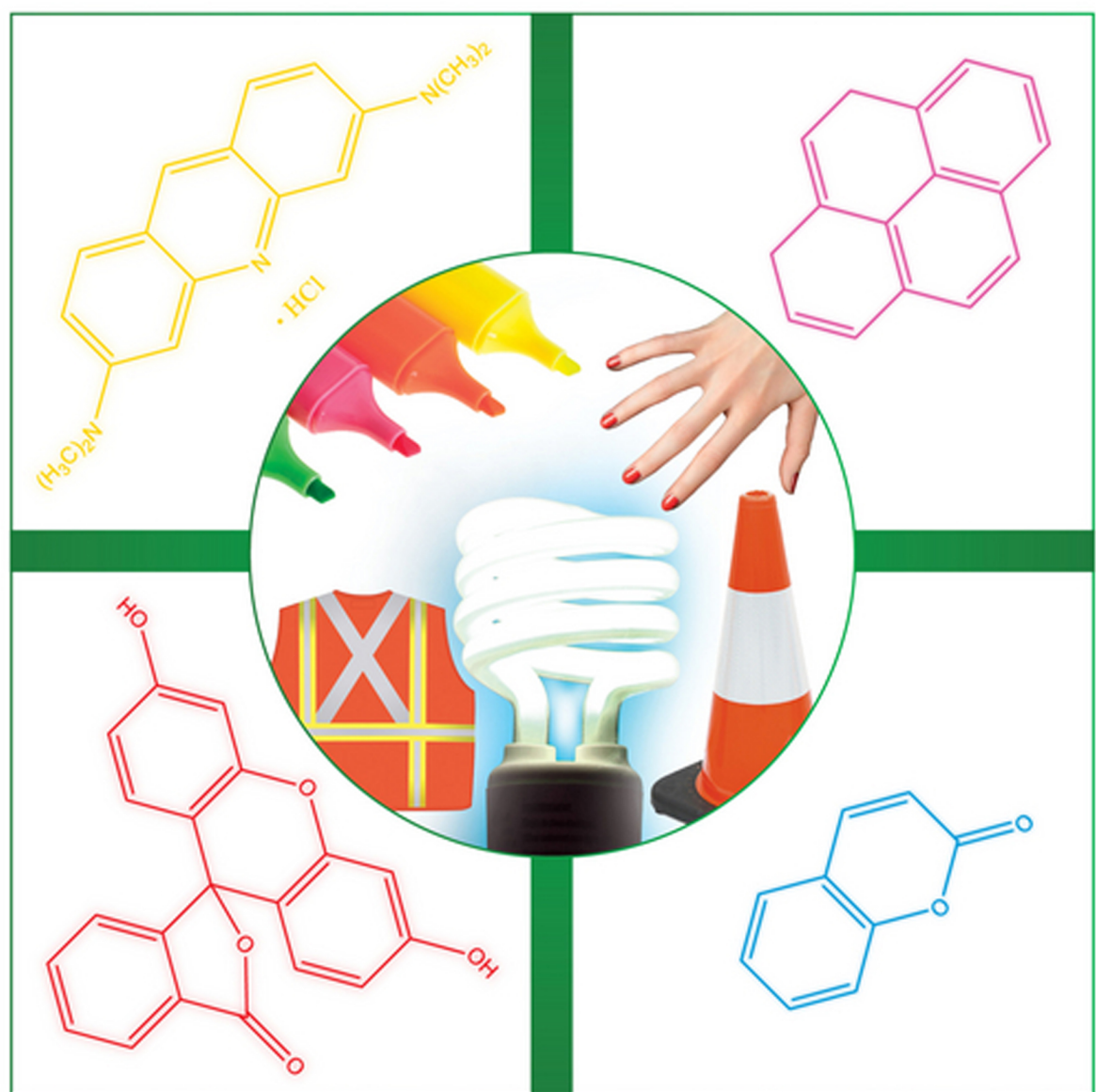


HANDBOOK OF FLUORESCENT DYES AND PROBES



R. W. SABNIS

WILEY

HANDBOOK OF FLUORESCENT DYES AND PROBES

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R. W. Sabnis, Ph.D.

WILEY

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1 2015

Dedicated To
My Wife
Mrs. Madhuri R. Sabnis

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Preface

Fluorescence has been a fascination for individuals for a long time. This book is intended as a reference guide on fluorescent dyes used in medicine, life science, imaging science, cell biology, labeling technology, clinical science, biological chemistry, dye chemistry, biological staining, color chemistry, environmental science, forensic science, organic chemistry, histochemistry, cytochemistry, medicinal chemistry, adhesives, agriculture, coatings, devices, electronics, petroleum, photography, plastics, polymers, security, textile and toys. There are hundreds of fluorescent dyes reported but this book mainly focuses on those dyes, which are widely used in various industrial and academic research.

There is no book available in the market directly on fluorescent dyes and probes, which provides information (such as CAS Registry Numbers, synthesis, various properties, safety/toxicity data and a wide variety of applications) in one source, even though use of fluorescent dyes is wide spread, growing rapidly and has exploded in the past decade. There was a need to publish a book that provided an immediate incentive for compiling the notes to update the scientific community with the wealth of information on fluorescent dyes and probes. To remedy this situation, we have undertaken an ambitious and monumental task of assembling in one publication all the critical data relevant in the field of fluorescent dyes. The dyestuff literature, particularly on fluorescent dyes, is largely in patents. The book provides systematic and up-to-date library of information on 150+ fluorescent dyes and probes as a reference handbook. The book is compiled as a resource guide for chemist and non-chemist in industry as well as in university.

Apart from supplying specific data, the comprehensive, interdisciplinary and comparative nature of the book will provide the user with an easy overview of the state of the art, pinpointing the gaps in the fluorescent dyes knowledge and providing a basis for further research. In addition, it will enable the researcher to use the book in most facile and user-friendly manner.

Fluorescent dyes and probes are arranged alphabetically by the most commonly used name. Again, the choice of primary name is somewhat arbitrary, but an effort has been made to strike a balance between names

that are easily recognizable and names that are chemically informative. The detail information of each fluorescent dye or probe is covered in the following order: CAS registry number, chemical structure, CA index name, other names, Merck index number (Merck Index 15th Edition, 2013), chemical/dye class, molecular formula, molecular weight, physical form, solubility, melting point, boiling point, pK_a , absorption (λ_{max}), emission (λ_{max}), molar extinction coefficient, quantum yield, synthesis, imaging/labeling applications, biological/medical applications, industrial applications, safety/toxicity and references. Where there are discrepancies between different values, the author used his judgment on selecting the most likely value.

Numerous recent references have been provided on various synthetic methods, imaging/labeling applications, biological/medical applications, industrial applications and safety/toxicity data. Space and format limitations prevent giving all the references for each dye. This is the first ever book which provides safety/toxicity data with reference to acute toxicity, aquatic toxicity, carcinogenicity, cytotoxicity, chronic toxicity, ecotoxicity, genotoxicity, hematotoxicity, hepatotoxicity, immunotoxicity, microbial toxicity, mutagenicity, nephrotoxicity, neurotoxicity, nucleic acid damage, oral toxicity, phototoxicity, phytotoxicity, skin toxicity, reproductive toxicity, and so on.

