

Biomaterials Science: Processing, Properties, and Applications V

Edited by

Roger Narayan

Susmita Bose

Amit Bandyopadhyay

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Contents

[PREFACE](#)

[NEXT GENERATION BIO CERAMICS](#)

[EVALUATION OF LONG-TERM MECHANICAL AND BIOLOGICAL BIOCOMPATIBILITY OF LOW-COST \$\beta\$ -TYPE Ti-Mn ALLOYS FOR BIOMEDICAL APPLICATIONS](#)

[ABSTRACT](#)

[INTRODUCTION](#)

[EXPERIMENTAL](#)

[RESULTS AND DISCUSSION](#)

[CONCLUSIONS](#)

[ACKNOWLEDGEMENT](#)

[REFERENCES](#)

[CONTROL OF Ag RELEASE FROM Ag-CONTAINING CALCIUM PHOSPHATES IN SIMULATED BODY FLUID](#)

[ABSTRACT](#)

[INTRODUCTION](#)

[EXPERIMENTAL PROCEDURES](#)

[RESULTS AND DISCUSSION](#)

[CONCLUSIONS](#)

[ACKNOWLEDGMENTS](#)

[REFERENCES](#)

[GALLIUM-CONTAINING FERRITES FOR HYPERTHERMIA TREATMENT](#)

[ABSTRACT](#)

[INTRODUCTION](#)

