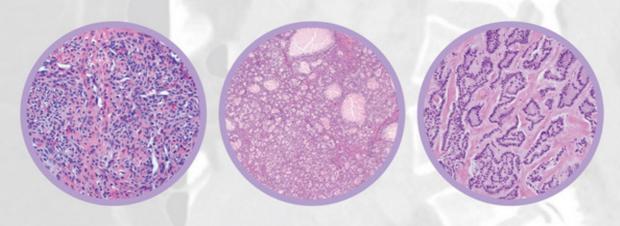
UNCOMMON GYNECOLOGIC CANCERS



Edited by Marcela G. del Carmen, Robert H. Young John O. Schorge, Michael J. Birrer

Uncommon Gynecologic Cancers

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EDITED BY

Marcela G. del Carmen, MD, MPH

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Robert H. Young, MD, FRCPATH

James Homer Wright Pathology Laboratories Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

John O. Schorge, мь

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Michael J. Birrer, MD, PHD

Department of Medicine Massachusetts General Hospital Boston, MA, USA

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Contents

List of Contributors, vi

Preface, viii

Part 1 General Principles, 1

- 1 Molecular Targets in Gynecologic Cancers, 3 Whitfield Growdon, Rosemary Foster, and Bo Rueda
- 2 Imaging of Rare Gynecologic Tumors, 18 Hima B. Prabhakar and Dinushi S. Perera
- 3 Pathology Overview of Rare Gynecologic Tumors, 35 *Robert H. Young*

Part 2 Rare Ovarian Cancers, 51

- 4 Ovarian Clear Cell Carcinoma, 53 Rachel M. Clark and John O. Schorge
- 5 Mucinous Epithelial Ovarian Cancer, 64 Marcela G. del Carmen
- 6 Pseudomyxoma Peritonei, 70 Leslie S. Bradford and Annekathryn Goodman
- 7 Squamous Cell Carcinoma of the Ovary, 81 *Ryan J. Spencer and Laurel W. Rice*
- 8 Endometriosis-Associated Ovarian Cancer, 94
 Leslie A. Garrett and Marcela G. del Carmen
- 9 Carcinosarcoma of the Ovary, 109 J. Alejandro Rauh-Hain and Marcela G. del Carmen
- 10 Malignant Germ Cell Tumors of the Ovary, 120 Marcela G. del Carmen
- 11 Malignant Sex Cord-Stromal Tumors of the Ovary, 132 Marcela G. del Carmen and Richard T. Penson
- 12 Small Cell and Neuroendocrine Cancers of the Ovary, 139 Ariel A. Kaphan and Cesar M. Castro
- 13 Carcinoid Tumors of the Ovary, 148

 Michael J. Worley, Jr. and Michael G. Muto

Part 3 Rare Uterine Cancers, 157

- 14 Uterine Carcinosarcoma, 159

 Premal H. Thaker and Don S. Dizon
- 15 Uterine Leiomyosarcoma, 167 Marcela G. del Carmen
- 16 Mucinous Tumors of the Uterine Corpus, 178 Rachel M. Clark and Marcela G. del Carmen
- 17 Uterine Clear Cell Carcinoma, 186 J. Alejandro Rauh-Hain, Alexander B. Olawaiye, and David M. Boruta
- 18 Updates on the Management of Uterine Serous Carcinoma, 195Amanda Ramos and Amanda Nickles Fader
- 19 Endometrial Stromal Sarcomas, 205 Kathleen M. Schmeler, J. Alejandro Rauh-Hain, and Marcela G. del Carmen

Part 4 Rare Cancers of the Cervix, Vulva and Vagina, 217

- 20 Small Cell Neuroendocrine Carcinoma of the Cervix, 219 Olivia W. Foley, Leslie S. Bradford, and Marcela G. del Carmen
- 21 Primary Malignant Melanoma of the Vagina and Vulva, 228 Alexander B. Olawaiye
- 22 Sarcomas of the Vulva and Vagina, 236 Rachel M. Clark, J. Alejandro Rauh-Hain, and Marcela G. del Carmen
- 23 Gestational Trophoblastic Disease, 245 Whitfield Growdon