Several appendixes have been provided at the end of the book for scientists to conveniently and easily find a dye as per their need. These appendixes include CAS Registry Numbers, Acridines, Anthracenes, Boron co-ordination compounds/dyes, Coumarins, Cyanines/Styryls, Heterocycles, Pyrenes and Xanthenes.

Omissions as well as errors of fact and interpretation are inevitable in dealing with so vast a subject as fluorescent dyes. I shall be glad to have my attention drawn to errors and to incorporate suggestions for improvement when a revision becomes possible.

I express my profound respect and appreciation to my Guru/Mentor/Advisor, Prof. D. W. Rangnekar, who brought me to this wonderful world of Color Science in the Department of Dyestuffs Technology, Institute of Chemical Technology (ICT), where I laid the foundation stone for my research career in Dye Chemistry.

It is a pleasure to make grateful acknowledgement to Dr. Alan Fanta, Dr. Doina Ene, and Dr. Jeffrey Talkington for extremely useful discussions, encouragement and inspiration.

Words are inadequate to express my sincere appreciation to my wife Madhuri and daughter Anika. It would not have been possible to write this book without

their encouragement and patience. It is a great pleasure to express my gratitude and appreciation to John Wiley & Sons Inc., for giving me an opportunity to write this book.

R. W. SABNIS

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About the Author



Ram W. Sabnis is a Senior Patent Agent in USA. His interests include dyes, pigments, organic chemistry, heterocycles, polymers, synthesis, formulations, coatings and patents. Presently, he focuses on drafting and prosecuting US and international patents. He is a registered patent agent with US Patent and Trademark

Office (USPTO). Prior to entering the legal (patents) field, he was a research chemist for Ascadia, General Electric, Brewer Science, U.S. Textiles and Thermo Fisher (Molecular Probes) in USA. He also worked as a Patent Agent at Squire Patton Boggs L.L.P. and Senior Manager at Pfizer Inc. (Wyeth).

Dr. Sabnis was born and raised in Mumbai, India. He received his Ph.D. in Organic Chemistry (Dye Chemistry) from Institute of Chemical Technology (ICT) (formerly UDCT), Mumbai, India. He is a Chartered Colourists, Fellow of Society of Dyers and Colourists (CCol FSDC).

Dr. Sabnis is one of the world's foremost experts in dyes, inventing world's first colored bubbles

(non-staining) and color changing dye system with many applications. He has immensely contributed to color science and technology for the past 25 years, particularly, dyes for biomedicine, personal care products, health/beauty products, electronics (displays, OLEDs), inks, paints, plastics, textiles and toys/bubbles.

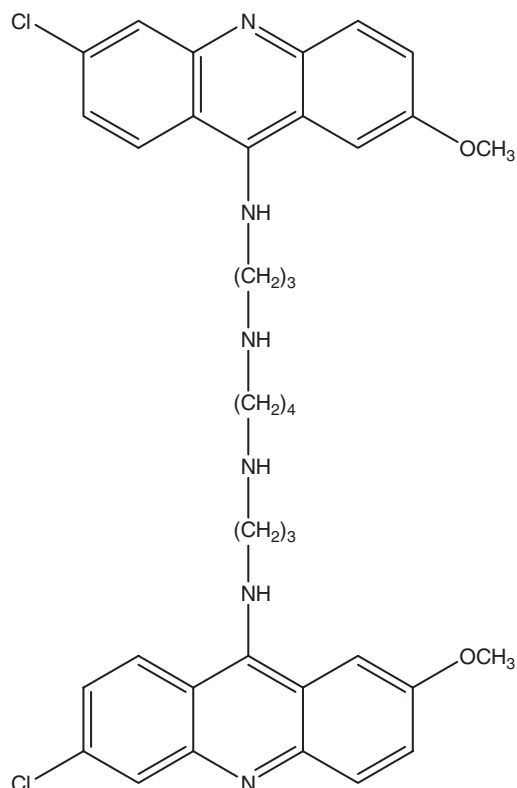
He has over 200 publications which include books, book chapters, encyclopedia chapters, patents, reviews, papers, and symposia presentations. He is also an inventor of several US and international patents (issued/published). Dr. Sabnis is the recipient of Pfish Award, Perkin Innovation Award, Grand Innovation Award, Competitive Spirit Award and Best Doctoral Thesis Award. Dr. Sabnis is awarded the Gold Medal for "Outstanding service to the coloration industry" by the Society of Dyers and Colourists, Bradford, U.K.

He has written two books on color chemistry, namely, "*Handbook of Biological Dyes and Stains*" and "*Handbook of Acid-Base Indicators*." He will continue to focus his activities on fascinating dye chemistry as well as demanding intellectual property in the years to come.

ACRIDINE HOMODIMER

CAS Registry Number 57576-49-5

Chemical Structure



CA Index Name 1,4-Butanediamine, *N,N'*-bis[3-[(6-chloro-2-methoxy-9-acridinyl)amino]propyl]-

Other Names Acridine homodimer; NSC 219743

Merck Index Number Not listed

Chemical/Dye Class Acridine

Molecular Formula C₃₈H₄₂Cl₂N₆O₂

Molecular Weight 685.69

Physical Form Orange-brown powder or yellow solid

Solubility Soluble in water, *N,N*-dimethylformamide, dimethyl sulfoxide, methanol

Melting Point 169–170 °C²

Boiling Point (Calcd.) 885.4 ± 65.0 °C Pressure: 760 Torr

pKa (Calcd.) 10.63±0.19 Most Basic Temperature: 25 °C

Absorption (λ_{max}) 431 nm (H₂O/DNA); 418 nm (MeOH)

Emission (λ_{max}) 498 nm (H₂O/DNA); 500 nm (MeOH)

Molar Extinction Coefficient 12,000 cm⁻¹ M⁻¹ (MeOH)

Synthesis Synthetic methods¹⁻³

Imaging/Labeling Applications Nucleic acids;¹⁻⁹ chromosomes¹⁰

Biological/Medical Applications Detecting nucleic acids;¹⁻⁹ diagnosis and selective tissue necrosis;¹¹ treating cancer,¹¹ malformed proteins causing neurodegenerative disease,¹³ prion disease¹²

Industrial Applications Not reported

Safety/Toxicity Neurotoxicity¹³

REFERENCES

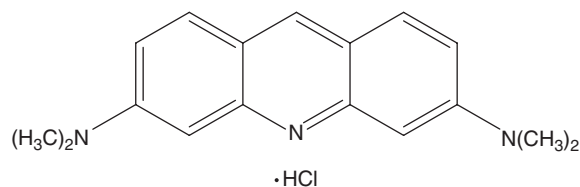
1. Sabnis, R. W. *Handbook of Biological Dyes and Stains*; John Wiley & Sons Inc.: Hoboken, **2010**; pp 3–4.
2. Canellakis, E. S.; Shaw, Y. H.; Hanners, W. E.; Schwartz, R. A. Diacridines: Bifunctional intercalators. I. Chemistry, physical chemistry and growth inhibitory properties. *Biochim. Biophys. Acta* **1976**, 418, 277–289.
3. Barbet, J.; Roques, B. P.; Le Pecq, J. B. Compounds from polyintercalating DNA. Synthesis of acridine dimers. *Compt. Rend. Seances Acad. Sci., Ser. D* **1975**, 281, 851–853.
4. Park, H. O.; Kim, H. B.; Chi, S. M. Detection method of DNA amplification using probes labeled with intercalating dye. PCT Int. Appl. WO 2006004267, 2006.
5. Markovits, J.; Garbay-Jaureguiberry, C.; Roques, B. P.; Le Pecq, J. B. Acridine dimers: influence of the intercalating ring and of the linking-chain nature on the equilibrium and kinetic DNA-binding parameters. *Eur. J. Biochem.* **1989**, 180, 359–366.
6. Bottiroli, G.; Giordano, P.; Prosperi, E. Fluorescent probes in nucleic acid research. *Acta Histochem., Suppl.* **1982**, 26, 189–194.