MATERIALS AND METHODS

RESULTS AND DISCUSSIONS

CONCLUSIONS

REFERENCES

EXPLORATION OF AMORPHOUS AND
CRYSTALLINE TRI-MAGNESIUM PHOSPHATES FOR
BONE CEMENTS

ABSTRACT

INTRODUCTION

EXPERIMENTAL

RESULTS AND DISCUSSION

CONCLUSION

ACKNOWLEDGEMENTS

REFERENCES

MICRO-X-RAY DIFFRACTION STUDY OF NEW
NICKEL-TITANIUM ROTARY ENDODONTIC
INSTRUMENTS*

ABSTRACT

INTRODUCTION

EXPERIMENTAL PROCEDURES

RESULTS

DISCUSSION

CONCLUSIONS

ACKNOWLEDGMENT

REFERENCES

TORSIONAL PROPERTIES OF NANOSTRUCTURED
TITANIUM CORTICAL BONE SCREWS

ABSTRACT

INTRODUCTION

MATERIALS AND METHODS

RESULTS AND DISCUSSION

CONCLUSIONS

RECOMMENDATION

ACKNOWLEDGEMENT

REFERENCES

STRENGTHENING BEHAVIORS OF LOW-PRECIOUS
Ag-Pd-Au-Zn ALLOYS FOR DENTAL APPLICATIONS

ABSTRACT

INTRODUCTION

UNIQUE HARDENING BEHAVIOR OF Ag-20Pd-
12Au-14.5Cu ALLOY

EVALUATION OF MICROSTRUCTURE AND
MECHANICAL PROPERTIES OF Ag-20Pd-12Au-
14.5Cu ALLOYS WITH AND WITHOUT A PHASE

SUMMARY

REFERENCES

EFFECT OF IMMERSION MEDIUM ON THE
DEGRADATION AND CONVERSION OF SILICATE
(13-93) BIOACTIVE GLASS SCAFFOLDS

ABSTRACT

1. INTRODUCTION

2. MATERIALS AND METHODS

3. RESULTS

4. DISCUSSION

5. CONCLUSIONS

ACKNOWLEDGEMENTS

REFERENCES

EVALUATION OF LONG-TERM BONE
REGENERATION IN RAT CALVARIAL DEFECTS

IMPLANTED WITH STRONG POROUS BIOACTIVE GLASS (13-93) SCAFFOLDS

ABSTRACT

1. INTRODUCTION

2. MATERIALS AND METHODS

3. RESULTS

4. DISCUSSION

5. CONCLUSIONS

ACKNOWLEDGEMENTS

REFERENCES

MAGNESIUM SINGLE CRYSTAL AS A BIODEGRADABLE IMPLANT MATERIAL

ABSTRACT

1. INTRODUCTION

2. EXPERIMENTAL DETAILS

3. RESULTS AND DISCUSSION

4. CONCLUSIONS

ACKNOWLEDGEMENTS

REFERENCES

SURFACE PROPERTIES OF BIOMATERIALS

DAMAGE EVALUATION OF TiO₂ NANOTUBES ON TITANIUM

ABSTRACT

1.0 INTRODUCTION

2.0 MATERIALS AND METHODS

3.0 RESULTS

4.0 DISCUSSIONS

5.0 CONCLUSIONS

6.0 ACKNOWLEDGEMENTS

7.0 REFERENCES

DRUG DELIVERY FROM SURFACE MODIFIED TITANIUM ALLOY FOR LOAD-BEARING IMPLANTS

ABSTRACT

INTRODUCTION

MATERIALS AND METHODS

RESULTS

DISCUSSION

ACKNOWLEDGEMENT

REFERENCES

A FAMILY OF NOVEL BIOSTABLE RETICULATED ELASTOMERIC AND RESILIENT BIOINTREGATIVE CROSSLINKED POLYURETHANE-UREA SCAFFOLDS

ABSTRACT

1. INTRODUCTION

2. MATERIALS

3. CHARACTERIZATION

4. RESULTS

5. CONCLUSIONS

6. REFERENCES

FIGURE

IN SITU NITRIDATION OF TITANIUM USING LENS[™]

ABSTRACT

1. INTRODUCTION

2. EXPERIMENTAL

3. RESULTS

4. DISCUSSIONS

5. CONCLUSIONS

6.0 ACKNOWLEDGEMENTS

REFERENCES

MAGNESIUM DOPED HYDROXYAPATITE: SYNTHESIS, CHARACTERIZATION AND BIOACTIVITY EVALUATION

ABSTRACT

INTRODUCTION

MATERIALS AND METHODS

RESULT AND DISCUSSION

CONCLUSIONS

ACKNOWLEDGEMENT

REFERENCES

NOVEL PLA- AND PCL-HA POROUS 3D SCAFFOLDS PREPARED BY ROBOCASTING FACILITATE MC3T3- E1 SUBCLONE 4 CELLULAR ATTACHMENT AND GROWTH

ABSTRACT

INTRODUCTION

MATERIALS AND METHODS

RESULTS

DISCUSSION

CONCLUSIONS

ACKNOWLEDGEMENTS

REFERENCES

DEXTRAN COATED CERIUM OXIDE NANOPARTICLES FOR INHIBITING BONE CANCER CELL FUNCTIONS

ABSTRACT

1. INTRODUCTION:

2. MATERIALS AND METHODS:

[3. RESULTS](#)
[4. DISCUSSION:](#)
[5. CONCLUSIONS:](#)
[ACKNOWLEDGEMENTS](#)
[REFERENCES](#)
[AUTHOR INDEX](#)
[EULA](#)

List of Tables

[EVALUATION OF LONG-TERM MECHANICAL AND BIOLOGICAL BIOCOMPATIBILITY OF LOW-COST \$\beta\$ -TYPE Ti-Mn ALLOYS FOR BIOMEDICAL APPLICATIONS](#)

[Table I](#)

[CONTROL OF Ag RELEASE FROM Ag-CONTAINING CALCIUM PHOSPHATES IN SIMULATED BODY FLUID](#)

[Table I](#)

[Table II](#)

[GALLIUM-CONTAINING FERRITES FOR HYPERTHERMIA TREATMENT](#)

[Table 1](#)

[Table 2](#)

[Table 3](#)

[Table 4](#)

[EXPLORATION OF AMORPHOUS AND CRYSTALLINE TRI-MAGNESIUM PHOSPHATES FOR BONE CEMENTS](#)

[Table 1](#)

[Table 2](#)

Table 3

MICRO-X-RAY DIFFRACTION STUDY OF NEW NICKEL-TITANIUM ROTARY ENDODONTIC INSTRUMENTS*

Table 1

TORSIONAL PROPERTIES OF NANOSTRUCTURED TITANIUM CORTICAL BONE SCREWS

Table I

Table II

Table III

Table IV

EFFECT OF IMMERSION MEDIUM ON THE DEGRADATION AND CONVERSION OF SILICATE (13-93) BIOACTIVE GLASS SCAFFOLDS

Table I

Table II

Table III

EVALUATION OF LONG-TERM BONE REGENERATION IN RAT CALVARIAL DEFECTS IMPLANTED WITH STRONG POROUS BIOACTIVE GLASS (13-93) SCAFFOLDS

Table I

Table II

Table III

MAGNESIUM SINGLE CRYSTAL AS A BIODEGRADABLE IMPLANT MATERIAL

Table I

Table II

Table III

[Table IV](#)