Index, 260

List of Contributors

David M. Boruta, MD

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology

Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Leslie S. Bradford, MD

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology

Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Cesar M. Castro, MD

Massachusetts General Hospital Cancer Center and Department of Medicine Harvard Medical School Boston, MA, USA

Rachel M. Clark, MD

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Marcela G. del Carmen, мр, мрн

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Don S. Dizon, MD, FACP

Oncology of Sexual Health Gillette Center for Gynecologic Oncology Massachusetts General Hospital Cancer Center Boston, MA, USA

Amanda Nickles Fader, мр

The Kelly Gynecologic Oncology Service Johns Hopkins Hospital Baltimore, MD, USA

Olivia W. Foley, BA

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Rosemary Foster, PhD

Vincent Center for Reproductive Biology Vincent Department of Obstetrics and Gynecology Massachusetts General Hospital Boston, MA, USA

Leslie A. Garrett, MD

Division of Gynecologic Oncology Department of Obstetrics and Gynecology Beth Israel Deaconess Hospital Boston, MA, USA

Annekathryn Goodman, мо

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Whitfield Growdon, мр

Vincent Center for Reproductive Biology Vincent Department of Obstetrics and Gynecology Massachusetts General Hospital Boston, MA, USA

Ariel A. Kaphan

Massachusetts General Hospital Cancer Center and Department of Medicine, Harvard Medical School, Boston, MA. USA

Michael G. Muto, MD

Division of Gynecologic Oncology Brigham and Women's Hospital Boston, MA, USA

Alexander B. Olawaiye, MD

Division of Gynecologic Oncology Magee-Women's Hospital Pittsburgh, PA, USA List of Contributors vii

Richard T. Penson, MD

Division of Medical Oncology Massachusetts General Hospital Harvard Medical School Boston, MA, USA

Dinushi S. Perera, MD

Pennsylvania Hospital University of Pennsylvania Health System Philadelphia, PA, USA

Hima B. Prabhakar, мо

Perelman School of Medicine of the University of Pennsylvania Division of Body Imaging, Pennsylvania Hospital, Philadelphia, PA, USA

Amanda Ramos, MD

The Kelly Gynecologic Oncology Service Johns Hopkins Hospital Baltimore, MD, USA

J. Alejandro Rauh-Hain, мр

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Laurel W. Rice, MD

Department of Obstetrics and Gynecology University of Wisconsin Hospital and Clinics Madison, WI, USA

Bo Rueda, PhD

Vincent Center for Reproductive Biology Vincent Department of Obstetrics and Gynecology Massachusetts General Hospital Boston, MA, USA

Kathleen M. Schmeler, мо

Department of Gynecologic Oncology The University of Texas MD Anderson Cancer Center Houston, TX, USA

John O. Schorge, MD

Division of Gynecologic Oncology, Vincent Obstetrics and Gynecology Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Ryan J. Spencer, MD

Department of Obstetrics and Gynecology University of Wisconsin Hospital and Clinics Madison, WI, USA

Premal H. Thaker, мо

Department of Obstetrics and Gynecology Washington University School of Medicine St. Louis, MO, USA

Michael J. Worley, Jr., MD

Division of Gynecologic Oncology Brigham and Women's Hospital Boston, MA, USA

Robert H. Young, MD, FRCPATH

James Homer Wright Pathology Laboratories Massachusetts General Hospital, Harvard Medical School Boston, MA, USA

Preface

Tumors of rare histology arising in the gynecologic tract account for a disproportionate number of deaths. Some in the field have advocated that patients with rare gynecologic cancers should be managed by select experts in the field who may have more experience in diagnosing and treating these tumors. This position has resulted in a small number of experts comfortable with the management of more rare and aggressive gynecologic malignancies.

The very rarity of these histologies has made it difficult to collect information for best management strategies derived from prospective clinical trials. Often the reported experience has been limited to retrospective reports from institutions of excellence with more experience.