7. Bottiroli, G.; Giordano, P.; Doglia, S.; Cionini, P. G. Employment of bis-intercalating dyes for the “*in situ*” study of DNA composition. *Basic Appl. Histochem.* **1979**, *23*, 59–63.
8. Le Bret, M.; Le Pecq, J. B.; Barbet, J.; Roques, B. P. A reexamination of the problem of resonance energy transfer between DNA intercalated chromophores using bisintercalating compounds. *Nucleic Acids Res.* **1977**, *4*, 1361–1379.
9. Le Pecq, J. B.; Le Bret, M.; Barbet, J.; Roques, B. DNA polyintercalating drugs. DNA binding of diacridine derivatives. *Proc. Natl. Acad. Sci. U.S.A.* **1975**, *72*, 2915–2919.
10. Van de Sande, J. H.; Lin, C. C.; Deugau, K. V. Clearly differentiated and stable chromosome bands produced by a spermine bis-acridine, a bifunctional intercalating analog of quinacrine. *Exp. Cell Res.* **1979**, *120*, 439–444.
11. Mills, R. L. Pharmaceuticals and apparatus based on Moessbauer isotopic resonant absorption of γ emission (MIRAGE) providing diagnosis and selective tissue necrosis. Can. Pat. Appl. CA 2005039, 1991.
12. May, B. C. H.; Fafarman, A. T.; Hong, S. B.; Rogers, M.; Deady, L. W.; Prusiner, S. B.; Cohen, F. E. Potent inhibition of scrapie prion replication in cultured cells by bis-acridines. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 3416–3421.
13. Prusiner, S. B.; Korth, C.; May, B. C. H. Cyclic bis-compounds clearing malformed proteins. U.S. Pat. Appl. Publ. US 2004229898, 2004.

ACRIDINE ORANGE (AO)

CAS Registry Number 65-61-2

Chemical Structure



CA Index Name 3,6-Acridinediamine, N^3,N^3,N^6,N^6 -tetramethyl-, hydrochloride (1:1)

Other Names 3,6-Acridinediamine, N,N,N',N' -tetramethyl-, monohydrochloride; Acridine Orange R; Acridine, 3,6-bis(dimethylamino)-, hydrochloride; Acridine, 3,6-bis(dimethylamino)-, monohydrochloride; 3,6-Bis(dimethylamino)acridine hydrochloride; Acridine Orange; AO; Acridine Orange N; Acridine Orange NO; Acridine Orange NS; Basic Orange 14; Basic Orange 3RN; C.I. 46005; C.I. Basic Orange 14; Rhoduline Orange NO; Sumitomo Acridine Orange NO; Sumitomo AcridineOrange RK conc

Merck Index Number Not listed

Chemical/Dye Class Acridine

Molecular Formula $C_{17}H_{20}ClN_3$

Molecular Weight 301.82

Physical Form Orange solid

Solubility Soluble in water, dimethyl sulfoxide, ethanol, methanol

Absorption ($\lambda_{\max}$) 500 nm (H_2O/DNA); 460 nm (H_2O/RNA); 489 nm (MeOH)

Emission ($\lambda_{\max}$) 526 nm (H_2O/DNA); 650 nm (H_2O/RNA); 520 nm (MeOH)

Molar Extinction Coefficient $53,000\text{ cm}^{-1}\text{ M}^{-1}$ (H_2O/DNA); $64,000\text{ cm}^{-1}\text{ M}^{-1}$ (MeOH)

Synthesis Synthetic methods¹⁻⁸

Imaging/Labeling Applications Bacteria;⁹⁻¹⁵ blood smears;¹⁶⁻¹⁸ casein;¹⁹ cells/tissues;²⁰⁻²³ chromosomes;^{24,25} endospores;²⁶ lignin;^{27,107} liposomes;²⁸ lysosomes;²⁹⁻³⁵ micronucleus;³⁶⁻³⁸ microorganisms;³⁹⁻⁴⁹ mucin;⁵⁰ nuclei;⁵¹ nucleic acids;⁵²⁻⁷⁴ parasites;⁷⁵⁻⁷⁹ sperms;^{80,81} tumors;^{82,83} yeast⁸⁴⁻⁸⁸

Biological/Medical Applications Analyzing/counting/measuring microorganisms;³⁹⁻⁴⁹ analyzing/detecting/identifying nucleic acids;⁵²⁻⁷⁴ counting/detecting cells/tissues;²⁰⁻²³ detecting parasites;⁷⁵⁻⁷⁹ measuring phagosome-lysosome fusion;^{31,33} for photodynamic therapy;^{82,83} monitoring atmospheric/indoor bioaerosols;^{89,90} apoptosis assay;⁹¹⁻⁹⁷ cytotoxicity assay;^{98,99} genotoxicity assay;⁹⁹ as temperature sensor;^{100,101} dental materials for crowns and bridges¹⁰²

Industrial Applications Adhesives;¹⁰³ aluminophosphate crystalline materials;¹⁰⁴ detecting clay particles;¹⁰⁵ display device;¹⁰⁶ evaluating fiber surface characteristics;¹⁰⁷ glass matrixes;¹⁰⁸ imaging material;¹⁰⁹ inks;^{110,111} lasers;¹¹² recording materials;¹¹³⁻¹¹⁵ photoresists;^{116,117} textiles;¹¹⁸ thin films;^{119,120} tracers for hydrology;¹²¹ wiring boards¹²²

Safety/Toxicity Carcinogenicity;¹²³⁻¹²⁵ cytotoxicity;^{126,127} DNA damage;¹²⁸ embryotoxicity;¹²⁹ genotoxicity;¹³⁰⁻¹³⁴ mutagenicity;¹³⁵⁻¹³⁸ photodynamic toxicity;¹³⁹ phototoxicity¹⁴⁰⁻¹⁴²