[Table V](#)

[Table VI](#)

[DAMAGE EVALUATION OF TiO₂ NANOTUBES ON TITANIUM](#)

[Table 1](#)

[A FAMILY OF NOVEL BIOSTABLE RETICULATED ELASTOMERIC AND RESILIENT BIOINTREGATIVE CROSSLINKED POLYURETHANE-UREA SCAFFOLDS](#)

[Table 1](#)

[MAGNESIUM DOPED HYDROXYAPATITE: SYNTHESIS, CHARACTERIZATION AND BIOACTIVITY EVALUATION](#)

[Table 1.](#)

[Table 2.](#)

[Table 3.](#)

List of Illustrations

[EVALUATION OF LONG-TERM MECHANICAL AND BIOLOGICAL BIOCOMPATIBILITY OF LOW-COST \$\beta\$ -TYPE Ti-Mn ALLOYS FOR BIOMEDICAL APPLICATIONS](#)

[Figure 1. Geometry of \(a\) tensile and \(b\) fatigue specimens.](#)

[Figure 2. Typical OM images of solutionized \(a\) Ti-8Mn and \(b\) Ti-13Mn.](#)

[Figure 3. Typical XRD profiles of solutionized \(a\) Ti-8Mn and \(b\) Ti-13Mn.](#)

[Figure 4. Typical SAED patterns of solutionized \(a\) Ti-8Mn and \(b\) Ti-13Mn.](#)

Figure 5. Tensile properties of solutionized Ti-8Mn and Ti-13Mn, along with those of aged Ti-6Al-4V.

Figure 6. S-N curves of solutionized Ti-8Mn and Ti-13Mn along with region of that of aged Ti-6Al-4V ELI¹².

Figure 7. SEM fractographs of crack initiation areas, stable crack propagation areas, and final fracture areas of (a) Ti-8Mn and (b) Ti-13Mn.

Figure 8. CMR images of cross-sections of the implants: (a) solutionized Ti-12Mn implant and (b) CP-Ti implant at 12 weeks, and (c) solutionized Ti-12Mn implant and (d) CP-Ti implant at 96 weeks after implantation⁸.

Figure 9. Relative bone contact ratios of solutionized Ti-12Mn implant and CP-Ti implant at 12, 52, and 96 weeks after implantation⁸.

CONTROL OF Ag RELEASE FROM Ag-CONTAINING CALCIUM PHOSPHATES IN SIMULATED BODY FLUID

Figure 1. Amounts of Ca, P, and Ag in TBS after immersion of as-precipitated powders for (a) 6 h and (b) 24 h.

Figure 2. XRD patterns of as-precipitated powders and residue powders on filter after immersion tests of (a) CaP1.33 and (b) CaP1.67 series in TBS for 24 h.

Figure 3. Amounts of (a) Ca, (b) P, and (c) Ag for sintered compacts in TBS.

Figure 4. Surface of sintered compacts of (a) 0AgCaP1.33 and (b) 10AgCaP1.33 after immersion for 15 d. (c) EDX spectrum of the region indicated by red rectangle in (b).

GALLIUM-CONTAINING FERRITES FOR HYPERTHERMIA TREATMENT

Figure 1. XRD patterns of the materials obtained using different Fe:Ga ratios (1:1, 2:1, 3:1, 4:1) after heat treatment for 1 h at 500 °C.

Figure 2. Hysteresis loops of the MNP's obtained using Fe:Ga ratios of 1:1, 2:1, 3:1 and 4:1.

Figure 3. SEM image and corresponding EDS spectrum of the magnetic material obtained using a Fe:Ga ratio of 2:1.

Figure 4. TEM and HRTEM images and particle size distribution of the magnetic material obtained using a Fe:Ga ratio of 2:1.

Figure 5. Behavior of temperature as a function of testing time of magnetic induction of the material obtained using a Fe:Ga ratio of 2:1.

Figure 6. MNP's after 7 days of immersion in SBF.

Figure 7. MNP's after 28 days of immersion in SBF: a) area EDS analysis, b) punctual EDS analysis.

Figure 8. MNP's after 14 days of immersion in SBF with replacement of this fluid at the seventh day.

Figure 9. MNP's after 28 days of immersion in SBF with replacement of this fluid every 7 days: a) area EDS analysis, b) punctual EDS analysis.

