices for the reclamation of salt affected soils

The biological reclamation techniques provide the earnings to the farmers without the need of costly drainage and reclamation work. Salt-tolerant trees and shrub species especially *Atriplex, Eucalyptus, Chenopodium, Suaeda, Salicornia, Kochia, Sesbania, Salsola, Juncus* and others can be grown efficiently in those areas of the land that cannot be restored (Aronson 1985; Le-Houerou 1986). For land reclamation purpose several investigators have studied the cultivation of salinity and sodocity-tolerant plants like grasses (Qadir et al. 1996), agronomic and horticultural crops (Ahmad et al. 1992, 2006, 2008a, b, 2010a, 2012a; Azooz et al. 2011), forest species. Furthermore this process can be made more effective by combining various ameliorative methods. As a result the outcome of this interaction becomes more impressive rather applied singly (Fig. 1.1).

#### 1.5 Plant Adaptations to Salt Stress

Under osmotic stress, plants accumulate osmotically active compounds called osmolytes in order to lower the osmotic potential. These are referred to as compatible metabolites because they do not apparently interfere with the normal cellular metabolism of the cell (Ahmad and Sharma 2008; Ahmad and Prasad 2012a, b). The primary function of compatible solutes is to maintain cell turgor and thus providing the driving gradient for water uptake. Recent studies indicate that compatible solutes can also act as free-radical scavengers or chemical chaperones by directly stabilizing membranes and/or proteins (Mc Neil et al. 1999; Diamant et al. 2001). Glycerol and sucrose helps by were discovered by empirical methods to protecting the biological macromolecules against the damaging effects of salinity. Later, a systematic examination of the molecules, which accumulate in halophytes and halo-tolerant organisms, led to the identification of a variety of molecules also able to provide protection (Arabawa and Timasheff 1985; Wiggins 1990). These molecules are not highly charged, but are polar, highly soluble and have a larger hydration shell. Such molecules will be preferentially solubilized in the bulk water of the cell where they could interact directly with the macromolecules.

#### 1.5.1 Glycine-Betaine

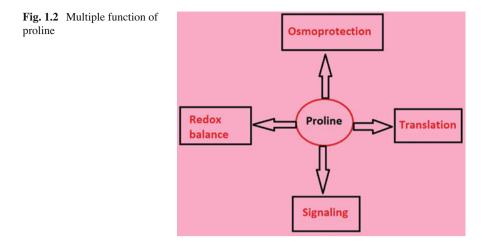
Plants synthesise glycine betaine-a major osmolyte that have been widely observed in various plant species conferring salt-tolerance (Rhodes and Hanson 1993; Hanson et al. 1994; Ahmad and Sharma 2008; Chen and Murata 2011; Koyro et al. 2012). Highly tolerant species, i.e., Spartina and Distichlis show highest accumulation, low tolerant accumulate average levels while as sensitive one's show low levels of accumulation of glycine-betaine (Rhodes et al. 1989). Glycinebetaine is synthesized from choline in two steps, the first being catalyzed by choline mono-oxygenase leading to synthesis of betaine aldehyde, which is further oxidized by betaine-aldehyde dehydrogenase (Ahmad and Sharma 2008; Chen and Murata 2011; Koyro et al. 2012). Genetic evidence showed that glycine-betaine has been obtained to develop the salinity tolerance for barley and maize (Rhodes et al. 1989; Grumet and Hanson 1986). Among these two plants, isogenic barley lines show different abilities to adjust osmotically. Transgenic rice plants expressing betaine-aldehyde dehydrogenase converted high levels of exogenously applied betaine aldehyde to glycinebetaine than did wild-type plants. The elevated level of glycinebetaine in transgenic plants conferred significant tolerance against salt, cold and heat stress (Chen and Murata 2011). Transgenic plants expressing bacterial gene for the synthesis of GB is given in table 1.4.

| Gene   | Plant species           | Tolerance to            | Reference             |
|--------|-------------------------|-------------------------|-----------------------|
| codA   | Arabidopsis             | Salt, Chilling stress   | Hayashi et al. 1997   |
| cox    | Arabidopsis             | Salt, drought, freezing | Huang et al. 2000     |
| codA   | Arabidopsis             | Salt                    | Sulpice et al. 2003   |
| codA   | Brassica juncea         | Salt                    | Prasad et al. 2000    |
| codA   | Diospyras kaki          | Salt                    | Gao et al. 2000       |
| codA   | Lycopersicon esculantum | Chilling, salt          | Park et al. 2004      |
| cox    | Nicotiana tabacum       | Salt                    | Huang et al. 2000     |
| codA   | Oryza sativa            | Salt                    | Mohanty et al. 2002   |
| cox    | Oryza sativa            | Salt                    | Su et al. 2006        |
| codA   | Soalnum tuberosum       | Salt, drought           | Ahmad et al. 2008     |
| BADH   | Nicotiana tabacum       | Salt                    | Zhou et al. 2008      |
| BADH   | Nicotiana tabacum       | Salt                    | Yang et al. 2008      |
| СМО    | Nicotiana tabacum       | Salt, drought           | Zhang et al. 2008     |
| BADH   | Lycopersicon esculantum | Salt                    | Zhou et al. 2007      |
| СМО    | Oryza sativa            | Salt                    | Shirasawa et al. 2006 |
| OsCMO  | Nicotiana tabacum       | Salt                    | Luo et al. 2012       |
| SoBADH | Ipomoea batatas         | Salt                    | Fan et al. 2012       |