REFERENCES

1. Sabnis, R. W. *Handbook of Biological Dyes and Stains*; John Wiley & Sons Inc.: Hoboken, **2010**; pp 5-7.
2. Albert, A. *The Acridines: Their Preparation, Physical, Chemical, and Biological Properties and Uses*; St. Martin's Press: New York, **1966**; p 113.
3. Acheson, R. M. *Acridines*; Interscience Publishers, Inc.: New York, **1956**; pp 26-35.
4. Glushko, V. N.; Parbuzina, I. L.; Petrova, G. S. Acridine orange hydrochloride. *Khim. Promyshl., Ser.: Reakt. Osobo Chistye Veshch.* **1980**, 3-4.
5. Glushko, V. N.; Parbuzina, I. L.; Petrova, G. S. 3,6-Bis(dimethylamino)acridine hydrochloride (Acridine Orange). U.S.S.R. SU 694525, 1979.
6. Albert, A. Acridine syntheses and reactions. III. Synthesis of aminoacridines from formic acid and amines. *J. Chem. Soc.* **1947**, 244-250.
7. Karr, A. E. Acridine oranges. *Text. Colorist* **1940**, 62, 604-607, 634, 676-679, 763-767, 836-837, 852.
8. Biehringer, J. Ueber die farbstoffe der pyroningruppe. *J. Prakt. Chem.* **1897**, 54, 217-258
9. Araki, H.; Okasawa, Y.; Kaneno, M. Acid-fast bacteria acridine orange fluorescent staining

- method. Jpn. Kokai Tokkyo Koho JP 2004069341, 2004.
10. Rychlik, I.; Cardova, L.; Sevcik, M.; Barrow, P. A. Flow cytometry characterization of *Salmonella typhimurium* mutants defective in proton translocating proteins and stationary-phase growth phenotype. *J. Microbiol. Methods* **2000**, *42*, 255–263.
11. Yano, R.; Nogami, T. A method and an apparatus for detecting live bacteria using fluorescence-labeled bacteriophage. Jpn. Kokai Tokkyo Koho JP 11318499, 1999.
12. Back, J. P.; Kroll, R. G. The differential fluorescence of bacteria stained with acridine orange and the effects of heat. *J. Appl. Bacteriol.* **1991**, *71*, 51–58.
13. Maki, J. S.; LaCroix, S. J.; Hopkins, B. S.; Staley, J. T. Recovery and diversity of heterotrophic bacteria from chlorinated drinking waters. *Appl. Environ. Microbiol.* **1986**, *51*, 1047–1055.
14. Bergstrom, I.; Heinanen, A.; Salonen, K. Comparison of acridine orange, acriflavine, and bisbenzimidazole stains for enumeration of bacteria in clear and humic waters. *Appl. Environ. Microbiol.* **1986**, *51*, 664–667.
15. Meseguer, M.; de Rafael, L.; Baquero, M.; Martinez, F. M.; Lopez-Brea, M. Acridine orange stain in the early detection of bacteria in blood cultures. *Eur. J. Clin. Microbiol.* **1984**, *3*, 113–115.
16. Kvetnaya, A. S.; Zhelezova, L. I. Prognosis of clinical course of acute intestinal infection in child based on evaluation of functional state of polymorphonuclear leukocytes. Russ. RU 2275634, 2006.
17. Sciotto, C. G.; Lauer, B. A.; White, W. L.; Istre, G. R. Detection of *Borrelia* in acridine orange-stained blood smears by fluorescence microscopy. *Arch. Pathol. Lab. Med.* **1983**, *107*, 384–386.
18. Mirrett, S.; Lauer, B. A.; Miller, G. A.; Reller, L. B. Comparison of acridine orange, methylene blue, and gram stains for blood cultures. *J. Clin. Microbiol.* **1982**, *15*, 562–566.
19. Das, M.; Roy, B. R.; Chattoraj, D. K. Binding of dyes to casein. *Indian J. Technol.* **1986**, *24*, 95–100.
20. Skyggebjerg, O.; Glensbjerg, M. A method and a system for counting cells from a plurality of species. PCT Int. Appl. WO 2002101087, 2002.
21. Foglieni, C.; Meoni, C.; Davalli, A. M. Fluorescent dyes for cell viability: an application on prefixed conditions. *Histochem. Cell Biol.* **2001**, *115*, 223–229.
22. Bartzatt, R. Acridine orange staining I. *In vitro* derived cells lines. *J. Histotechnol.* **1987**, *10*, 91–93.
23. Schmitz-Moormann, P. Tissue staining by basic dyes. I. Influence of the pH of the staining solution and of the dye-affinity on the adsorption of the dye. *Histochemie* **1968**, *16*, 23–35.
24. Lin, C. C.; Jorgenson, K. F.; Van de Sande, J. H. Specific fluorescent bands on chromosomes produced by acridine orange after prestaining with base specific non-fluorescent DNA ligands. *Chromosoma* **1980**, *79*, 271–286.
25. Forabosco, A.; Couturier, J.; Dutrillaux, B. Effects of pH on the staining of human chromosomes with acridine orange. *Exp. Cell Res.* **1974**, *88*, 418–421.
26. Schichnes, D.; Nemson, J. A.; Ruzin, S. E. Fluorescent staining method for bacterial endospores. *Microscope* **2006**, *54*, 91–93.
27. Perdih, F.; Perdih, A. Lignin selective dyes: quantum-mechanical study of their characteristics. *Cellulose* **2011**, *18*, 11392–1150.
28. Van Rooijen, N.; Van Nieuwmegen, R. Fluorochrome staining of multilamellar liposomes. *Stain Technol.* **1978**, *53*, 307–310.
29. Krolenko, S. A.; Adamyan, S. Ya.; Belyaeva, T. N.; Mozhenok, T. P. Acridine orange bioaccumulation in acid organelles of normal and vacuolated frog skeletal muscle fibres. *Cell Biol. Int.* **2006**, *30*, 933–939.
30. Traganos, F.; Darzynkiewicz, Z. Lysosomal proton pump activity: Supravital cell staining with acridine orange differentiates leukocyte subpopulations. *Methods Cell Biol.* **1994**, *41*, 185–194.
31. Steinberg, T. H.; Swanson, J. A. Measurement of phagosome-lysosome fusion and phagosomal pH. *Methods Enzymol.* **1994**, *236*, 147–160.
32. Rashid, F.; Horobin, R. W.; Williams, M. A. Predicting the behavior and selectivity of fluorescent probes for lysosomes and related structures by means of structure-activity models. *Histochem. J.* **1991**, *23*, 450–459.
33. Kielian, M. Assay of phagosome-lysosome fusion. *Methods Enzymol.* **1986**, *132*, 257–267.
34. Moriyama, Y.; Takano, T.; Ohkuma, S. Acridine orange as a fluorescent probe for lysosomal proton pump. *J. Biochem.* **1982**, *92*, 1333–1336.
35. Wilson, C. L.; Jumper, G. A.; Mason, D. L. Acridine orange as a lysosome marker in fungal spores. *Phytopathology* **1978**, *68*, 1564–1567.