EXPLORATION OF AMORPHOUS AND CRYSTALLINE TRI-MAGNESIUM PHOSPHATES FOR BONE CEMENTS

Figure 1: TGA and DSC of the synthesized and thermally treated $\text{Mg}_3(\text{PO}_4)_2$ powders.

Figure 2: XRD of the synthesized $\text{Mg}_3(\text{PO}_4)_2$ powders.

Figure 3: FTIR spectra of synthesized and thermally treated $\text{Mg}_3(\text{PO}_4)_2$ powders.

Figure 4: Scanning electron microscopy images of synthesized and thermally treated $\text{Mg}_3(\text{PO}_4)_2$ powders (Scale bar: 50 μm).

Figure 5: XRD spectra of 400°C, P:L of 1:2, 600°C, P:L of 1:2, and 800°C, P:L of 1:2 cements as reacted and after soaking in PBS for 2 days and 7 days. [# is $\text{Mg}_3(\text{NH}_4)_2\text{H}_4(\text{PO}_4)_4 \cdot 8\text{H}_2\text{O}$, * is $\text{Mg}_3(\text{PO}_4)_2$, + is $\text{MgNH}_4\text{PO}_4 \cdot 6\text{H}_2\text{O}$, = is $\text{MgHPO}_4 \cdot 3\text{H}_2\text{O}$, * is $\text{Mg}_3(\text{PO}_4)_2$].

Figure 6: FTIR spectra of 400°C, P:L of 1:2, 600°C, P:L of 1:2, and 800°C, P:L of 1:2, cements as reacted and after soaking in PBS for 2 days and 7 days.

Figure 7: SEM images of 400°C, 600°C, and 800°C cements, P:L of 1:2, 1:2 and 1:1 respectively. The left set of images shows the fracture surface of cements while the right set of images shows the exposed (outside) surface of the same cements. Each set compares after reaction completion and after PBS incubation. (Scale bar: 50 μm).

MICRO-X-RAY DIFFRACTION STUDY OF NEW NICKEL-TITANIUM ROTARY ENDODONTIC INSTRUMENTS*

Figure 1. Micro-XRD patterns for HyFlex® CM™ .04/20 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

Figure 2. Micro-XRD patterns for HyFlex® CM™ .04/40 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

Figure 3. Micro-XRD patterns for HyFlex® CM™ .08/25 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

Figure 4. Micro-XRD patterns for Vortex Blue® .04/20 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

Figure 5. Micro-XRD patterns for Vortex Blue® .04/30 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

Figure 6. Micro-XRD patterns for Vortex Blue® .04/40 rotary instrument at tip region (bottom) and at 3 mm, 6 mm, 9 mm and 12 mm (top) from tip.

TORSIONAL PROPERTIES OF NANOSTRUCTURED TITANIUM CORTICAL BONE SCREWS

Figure 2a. EBSD Grain Size Diameter (Control 2500X).

Figure 2b. EBSD Grain Size Diameter (Nano 10,000X).

Figure 3a. Control Torque vs. Rotation

Figure 3b. Nano Torque vs Rotation

Figure 4. Torsional Failure Location of Nano Bone Screws

Figure 5a. Fracture Surface of Nano Distal Screw Shaft (35X).

Figure 5b. Fracture Surface of Nano Prox Screw Shaft (35X).

STRENGTHENING BEHAVIORS OF LOW-PRECIOUS Ag-Pd-Au-Zn ALLOYS FOR DENTAL APPLICATIONS

Figure 1. Schematic drawing of solution treatment (ST) for LR alloy.

Figure 2. Schematic drawing of aging treatment (AT) for LR alloy.

Figure 3. Schematic drawing of solution-treatment for aging-treated LR alloy.

Figure 4. XRD profiles change and change in Vickers hardness of WP and WP-ST alloys.

Figure 5. TEM (a, d) bright field images, (b, e) selected area diffraction patterns and (c, f) key diagrams of WP and WP-ST alloys. Beam direction is parallel to [001].

Figure 6. XRD profile change of LR and LR-ST alloys.

Figure 7. TEM (a, d) bright field images, (b, e) selected area diffraction patterns and (c, f) key diagrams of LR and LR-ST alloys. Beam direction is parallel to [001].

Figure 8. Changes in Vickers hardness of (a) WP and WP-ST alloys, and (b) LR and LR-ST alloys.

Figure 9. Tensile properties of WP, WP-ST, LR and LR-ST alloys.