 Table 1.4
 Transgenic plants expressing bacterial gene for the synthesis of GB and their tolerance to salt stress

#### 1.5.2 Polyamines

Any stress factor like osmotic stress, low pH, potassium deficiency, nutrient deficiency or light leads to accumulation of polyamines. Among polyamines, putrescine accumulation is correlated with increased argenine decarboxylase (ADC) activity in oats. Similar studies reports that transgenic carrot cells over expressing ornithine decarboxylase (ODC) cDNA considerably show that these cells were significantly more tolerant to both salt stress as well as water stress (Bohnert et al. 1995). It has been demonstrated that increased ethylene synthesis and seed germination leads to the suppression of polyamine biosynthesis (Gallardo et al. 1995). Lin and Kao (1995) observed that spermidine increases and putrescine decreases in the shoot and roots of rice seedlings and their accumulation including spermine with ADC activity signifies a specific role in salt-tolerance (Ahmad et al. 2012b). Polyamines such as spermine and spermidine are derived from methionine and ornithine while as putriscine from arginine. The first step involves the decarboxylation of ornithine catalysed by ODC (Ahmad et al. 2012b). Furthermore polyamines and their corresponding enzyme activities are substantially enhanced under salt and drought stress (Lefevre and Lutts 2000). Nuclear DNA is found to be stabilized by histones in eukaryotic organisms while as putrescine and polyamines take over the role of histones in bacteria besides the regulation of DNA in plant mitochondria and chloroplasts. Moreover polyamines stimulate several protein biosynthesis mostly through nucleic acid interaction. They also play a key role in stabilization of bio-membranes.



#### 1.5.3 Proline

Proline plays a key role in osmoregulation in plants subjected to hyperosmotic stresses, primarily drought and salinity stress (Ahmad and Jhon 2005; Ahmad et al. 2006, 2010a, 2012a; Ahmad and Sharma 2010). Accumulation of proline is a means of adaptation to abiotic stress and has been observed in rye grass for the first time by Kemble and MacPherson (1954). There are various other compatible solutes including glycine betaine (Mc Cue and Hanson 1990), and polyols that have been shown to accumulate in plants subjected to osmotic stress conditions (Adams et al. 1992). Proline accumulation influences stress tolerance in different ways (Fig. 1.2). Proline acts as a molecular chaperone protecting protein integrity and thereby increase the activities of many enzymes. Besides plants, proline accumulation has been found most widely distributed osmolyte in eubacteria, protozoa, marine invertebrates and algae (Mc Cue and Hanson 1990). Investigations shows that enhancement of proline is due to the stimulation of biosynthetic pathway in plants. Glutamate or ornithine is utilized for the synthesis of proline, glutamate being the primary precursor in osmotically stressed cells (Ahmad and Sharma 2008; Koyro et al. 2012). Transcripts corresponding to both cDNAs that accumulate against NaCl stress are found to play a regulatory key to build up strategies for overproduction of proline in a selected plant species. Furthermore the intermediates of proline biosynthetic pathways also enhance the expression of numerous genes that are regulated osmotically in rice (Iyer and Caplan 1998). Evidence also shows that proline degradation in the mitochondria is linked to respiratory electron transport system as well as ATP production. Expression of P5CS transgenic rice from moth-bean led to stress-induced overproduction of the P5CS enzyme and proline accumulation in transgenic rice plants under the control of an inducible promoter. While as second generation (RI) transgenic plants observed an enhancement of biomass in response to salt and water stress (Zhu et al. 1998.). Table 1.5 shows response of different plants towards proline accumulation under salt stress.