36. Polard, T.; Jean, S.; Merlina, G.; Laplanche, C.; Pinelli, E.; Gauthier, L. Giemsa versus acridine orange staining in the fish micronucleus assay and validation for use in water quality monitoring. *Ecotoxicol. Environ. Saf.* **2011**, *74*, 144–149.
37. Nersesyan, A.; Kundi, M.; Atefie, K.; Schulte-Hermann, R.; Knasmueller, S. Effect of staining procedures on the results of micronucleus assays with exfoliated oral mucosa cells. *Cancer Epidemiol., Biomarkers Prev.* **2006**, *15*, 1835–1840.
38. Hayashi, M.; Sofuni, T.; Ishidate, M., Jr., An application of acridine orange fluorescent staining to the micronucleus test. *Mutat. Res. Lett.* **1983**, *120*, 241–247.
39. Li, C. S.; Chia, W. C.; Chen, P. S. Fluorochrome and flow cytometry to monitor microorganisms in treated hospital wastewater. *J. Environ. Sci. Health, Part A* **2007**, *42*, 195–203.
40. Horikiri, S. Microorganism cell detection method using multiple fluorescent indicators. Jpn. Kokai Tokkyo Koho JP 2006238779, 2006.
41. Noda, N.; Mizutani, T. Microorganism-measuring method using multiple staining. Jpn. Kokai Tokkyo Koho JP 2006340684, 2006.
42. Chen, P.; Li, C. Real-time quantitative PCR with gene probe, fluorochrome and flow cytometry for microorganism analysis. *J. Environ. Monit.* **2005**, *7*, 257–262.
43. Besson, F. I.; Hermet, J. P.; Ribault, S. Reaction medium and process for universal detection of microorganisms. Fr. Demande FR 2847589, 2004.
44. Sunamura, T.; Maruyama, A.; Kurane, R. Method for detecting and counting microorganism. Jpn. Kokai Tokkyo Koho JP 2002291499, 2002.
45. Giorgio, A.; Rambaldi, M.; Maccario, P.; Ambrosone, L.; Moles, D. A. Detection of microorganisms in clinical specimens using slides prestained with acridine orange (AOS). *Microbiologica* **1989**, *12*, 97–100.
46. Tsuji, T.; Karasawa, M. Method for counting of microorganisms. Jpn. Kokai Tokkyo Koho JP 60210997, 1985.
47. Lauer, B. A.; Reller, L. B.; Mirrett, S. Comparison of acridine orange and gram stains for detection of microorganisms in cerebrospinal fluid and other clinical specimens. *J. Clin. Microbiol.* **1981**, *14*, 201–205.
48. McCarthy, L. R.; Senne, J. E. Evaluation of acridine orange stain for detection of microorganisms in blood cultures. *J. Clin. Microbiol.* **1980**, *11*, 281–285.
49. Scholefield, J. Staining microorganisms. Ger. Offen. DE 2728077, 1978.
50. Hicks, J. D.; Matthaie, E. A selected fluorescence stain for mucin. *J. Pathol. Bacteriol.* **1958**, *75*, 473–476.
51. Horobin, R. W.; Stockert, J. C.; Rashid-Doubell, F. Fluorescent cationic probes for nuclei of living cells: Why are they selective? A quantitative structure-activity relations analysis. *Histochem. Cell Biol.* **2006**, *126*, 165–175.
52. Kjaerulff, S.; Glensbjerg, M. Method for analysis of cellular DNA content. PCT Int. Appl. WO 2011098085, 2011.
53. Miyamoto, S.; Kato, T.; Tomono, J. Nucleic acid identification method. Jpn. Kokai Tokkyo Koho JP 2010233530, 2010.
54. Lai, S.; Chang, X.; Tian, L.; Wang, S.; Bai, Y.; Zhai, Y. Fluorometric determination of DNA using nano-SiO₂ particles as an effective dispersant and stabilizer for acridine orange. *Microchim. Acta* **2007**, *156*, 225–230.
55. Bi, S.; Qiao, C.; Song, D.; Tian, Y.; Gao, D.; Sun, Y.; Zhang, H. Study of interactions of flavonoids with DNA using acridine orange as a fluorescence probe. *Sens. Actuators, B: Chem.* **2006**, *B119*, 199–208.
56. Martens-Habbena, W.; Sass, H. Sensitive determination of microbial growth by nucleic acid staining in aqueous suspension. *Appl. Environ. Microbiol.* **2006**, *72*, 87–95.
57. Park, H. O.; Kim, H. B.; Chi, S. M. Detection method of DNA amplification using probes labeled with intercalating dye. PCT Int. Appl. WO 2006004267, 2006.
58. Ovadekova, R.; Jantova, S.; Labuda, J. Detection of the effective DNA protection by quinazolines using a DNA-based electrochemical biosensor. *Anal. Lett.* **2005**, *38*, 2625–2638.
59. El-Naggar, A. K. Concurrent flow cytometric analysis of DNA and RNA. *Methods Mol. Biol.* **2004**, *263*, 371–384.
60. Lauretti, F.; Lucas de Melo, F.; Benati, F. J.; de Mello, V. E.; Santos, N.; Linhares, R. E. C.; Nozawa, C. Use of acridine orange staining for the detection of rotavirus RNA in polyacrylamide gels. *J. Virol. Methods* **2003**, *114*, 29–35.
61. Tomita, N.; Mori, Y. Method for efficiently detecting double-stranded nucleic acid. PCT Int. Appl. WO 2002103053, 2002.