Fig. 10 Change in Vickers hardness of 1173WQ_{lr}/3.6 ks, 1023WQ_{lr}/1.8 ks, 1023WQ_{lr}/3.6 ks, 1023WQ_{lr}/7.2 ks, 1023WQ_{lr}/14.4 ks and 1023WQ_{lr}/28.8 ks alloys.

Figure 11. Change in Vickers hardness of 1173WQ_{lr}/3.6 ks, 673WQ_{lr}/1.8 ks, 673WQ_{lr}/3.6 ks, 673WQ_{lr}/7.2 ks, 673WQ_{lr}/14.4 ks and 673WQ_{lr}/28.8 ks alloys.

Figure 12. Vickers hardness of 673WQ_{lr}/1.8 ks and 673WQ_{lr}/1.8 ks-ST alloys.

EFFECT OF IMMERSION MEDIUM ON THE DEGRADATION AND CONVERSION OF SILICATE (13-93) BIOACTIVE GLASS SCAFFOLDS

Fig. 1. (a) SEM image of the as-fabricated silicate 13-93 bioactive glass scaffold and (b) EDS spectrum of the surface of the as-fabricated scaffold.

Fig. 2. SEM images of the surface of 13-93 bioactive glass scaffolds after immersion for 2, 4 and 6 weeks in DMEM (a, b, c) and SBF (e, f, g). The EDS spectra of the reacted surface at 6 weeks in DMEM and in SBF are shown in (d) and (h), respectively.

Fig. 3. SEM backscattered electron images (left) and X-ray maps of the cross section of 13-93 bioactive glass scaffolds immersed in DMEM and SBF for 6 weeks showing the Ca (K), P (K), Si (K) distribution.

Fig. 4. (a) FTIR spectra of 13-93 glass scaffolds and (b) thin-film XRD patterns of 13-93 glass disks, as prepared and after immersion in DMEM and SBF for 2, 4 and 6 weeks.

Fig. 5. Cumulative amount of Si, K, Ca and P released from the 13-93 bioactive glass scaffolds into (a) DMEM and (b) SBF as a function of immersion time.

EVALUATION OF LONG-TERM BONE REGENERATION IN RAT CALVARIAL DEFECTS IMPLANTED WITH STRONG POROUS BIOACTIVE GLASS (13-93) SCAFFOLDS

Fig. 1. (a) Optical image of 13-93 bioactive glass scaffold prepared by robocasting for implantation in rat calvarial defects. (b) Higher-magnification SEM image of the scaffold showing dense glass filaments and porous grid-like architecture in the plane of deposition (xy plane). Inset: SEM image in z direction. The scaffolds had a porosity of $47 \pm 1\%$, a pore width of $300 \pm 10 \mu\text{m}$ in the xy plane and $150 \pm 10 \mu\text{m}$ in z direction.

Fig. 2. Transmitted light images of H&E-stained sections of rat calvarial defects implanted for 6 weeks (a1, a2), 12 weeks (b1, b2) and 24 weeks (c1, c2) with the as-fabricated scaffolds (a1-c1) and the BMP2-loaded scaffolds (a2-c2). N = new bone; O = old (host) bone; * = bony island; G = bioactive glass; arrowheads indicate the edges of old bone.

Fig. 3. Percent new bone in rat calvarial defects implanted with the as-fabricated and BMP2-loaded scaffolds at 6, 12 and 24 weeks. The amount of new bone is shown as a percent of the pore space (area) of the scaffolds. (*significant difference within each group; **significant difference among groups for a given implantation time; $p < 0.05$).

Fig. 4. Transmitted light images of von Kossa stained sections of rat calvarial defects implanted for 6 weeks (a1, a2), 12 weeks (b1, b2) and 24 weeks (c1, c2) with the as-fabricated scaffolds (a1-c1) and BMP2-loaded scaffolds (a2-c2). N = new bone; O = old bone; G = bioactive glass; arrowheads indicate the edges of old bone.

Fig. 5. Percent total von Kossa positive (vK+) area, determined as a fraction of the total defect area, for rat calvarial defects implanted with the as-fabricated and BMP2-loaded scaffolds at 6, 12 and 24 weeks postimplantation. (*significant difference within each group; **significant difference when compared to as-fabricated scaffold at the same implantation time; $p < 0.05$).