|               | Species                      | 11 1.                                  |                                     |
|---------------|------------------------------|--|-------------------------------------|
| <u></u>       | T T T T T T                  | salinity                               | References                          |
| Soluble       | Pisum sativum                | Increase                               | Ahmad and Jhon 2005                 |
| protein       | Oryza sativa                 | Decrease                               | Alamgir and Ali 1999                |
|               | Vicia faba                   | Decrease                               | Gadallah 1999                       |
|               | Amaranthus tricolor          | Decrease                               | Wang and Nil 2000                   |
|               | Bruguiera parviflora         | Decrease                               | Parida et al. 2002                  |
|               | Pancratium maritimum         | Increases at low salinity              | Khedr et al. 2003                   |
|               |                              | Decrease at high salinity              |                                     |
|               | Arabidopsis thaliana         | Increase                               | Quintero et al. 1996                |
|               | Morus alba                   | Increase                               | Ahmad and Sharma 2010               |
|               | Brassica juncea var. Bio902  | Increase                               | Mittal et al. 2012                  |
|               | Brassica juncea var. Urvashi | Decrease                               | Mittal et al. 2012                  |
|               | Beta vulgarus                | Decrease                               | Jamil et al. 2012a                  |
|               | Oryza sativa                 | Decrease                               | Jamil et al. 2012b                  |
|               | Portulaca oleraceae          | Decrease                               | Rahdari et al. 2012                 |
|               | Setaria italica              | Increase                               | Hendawy et al. 2012                 |
|               | Portulaca oleraceae          | Increases                              | Rahdari et al. 2012                 |
|               | Prunus species               | Decrease                               | Sorkheh et al. 2012                 |
|               | Borago officinalis           | Decrease                               | Enteshari et al. 2011               |
| Proline       | Pisum sativum                | Increase                               | Ahmad and Jhon 2005                 |
|               | Portulaca oleraceae          | Increases                              | Rahdari et al. 2012                 |
|               | Matricaria chamomilla        | Increases                              | Heidari and Sarani 2012             |
|               | Borago officinalis           | Increases                              | Enteshari et al. 2011               |
|               | Brassica juncea var. Bio902  | Increase                               | Mittal et al. 2012                  |
|               | Brassica juncea var. Urvashi | Decrease                               | Mittal et al. 2012                  |
|               | Suaeda maritima              | Increase                               | Rajaravindran and<br>Natarajan 2012 |
|               | Lycopersicon esculantum      | Increase                               | Babu et al. 2012                    |
|               | Morus alba                   | Increase                               | Ahmad and Sharma 2010               |
| Carbohydrates | Portulaca oleraceae          | Increases                              | Rahdari et al. 2012                 |
|               | Matricaria chamomilla        | Increases                              | Heidari and Sarani 2012             |
|               | Beta vulgaris L.             | Increases                              | Dadkhah 2010                        |
|               | Borago officinalis           | Decrease                               | Enteshari et al. 2011               |
|               | Prosopis alba                | Increase in<br>soluble<br>carbohydrate | Meloni et al. 2004                  |
|               | Morus alba                   | Increase                               | Ahmad and Sharma 2010               |

 Table 1.5
 Responses of biochemical attributes under salt stress

#### 1.5.4 Carbohydrates

Osmotic potential accounts more than 50% of sugar in glycophytes subjected to saline conditions (Cram 1976). Despite a significant decrease in net  $CO_2$  assimilation rate its accumulation in plants have been widely reported (Murakeozy et al. 2003).

Accumulation of carbohydrates plays a central role in osmoprotection, osmotic adjustment, carbon storage and radical scavenging under salt stress (Ahmad and Sharma 2008; Koyro et al. 2012). Trehalose, have shown to accumulate and protects membranes and proteins in cells against water deficit and reduced aggregation of denatured proteins (Singer and Lindquist 1998). At the same time suppressive effect has been observed by trehalose on apoptotic cell death (Yamada et al. 2003) suggesting the presence of trace amounts in vascular plants, including major crops, though the definite role of this osmolyte in metabolism is still unclear.

Role of sugars in adaptation of plants to salinity have been concluded to be universally associated with salt tolerance. However, this does not signify the role of indicator for salt tolerance in breeding programs for some species. Table 1.5 shows response of different plants towards carbohydrate accumulation under salt stress.

#### 1.5.5 Proteins

Salt-induced proteins in plants have been classified into two major groups (Mansour 2000), i.e., salt stress proteins, which accumulate only due to salt stress, and stress associated proteins, that accumulates in response to various abiotic stress like heat, cold, drought, water logging, and high and low mineral nutrients. Protein accumulation also provides a storage pool of nitrogen to be re-utilized later (Singh et al. 1987) and also a key role in osmotic adjustment. Large number of cytoplasmic proteins cause alterations in cytoplasmic viscosity of the cells stimulated by salinity (Hasegawa et al. 2000). Proteins have shown to increase on exposure to salt stress and can be synthesized *de novo* in response to salt stress or may be present constitutively at low concentration (Pareek et al. 1997). Protein with 26 kDa named as osmotin have been detected in tobacco in response to salt stress (Singh et al. 1987). In salt stressed *Mesembryanthemum crystallinum* another osmotin-like protein was also observed to increase as compared to non-stressed plants (Thomas and Bohnert 1993). In barley, two 26 kDa polypeptides, identified as germin which is not immunologically related to osmotin, have been found to increase in response to salt stress (Hurkman et al. 1991). Similar reports have been found in radish with 22 kDa protein (Lopez et al. 1994). Salt tolerant shows higher soluble proteins than salt sensitive species of barley (Hurkman et al. 1989), sunflower, finger millet (Uma et al. 1995), and rice (Pareek et al. 1997). Table 1.5 shows response of different plants towards protein accumulation under salt stress.

#### 1.6 Reactive Oxygen Species and Antioxidants

Adaptation of the plant cell to high salinity involves osmotic adjustment and the compartmentation of toxic ions, whereas an increasing body of evidence suggests that high salinity also induces the generation of reactive oxygen species (ROS) and