62. Gonzalez, K.; McVey, S.; Cunnick, J.; Udovichenko, I. P.; Takemoto, D. J. Acridine orange differential staining of total DNA and RNA in normal and galactosemic lens epithelial cells in culture using flow cytometry. *Curr. Eye Res.* **1995**, *14*, 269–273.
63. Evenson, D.; Darzynkiewicz, Z.; Jost, L.; Janca, F.; Ballachey, B. Changes in accessibility of DNA to various fluorochromes during spermatogenesis. *Cytometry* **1986**, *7*, 45–53.
64. Villanueva, A.; Stockert, J. C.; Armas-Portela, R. A simple method for the fluorescence analysis of nucleic acid-dye complexes in cytological preparations. *Histochemistry* **1984**, *81*, 103–104.
65. Wallen, C. A.; Higashikubo, R.; Dethlefsen, L. A. Comparison of two flow cytometric assays for cellular RNA - acridine orange and propidium iodide. *Cytometry* **1982**, *3*, 155–160.
66. Curtis, S. K.; Cowden, R. R. Four fluorochromes for the demonstration and microfluorometric estimation of RNA. *Histochemistry* **1981**, *72*, 39–48.
67. Dutt, M. K. Acridine orange - its use in the specific staining of DNA in mammalian tissue sections. *Microsc. Acta* **1981**, *84*, 37–42.
68. Carmichael, G. G.; McMaster, G. K. The analysis of nucleic acids in gels using glyoxal and acridine orange. *Methods Enzymol.* **1980**, *65*, 380–391.
69. Taylor, I. W.; Milthorpe, B. K. An evaluation of DNA fluorochromes, staining techniques, and analysis for flow cytometry. I. Unperturbed cell populations. *J. Histochem. Cytochem.* **1980**, *28*, 1224–1232.
70. Coulson, P. B.; Bishop, A. O.; Lenarduzzi, R. Quantitation of cellular deoxyribonucleic acid by flow microfluorometry. *J. Histochem. Cytochem.* **1977**, *25*, 1147–1153.
71. Tomita, G. Molecular complexes of acridine orange and nucleosides. *Biophysik* **1967**, *4*, 118–128.
72. Yamabe, S. A spectrophotometric study on binding of acridine orange with DNA. *Mol. Pharmacol.* **1967**, *3*, 556–560.
73. Kasten, F. H. Cytochemical studies with acridine orange and the influence of dye contaminants in the staining of nucleic acids. *Int. Rev. Cytol.* **1967**, *21*, 141–202.
74. Lecatsas, G. Detection of plant nucleic acids with acridine orange. *S. Afr. J. Sci.* **1967**, *63*, 61.
75. Xu, L.; Chaudhuri, A. *Plasmodium yoelii*: a differential fluorescent technique using acridine orange to identify infected erythrocytes and reticulocytes in Duffy knockout mouse. *Exp. Parasitol.* **2005**, *110*, 80–87.
76. Saito-Ito, A.; Akai, Y.; He, S.; Kimura, M.; Kawabata, M. A rapid, simple and sensitive flow cytometric system for detection of *Plasmodium falciparum*. *Parasitol. Int.* **2001**, *50*, 249–257.
77. Freedman, D. O.; Berry, R. S. Rapid diagnosis of *Bancroftian filariasis* by acridine orange staining of centrifuged parasites. *Am. J. Trop. Med. Hyg.* **1992**, *47*, 787–793.
78. Rickman, L. S.; Long, G. W.; Oberst, R.; Cabanban, A.; Sangalang, R.; Smith, J. I.; Chulay, J. D.; Hoffman, S. L. Rapid diagnosis of malaria by acridine orange staining of centrifuged parasites. *Lancet* **1989**, *1*, 68–71.
79. Wongsrichanalai, C.; Webster, H. K.; Brown, A. E. Rapid diagnosis of malaria by acridine orange staining of centrifuged parasites. *Lancet* **1989**, *1*, 967.
80. Yaniz, Y. L.; Palacin, I.; Vicente-Fiel, S.; Gosalvez, J.; Lopez-Fernandez, C.; Santolaria, P. Comparison of membrane-permeant fluorescent probes for sperm viability assessment in the ram. *Reprod. Dom. Anim.* **2013**, *48*, 598–603.
81. Chohan, K. R.; Griffin, J. T.; Lafromboise, M.; De Jonge, C. J.; Carrell, D. T. Comparison of chromatin assays for DNA fragmentation evaluation in human sperm. *J. Androl.* **2006**, *27*, 53–59.
82. Matsubara, T.; Kusuzaki, K.; Matsumine, A.; Shintani, K.; Satonaka, H.; Uchida, A. Acridine orange used for photodynamic therapy accumulates in malignant musculoskeletal tumors depending on pH gradient. *Anticancer Res.* **2006**, *26*, 187–193.
83. Ueda, H.; Murata, H.; Takeshita, H.; Minami, G.; Hashiguchi, S.; Kubo, T. Unfiltered xenon light is useful for photodynamic therapy with acridine orange. *Anticancer Res.* **2005**, *25*, 3979–3983.
84. Muro, K.; Izumi, K. Method for evaluating yeast activity. Jpn. Kokai Tokkyo Koho JP 2004357618, 2004.
85. Bogen, H. J.; Elste, U. Physiology of the effect of acridine orange (AO). Studies of yeast cells. *Planta* **1955**, *45*, 325–375.
86. Bogen, H. J. Staining, damaging, and destroying of yeast cells by acridine orange. *Arch. Mikrobiol.* **1953**, *18*, 170–197.
87. Kolbel, H. Quantitative investigation of acridine orange uptake of living and dead yeast cells and correlation with electrical conditions of the cell. *Z. Naturforsch.* **1947**, *26*, 382–392.

88. Strugger, S. Investigations on the vital fluorochroming (fluorostaining) of yeast cells. *Flora* **1943**, *37*, 73–94.
89. Chi, M.; Li, C. Fluorochrome in monitoring atmospheric bioaerosols and correlations with meteorological factors and air pollutants. *Aerosol Sci. Technol.* **2007**, *41*, 672–678.
90. Li, C.; Huang, T. Fluorochrome in monitoring indoor bioaerosols. *Aerosol Sci. Technol.* **2006**, *40*, 237–241.
91. Verduzco, D.; Amatruda, J. F. Analysis of cell proliferation, senescence, and cell death in zebrafish embryos. *Methods Cell Biol.* **2011**, *101*, 19–38.
92. Alvarez, M.; Villanueva, A.; Acedo, P.; Canete, M.; Stockert, J. C. Cell death causes relocation of photosensitizing fluorescent probes. *Acta Histochem.* **2011**, *113*, 363–368.
93. Kajstura, M.; Halicka, H. D.; Pryjma, J.; Darzynkiewicz, Z. Discontinuous fragmentation of nuclear DNA during apoptosis revealed by discrete "sub-G1" peaks on DNA content histograms. *Cytometry, Part A* **2007**, *71A*, 125–131.
94. Hiruma, H.; Katakura, T.; Takenami, T.; Igawa, S.; Kanoh, M.; Fujimura, T.; Kawakami, T. Vesicle disruption, plasma membrane bleb formation, and acute cell death caused by illumination with blue light in acridine orange-loaded malignant melanoma cells. *J. Photochem. Photobiol., B: Biol.* **2007**, *86*, 1–8.
95. Baskic, D.; Popovic, S.; Ristic, P.; Arsenijevic, N. N. Analysis of cycloheximide-induced apoptosis in human leukocytes: Fluorescence microscopy using annexin V/propidium iodide versus acridine orange/ethidium bromide. *Cell Biol. Int.* **2006**, *30*, 924–932.
96. Giuliano, M.; Bellavia, G.; Lauricella, M.; D'Anne, A.; Vassallo, B.; Vento, R.; Tesoriere, G. Staurosporine-induced apoptosis in Chang liver cells is associated with down-regulation of Bcl-2 and Bcl-XL. *Int. J. Mol. Med.* **2004**, *13*, 565–571.
97. Loweth, A. C.; Morgan, N. G. Methods for the study of NO-induced apoptosis in cultured cells. *Methods Mol. Biol.* **1998**, *100*, 311–320.
98. Odawara, K. Visible light cytotoxicity expression ability assay method, and its use. Jpn. Kokai Tokkyo Koho JP 2007143465, 2007.
99. Chreno, O. Method for testing cytotoxicity and genotoxicity of chemical substances. Slovakia SK 278857, 1998.
100. Bousseksou, A.; Salmon, L.; Molnar, G.; Cobo, S. Materials with thermochromic spin transition doped with one or more fluorescent agents for use as temperature sensor. Fr. Demande FR 2952371, 2011.
101. Bousseksou, A.; Salmon, L.; Molnar, G.; Cobo, S. Heat-sensitive spin-transition materials doped with one or more fluorescent agents for use as temperature sensor. PCT Int. Appl. WO 2011058277, 2011.
102. Klemm, E.; Hoerhold, H. H.; Doms, I. Light-curing dental materials for crowns and bridges. Ger. (East) DD 215699, 1984.
103. Hoerhold, H. H.; Klemm, E.; Flammersheim, H. J.; Maertin, R.; Wolf, H. Adhesives. Ger. Offen. DE 3431440, 1985.
104. Gandara, F.; Lopez-Arbeloa, F.; Ruiz-Hitzky, E.; Cambor, M. A. "Bottle-around-a-ship" confinement of high loadings of acridine orange in new aluminophosphate crystalline materials. *J. Mater. Chem.* **2006**, *16*, 1765–1771.
105. Jain, R. K.; D'Hoore, J. Detection and determination of clay particles in natural waters. *J. Indian Soc. Soil Sci.* **1982**, *30*, 415–417.
106. Liu, T. Ion-color controlling electrophoresis display device. Faming Zhuanli Shenqing CN 1461967, 2003.
107. Drnovsek, T.; Perdih, A.; Perdih, M. Fiber surface characteristics evaluated by principal component analysis. *J. Wood Sci.* **2005**, *51*, 507–513.
108. Gaponenko, S. V.; Germanenko, I. N.; Stupak, A. P.; Eyal, M.; Brusilovsky, D.; Reinfeld, R.; Graham, S.; Klingshirn, C. Fluorescence of acridine orange in inorganic glass matrixes. *Appl. Phys. B: Lasers Opt.* **1994**, *B58*, 283–288.
109. Namiki, T.; Shinozaki, F.; Ikeda, T. Light-sensitive imaging material. Ger. Offen. DE 2831101, 1979.
110. Ueda, T.; Yasutomi, H. Aqueous ink-jet inks with mildew growth prevention. Jpn. Kokai Tokkyo Koho JP 10007960, 1998.
111. Ishii, K.; Takahashi, H.; Watanabe, K. Water-thinned fluorescent inks for ballpoint pens. Jpn. Kokai Tokkyo Koho JP 06049405, 1994.
112. Volkin, H. C. Direct solar pumped laser. U.S. Patent 4281294, 1981.
113. Ikoma, K.; Miura, K.; Kawade, I.; Oguchi, Y.; Myagawa, M. Optical recording material containing light-emitting dye, and recording method. Jpn. Kokai Tokkyo Koho JP 63062792, 1988.
114. Baumann, R.; Meisel, A.; Singer, W.; Fritzsche, K.; Bauriegel, L. Laser-sensitive information recording material. Ger. (East) DD 223834, 1985.