Fig. 6. Backscattered SEM images of rat calvarial defects implanted with the as-fabricated scaffolds (a1, a2) and BMP2-loaded scaffolds (b1, b2) at 24 weeks postimplantation. N = new bone; G = bioactive glass. The approximate thickness of the converted

surface layer on the glass filaments is shown in (a2) and (b2).

MAGNESIUM SINGLE CRYSTAL AS A BIODEGRADABLE IMPLANT MATERIAL

Figure 1 Typical voltage vs. time graph during micro arc oxidation of magnesium

Figure 2. (A) Single Crystal furnace and (B) as cast single crystal

Figure 3. Setup used for anodizing the Mg crystals

Figure 4. XRD scan along single crystal length

Figure 5 Representative optical image of MAO coated sample (left) and the sparking observed during the process (right).

Figure 6. Polarization curves of Anodized (S2-upper brown curve) and non-anodized (S1-lower blue curve) Mg single crystals in Hank's solution

Figure 7. Polarization curves of Polycrystalline (Poly), polished single crystalline (S1 and 2S1) and Anodized single crystalline (2S2) Mg in 0.15M NaCl solution

Figure 8. Potentiodynamic DC polarization curves of the samples with and without MAO coating

Figure 9. Optical images of immersion test samples (a) Single crystal polished till 1200 grit, (b) Etched single crystals for 10 seconds, (c) Polycrystalline Mg polished till 1200 grit, (d) MAO coated single crystal namely before immersion, after removal from the solution and after cleaning in chromate solution at 3 different time points

Figure 10. Weight loss vs. Time graph for immersion test in DMEM+HEPES

[Figure. 11. Optical images of non-anodized \(S1\) and anodized \(S2\) samples after polarization test](#)

[Figure 12. SEM images of anodized \(S2\) samples \(a\) before and \(b\) after polarization test \(Magnification 500X\).](#)

[Fig 13 Optical microscope images of the samples after the polarization tests in 0.15 M NaCl \(A\). Polycrystalline Mg \(B\) Anodized Mg for 6 hours \(2S2\). \(C\) Single crystal Mg \(2S1\) \(D\) Single crystal Mg \(S1\).](#)

[Figure 14. Surface and cross section SEM images of the samples anodized for different times \(a\) and \(d\) 5 min, \(b\) and \(e\) 7 min, \(c\) and \(f\) 10 min.](#)

[Figure 15. SEM and AFM images of etched single crystal sample](#)

[Figure 16. Weight loss vs. Time data for in vivo testing in mice](#)

[Figure 17. Bubble volume vs. Time data for in vivo testing in mice](#)

[DAMAGE EVALUATION OF TiO₂ NANOTUBES ON TITANIUM](#)

[Figure 1: \(a\),\(b\) Low and high magnification SEM images of the anodized Ti surface showing TiO₂ nanotubes. \(c\) SEM image showing the anodized Ti surface at 30 mins showing partial coverage of the surface with nanotubes.](#)

[Figure 2: Cross-section of the anodized surface showing nanotube of length 300nm± 25nm](#)

[Figure 3: SEM images of silver nanoparticles electrodeposited on the TiO₂ nanotubes showing the coverage of the deposition on the entire surface.](#)

Figure 4: SEM Images of (a) Before Insertion and (b) After Insertion into the bone showing no significant damages of the nanotubes due to implantation.

Figure 5: Silver nanoparticles electrodeposited on TiO₂ nanotubes (a) SEM image before implantation; (b) SEM image just after implantation.

Figure 6: SEM images showing the thermal degradation for nanotubes of length 300nm from 400°C- 800°C.

Figure 7: XRD analysis of (a) as anodized Ti sample, (b) 300nm TiO₂ nanotubes sample heat treated at 800.

DRUG DELIVERY FROM SURFACE MODIFIED TITANIUM ALLOY FOR LOAD-BEARING IMPLANTS

Figure 1: Schematic diagram of the experimental setup of anodization.

Figure 2: A schematic representation of the PCL coating on the drug coating.

Figure 3: A schematic diagram of the coating by the blend.

Figure 4: The figure shows the anodized surface morphology of the Ti6Al4V samples at two different magnifications.

Figure 5: It shows the coating thickness of the anodized Ti samples with a blend of polymer and drug.

Figure 6: SEM morphologies of the (a) Ti alloy coated with drug, (b) anodized Ti alloy coated with drug SIM, (c) anodized Ti alloy consecutively coated with the drug and PCL and (d) anodized Ti alloy coated with a blend of PCL and drug SIM.

Figure 7: The figure shows the surface morphology of (a) the Ti-NT-SIM-PCL sample before any release kinetics were done and the surface morphologies of the samples are shown after the release study was performed at (b) pH 7.4 and (c) 5.0. (d) shows the whereas the surface morphology after release from the same sample at (e) pH 7.4 and (f) pH 5.0.

Figure 8: Simvastatin (SIM) release from bare Ti6Al4V and Ti-NT substrates for 2 hours at (a) pH 7.4 and (b) pH 5.0. Simvastatin (SIM) release from bare Ti6Al4V and Ti-NT substrates coated consecutively with drug SIM and PCL and coated with the blend of polymer and drug respectively for 15 days at (c) pH 7.4 and (d) pH 5.0

Figure 9: Osteoblast interactions on the various substrates after one day of incubation.

A FAMILY OF NOVEL BIOSTABLE RETICULATED ELASTOMERIC AND RESILIENT BIOINTREGATIVE CROSSLINKED POLYURETHANE-UREA SCAFFOLDS

Figure 1. SEMS of a) Biomerix PCPUU matrix b) Bard PP mesh c) Gore non-woven Bio-A d) Salt leached PLGA e) Lyophilized PLGA f) Porcine Dermal Matrix

Figure 2. Assure™ Surgical Mesh is a tri-layered composite featuring a knitted polypropylene mesh sandwiched between an adhesively bonded layer of BMX PCPUU matrix on one side and melt bonded resorbable thin anti-adhesion layer on the other side.

Figure 3. Assure™ histology at 26 weeks with (*, Biomerix matrix [}], PLLA/PCL layer [≤]), mononuclear cell infiltration (◄), fibrous tissue formation (v), blood vessel (ß), multi nucleate giant cells (ï), skeletal muscle (sm). 2x=1mm, 10x=500µm, 20x=250µm, 40x=100µm.

Figure 4. Picture of Rotator Cuff Repair device with BMX PCPUU matrix sheet, reinforced with size 5-0 braided polyester suture grid.

Figure 5. SEM's and photographs of the Neurostring coil devices comprises an outer "jacket" of BMX PCPUU matrix on a platinum coil with a shape-set superelastic nitinol wire as inner member.

Figure 6. NS coil histology at 26 week showing that the devices are fully incorporated by organized fibrous tissue with moderate to focally marked angiogenesis surrounding the embolic material, both at the periphery and the center of the sac and that prolific ingrowth has occurred into the BMX matrix.

IN SITU NITRIDATION OF TITANIUM USING LENS™

Figure 1 : X-ray diffraction pattern confirms the formation of nitrides on Ti.

Figure 2 : Microstructural depth profile of laser nitrided sample at 425W with 2 laser passes.

Figure 3: SEM image of sample nitrided at 475W with 2 laser passes.

Figure 4: Chart comparing maximum hardness of untreated CPTi plate to maximum surface hardness of treated samples.

Figure 5 : Chart showing difference in wear track volume normalized for load and distance between treated and cp-Ti samples in DI water medium.

Figure 6: Wear damage on *in situ* nitride cp-Ti surface.

MAGNESIUM DOPED HYDROXYAPATITE: SYNTHESIS, CHARACTERIZATION AND BIOACTIVITY EVALUATION

Figure 1. XRD Analysis of as synthesized MgHAP

Figure 2. A comparison of XRD spectrum of as synthesized MgHAP, MgHAP8 and MgHAP10

Figure 3. Thermogravimetric curve of as synthesized MgHAP

Figure 4. FTIR spectra of as synthesized MgHAP, MgHAP8 and MgHAP10

Figure 5. TEM images and particle size estimations of as synthesized MgHAP, MgHAP8, and MgHAP10

Figure 6. Variation in pH with soaking time of as synthesized MgHAP, MgHAP8, and MgHAP10 soaked in SBF for 28 days.

Figure 7. FTIR spectrum of as synthesized MgHAP, MgHAP8 and MgHAP10 soaked in SBF for 28 days.

Figure 8. TEM images of SBF soaked MgHAP, MgHAP8 and MgHAP10 powders

NOVEL PLA- AND PCL-HA POROUS 3D SCAFFOLDS PREPARED BY ROBOCASTING FACILITATE MC3T3-E1 SUBCLONE 4 CELLULAR ATTACHMENT AND GROWTH

Figure 1. Scanning electron micrographs of polymer-HA scaffolds (70 wt.% HA) as-deposited. [8] The scaffolds are a) PCL-HA and b) PLA-HA and have a three-dimensional, mesh-like arrangement. The scaffold pictured above were representative of the set of scaffold geometries that can be manufactured using robocasting. Scaffolds used for cyto-compatibility studies had a 1 mm rod center-to-center diameter with 0.3 mm rod edge-to-edge spacing.