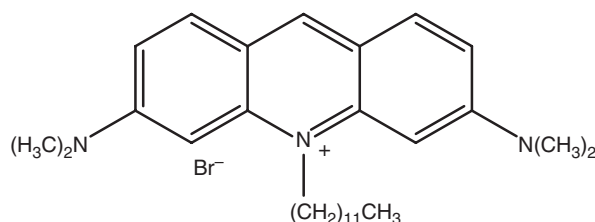
115. Ikegami, K.; Okuyama, H. Holographic recording material and process for producing holograms. Eur. Pat. Appl. EP 0084452, 1983.
116. Endo, M.; Sasako, M.; Tani, Y.; Ogawa, K. Resist compositions containing acridine or its derivatives. Jpn. Kokai Tokkyo Koho JP 01010236, 1989.
117. Farid, S. Y.; Haley, N. F.; Moody, R. E.; Specht, D. P. Negative working photoresists responsive to shorter visible wavelengths and novel coated articles. U.S. Patent 4743529, 1988.
118. Gamblin, R. L. Surfactant enhanced dyeing. U.S. Patent 5593459, 1997.
119. Machida, S.; Wakamatsu, T.; Masuo, S.; Jinnai, H.; Itaya, A. Morphology and photophysical properties of polymer thin films dispersed with dye nanoparticle. *Thin Solid Films* **2008**, *516*, 2615–2619.
120. Li, F.; Pfeiffer, M.; Werner, A.; Harada, K.; Leo, K.; Hayashi, N.; Seki, K.; Liu, X.; Dang, X. D. Acridine orange base as a dopant for n doping of C60 thin films. *J. Appl. Phys.* **2006**, *100*, 023716/1–023716/9.
121. Viriot, M. L.; Andre, J. C. Fluorescent dyes: a search for new tracers for hydrology. *Analisis* **1989**, *17*, 97–111.
122. Mori, K.; Ito, K. Method for pattern formation of metal deposition layers, and manufacture of wiring boards. Jpn. Kokai Tokkyo Koho JP 2007177322, 2007.
123. Schehrer, L.; Regan, J. D.; Westendorf, J. UDS induction by an array of standard carcinogens in human and rodent hepatocytes: effect of cryopreservation. *Toxicology* **2000**, *147*, 177–191.
124. Kowalski, L. A.; Laitinen, A. M.; Martazavi-Asl, B.; Wee, R. K. H.; Erb, H. E.; Assi, K. P.; Madden, Z. *In vitro* determination of carcinogenicity of sixty-four compounds using a bovine papillomavirus DNA-carrying C3H/10T1/2 cell line. *Environ. Mol. Mutagen.* **2000**, *35*, 300–311.
125. Heil, J.; Reifferscheid, G. Detection of mammalian carcinogens with an immunological DNA synthesis-inhibition test. *Carcinogenesis* **1992**, *13*, 2389–2394.
126. Bradburne, C. E.; Delehanty, J. B.; Gemmill, K. B.; Mei, B. C.; Mattoussi, H.; Susumu, K.; Blanco-Canosa, J. B.; Dawson, P. E.; Medintz, I. L. Cytotoxicity of quantum dots used for *in vitro* cellular labeling: Role of QD surface ligand, delivery modality, cell type, and direct comparison to organic fluorophores. *Bioconjugate Chem.* **2013**, *24*, 1570–1583.
127. Chang, Y. S.; Wu, C. L.; Tseng, S. H.; Kuo, P. Y.; Tseng, S. Y. Cytotoxicity of triamcinolone acetonide on human retinal pigment epithelial cells. *Invest. Ophthalmol. Vis. Sci.* **2007**, *48*, 2792–2798.
128. McCarroll, N. E.; Piper, C. E.; Keech, B. H. An *E. coli* microsuspension assay for the detection of DNA damage induced by direct-acting agents and promutagens. *Environ. Mutagen.* **1981**, *3*, 429–444.
129. Kohler, M.; Kundig, A.; Reist, H. W.; Michel, C. Modification of *in vitro* mouse embryogenesis by x-rays and fluorochromes. *Radiat. Environ. Biophys.* **1994**, *33*, 341–351.
130. Mascini, M. Determination of the genotoxicity of aqueous samples by using a DNA-based biosensor and dedicated apparatus. Ital. Appl. IT 2004RM0559, 2005.
131. Gonzalez, N. V.; Soloneski, S.; Larramendy, M. L. Dicamba-induced genotoxicity in Chinese hamster ovary (CHO) cells is prevented by vitamin E. *J. Hazard. Mater.* **2009**, *163*, 337–343.
132. Knight, A. W.; Billinton, N.; Cahill, P. A.; Scott, A.; Harvey, J. S.; Roberts, K. J.; Tweats, D. J.; Keenan, P. O.; Walmsley, R. M. An analysis of results from 305 compounds tested with the yeast RAD54-GFP genotoxicity assay (GreenScreen GC) - including relative predictivity of regulatory tests and rodent carcinogenesis and performance with autofluorescent and colored compounds. *Mutagenesis* **2007**, *22*, 409–416.
133. He, L.; Jurs, P. C.; Custer, L. L.; Durham, S. K.; Pearl, G. M. Predicting the genotoxicity of polycyclic aromatic compounds from molecular structure with different classifiers. *Chem. Res. Toxicol.* **2003**, *16*, 1567–1580.
134. Fernandez, M.; Gauthier, L.; Jaylet, A. Use of newt larvae for *in vivo* genotoxicity testing of water: Results on 19 compounds evaluated by the micronucleus test. *Mutagenesis* **1989**, *4*, 17–26.
135. Lee, I. E.; Nguyen, V. C.; Hayase, F.; Kato, H. Desmutagenicity of melanoidins against various kinds of mutagens and activated mutagens. *Biosci., Biotechnol., Biochem.* **1994**, *58*, 18–23.
136. Klopman, G.; Frierson, M. R.; Rosenkranz, H. S. The structural basis of the mutagenicity of chemicals in *Salmonella typhimurium*: The Gene-Tox data base. *Mutat. Res.* **1990**, *228*, 1–50.
137. Xamena, N.; Creus, A.; Marcos, R. Mutagenic activity of some intercalating compounds in the

- Drosophila zeste* somatic eye mutation test. *Mutat. Res.* **1984**, 138, 169–173.
138. Rogers, A. M.; Back, K. C. Comparative mutagenicity of 4 DNA-intercalating agents in L5178Y mouse lymphoma cells. *Mutat. Res.* **1982**, 102, 447–455.
139. Herkovits, J.; Perez-Coll, C. S.; Stockert, J. C.; Blazquez, A. The screening of photodynamic toxicity of dyes by means of a bioassay using amphibian embryos. *Res. J. Chem. Environ.* **2007**, 11, 86–91.
140. Harris, A. G.; Sinitsina, I.; Messmer, K. Intravital fluorescence microscopy and phototoxicity: effects on leukocytes. *Eur. J. Med. Res.* **2002**, 7, 117–124.
141. Saetzler, R. K.; Jallo, J.; Lehr, H. A.; Philips, C.M.; Vasthare, U.; Arfors, K. E.; Tuma, R. F. Intravital fluorescence microscopy: impact of light-induced phototoxicity on adhesion of fluorescently labeled leukocytes. *J. Histochem. Cytochem.* **1997**, 45, 505–573.
142. Lukasiak-Bachurzevska, B.; Dulczewska-Klopowska, M. Studies on the phototoxic properties of some coal derivatives. *Przegląd Dermatol.* **1981**, 68, 33–37.

ACRIDINE ORANGE 10-DODECYL BROMIDE (DODECYL-ACRIDINE ORANGE (DAO))

CAS Registry Number 41387-42-2

Chemical Structure



CA Index Name Acridinium, 3,6-bis(dimethylamino)-10-dodecyl-, bromide (1:1)

Other Names Acridinium, 3,6-bis(dimethylamino)-10-dodecyl-, bromide; 10-Dodecylacridine Orange Bromide; 3,6-Bis(dimethylamino)-10-dodecylacridinium bromide; AO 10 Dodecylbromide; Acridine orange 10-dodecyl bromide; Dodecyl-Acridine Orange; BDA; D 455; DADAB; DAO

Merck Index Number Not listed

Chemical/Dye Class Acridine

Molecular Formula C₂₉H₄₄BrN₃

Molecular Weight 514.59

Physical Form Orange solid

Solubility Soluble in dimethyl sulfoxide, ethanol, methanol

Melting Point >250 °C

Absorption (λ_{max}) 495 nm (MeOH)

Emission (λ_{max}) 520 nm (MeOH)

Molar Extinction Coefficient 87,000 cm⁻¹ M⁻¹ (MeOH)

Synthesis Synthetic methods¹⁻⁴

Imaging/Labeling Applications Chloride ions;⁵ keratin fibers/hairs;^{6,7} Langmuir-Blodgett (LB) monolayers;²³ micelles;^{2-4,8,9} mitochondrial membranes;¹⁰ proteins¹¹⁻¹⁴

Biological/Medical Applications Analyzing chloride ions;⁵ characterizing drug binding sites on glycoproteins;¹⁴ ophthalmic devices (intraocular lenses (IOL))¹⁵

Industrial Applications Determining cationic surfactants;¹⁶ electroluminescent devices;¹⁷⁻¹⁹ Langmuir-Blodgett films;^{20,23} photographic imaging system;²¹ photoresists;²² semiconductor electrodes;²³⁻²⁵ silica-surfactant composite films²⁶

Safety/Toxicity No data available

REFERENCES

- Haugland, R. P. *Handbook of Fluorescent Probes and Research Chemicals*; Molecular Probes Inc.: Eugene, **1996**; pp 314–316.
- Usui, Y.; Saga, K. The photoreduction and photosensitized reduction of dyes bound to a surfactant micellar surface. *Bull. Chem. Soc. Jpn.* **1982**, 55, 3302–3307.
- Yamagishi, A.; Masui, T.; Watanabe, F. Selective activation of reactant molecules by reversed micelles. *J. Phys. Chem.* **1981**, 85, 281–285.
- Kubota, Y.; Kodama, M.; Miura, M. Second CMC [critical micelle concentration] of an aqueous solution of sodium dodecyl sulfate. IV. Fluorescence depolarization. *Bull. Chem. Soc. Jpn.* **1973**, 46, 100–103.
- Kawabata, Y.; Toge, Y. Fluorometric analysis of chloride ion and chemical sensor therefor. U.S. Patent 5691205, 1997.
- Schaefer, K. Microscopic investigation of the diffusion of dyes in keratin fibers. *Melliand Textilber.* **1993**, 74, E382–E385, 1138, 1141–1142, 1145–1148, 1151–1152.
- Schaefer, K.; Koch, U. Investigation of the diffusion of dyes in keratin fibers by fluorescence microscopy (AiF 8415). *DWI Rep.* **1992**, 109, 699–716.
- Mitsuzuka, M.; Kikuchi, K.; Kokubun, H.; Usui, Y. The primary processes of the photosensitized reduction of methylene blue in an aqueous sodium dodecylsulfate micellar solution. *J. Photochem.* **1985**, 29, 363–373.
- Usui, Y.; Gotou, A. Migration of singlet excitation energy from acridine orange-10-dodecyl bromide to methylene blue in micelles. *Photochem. Photobiol.* **1979**, 29, 165–167.
- Hedley, D.; Chow, S. Flow cytometric measurement of lipid peroxidation in vital cells using parinaric acid. *Cytometry* **1992**, 13, 686–692.
- Fitos, I.; Visy, J.; Zsila, F.; Bikadi, Z.; Mady, G.; Simonyi, M. Specific ligand binding on genetic variants of human α 1-acid glycoprotein studied