Figure 2. Energy Dispersive Spectroscopy analysis of a) pure HA and b) robocasted PCL-HA (PLA-HA had a similar peak profile). All scaffolds had evidence of

calcium and phosphorous elemental composition within their matrices.

Figure 3. X-ray diffraction patterns of a) pure HA and b) robocasted PCL-HA (PLA-HA had a same peak profile). In each figure, the top pattern is the sample pattern and the bottom pattern is the powder diffraction file for HA. No crystallinity was observed for the polymers. Note the random orientation of crystallites in each sample.

Figure 4. Comparison of osteoblast proliferation on 2D and 3D scaffolds prepared by robocasting: Cell densities are reported with proliferation studied over 7 d. Control cell proliferation was approximately 5x initial seeding density (50 000 cells cm⁻²).

Figure 5. Comparison of osteoblast proliferation on 3D scaffolds and HA: Cell densities are reported with proliferation studied over 7 d. Control cell proliferation was approximately 5x initial seeding density (50 000 cells cm⁻²).

Figure 6. Electron micrograph of MC3T3-E1.4 layer on robocasted PCL-HA. Cell layers (white) on PCL-HA 3D scaffold (dark gray) at A) 100x, B) 5000x, with individual cells shown to be adherent (C) 2000x and D) 1000x). (O - osteoblast or osteoblast layers, L - lamellipodia, F - filopodia, S - scaffold).

Figure 7. Electron micrograph of MC3T3-E1.4 layer on robocasted PLA-HA. Overall views of cell layers (white) on PLA-HA 3D scaffold (dark gray) at A) 100x, B) 2000x, C) 5000x. Individual cells were seen attached to the PLA-HA surface at D) 1000x. (O - osteoblast or osteoblast layers, L - lamellipodia, F - filopodia, S - scaffold, PD - polymer degradation).

DEXTRAN COATED CERIUM OXIDE NANOPARTICLES FOR INHIBITING BONE CANCER CELL FUNCTIONS

Figure 1: Schematic of the synthesis and characterization of dextran coated ceria nanoparticles and cell culture experiments.

Figure 2: Figure 2a) TEM micrograph of 0.1M (left) and 0.01M (right) dextran coated nanoceria; b) X-ray diffraction of 0.1M and 0.01 M dextran coated nanoparticles; and c) X-ray photoelectron spectra.

Figure 3: Toxicity effect of 0.1 M and 0.01M dextran coated ceria nanoparticles (DCN) against to osteosarcoma cells at various concentrations after 1-day treatment. Data = mean +/- SEM and *p < 0.01 compared to control at the same DCN concentration.

Figure 4: Toxicity effect of 0.1 M dextran coated nanoparticles (DCN) against osteosarcoma cells at various concentrations after 1, 3 and 5 days treatment. Data = mean +/- SEM and *p < 0.01 compared to control at the same DCN concentration.

Preface

This volume is a collection of research papers from the Next Generation Biomaterials and Surface Properties of Biomaterials symposia, which took place during the Materials Science & Technology 2014 Conference & Exhibition (MS&T'14), at the David L. Lawrence Convention Center, Pittsburgh, Pennsylvania. These symposia focused on several key areas, including biomaterials for tissue engineering, ceramic biomaterials, metallic biomaterials, biomaterials for drug delivery, nanostructured biomaterials, biomedical coatings, and surface modification technologies.

We would like to thank the following symposium organizers for their valuable assistance: Kalpana Katti, North Dakota State University; Mukesh Kumar, Biomet Inc; Kajal Mallick, University of Warwick; Sharmila Mukhopadhyay, Wright State University; Vilupanur Ravi, California State Polytechnic University, Pomona; and Varshni Singh, Louisiana State University. Thanks also to all of the authors, participants, and reviewers of this *Ceramic Transactions* proceedings issue.

We hope that this issue becomes a useful resource in the area of biomaterials research that not only contributes to the overall advancement of this field but also signifies the growing roles of The American Ceramic Society and its partner materials societies in this rapidly developing field.

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Next Generation Bioceramics