The primary rationale for this book is to provide a central point of access that will disseminate the most novel diagnostic and treatment strategies for rare gynecologic cancers and make the information accessible to all clinicians caring for these patients. The book aims to place equal emphasis on the clinical and pathological challenges that arise in managing and diagnosing patients with rare gynecologic tumors. To this end, I was privileged to recruit Dr. Robin Young's participation as coeditor. Each chapter contains a section describing the pathologic hallmarks characteristic of each of these rare tumors.

In the last 5 years, advances in the field and new understanding of the molecular biology driving these cancers have resulted in a shift in paradigm for their treatment. Novel management options are centered on the concept of targeted therapies for some of these tumors in place of treatment directed primarily by anatomic site of origin. Even in the modern area of electronic communication,

immediate and central access to these newer principles and strategies is difficult. Textbooks contain limited information helpful to clinicians faced with a patient presenting with one of these cancers. The goal of this book is to summarize the available literature as it pertains to the biology, molecular science advances, pathologic diagnosis, imaging options, and treatment strategies for the management of more rare and aggressive gynecologic cancers.

We intend to organize the book so that each tumor is addressed systematically, proving the reader with the same information and clinical tools across each tumor type, so as to better understand the disease process, its diagnosis and appropriate work up, as well as available treatment options. Invited authors represent leaders in the field with recognized expertise in the treatment of rare gynecologic cancers. Given that these conditions often require expertise from a multidisciplinary team, contributors' expertise will be inclusive of basic science, pathology, diagnostic imaging, radiation oncology, gynecologic oncology, and medical oncology. We hope that the reader will find this book systematically organized and easy to access and that it will serve as a guide in the evaluation, diagnosis, and management of patients with more rare gynecologic malignancies. This book is dedicated to our families, our mentors, and all of our patients.

> Marcela G. del Carmen, MD, MPH Division of Gynecologic Oncology Massachusetts General Hospital Harvard Medical School, 2014

1

PART 1

General Principles

1

CHAPTER 1

Molecular Targets in Gynecologic Cancers

Whitfield Growdon, Rosemary Foster, and Bo Rueda

Vincent Center for Reproductive Biology, Vincent Department of Obstetrics and Gynecology, Massachusetts General Hospital, Boston, MA, USA

Introduction

The past two decades have brought an exponential increase in our understanding of the molecular drivers of cancer. These insights have led to the concept of personalized cancer care where an individual tumor can be interrogated for specific molecular alterations that may render the cancer susceptible to novel therapeutics that target that particular alteration. The advent of HER2 (ERBB2) targeted therapies for HER2 overexpressing breast cancer and EGFR inhibitors for EGFR (ERBB1) gene-mutated lung cancers are notable successes that support the concept that targeting specific molecular profiles can lead to clinical benefit.

Regarding gynecologic cancers, investigators now understand that the underlying drivers of any individual tumor may exhibit marked diversity even if both tumors have identical histology. Identifying key molecular pathways that drive subsets of tumors within ovarian, endometrial, and cervical cancer is crucial to the development of clinical trials utilizing the next generation of targeted therapeutics. This chapter seeks to explore several molecular pathways and proteins that have been shown to contribute to the pathology of significant subsets of ovarian, endometrial, cervical, and vulvar cancers. While the promise of personalized cancer medicine has yet to be fulfilled in gynecologic cancers, therapies targeting the PI3K, MAPK signaling pathways, as well as HER2 and VEGF receptors and PARP protein have been shown to have the potential to improve the therapeutic options for patients.

Phosphoinositol 3-kinase (PI3K) pathway

Oncogenic alterations in the phosphoinositol 3-kinase (PI3K) pathway (Figure 1.1) are frequent in endometrial and ovarian carcinomas [1–3]. PI3K is the upstream activator of Akt, and ultimately mTOR and it contributes to regulation of cell growth, angiogenesis, migration, and survival [2,4]. While three classes of PI3K enzymes have been described, class IA PI3Ks have been most associated with promoting carcinogenesis [5]. PI3K enzymes are activated by receptor tyrosine kinases and G-protein-coupled receptors and transfer phosphate groups to the inositol ring of phosphatidylinositol 4,5 bi-phosphate (PIP2) to produce the signaling molecule phosphatidylinositol 3,4,5 tri-phosphate (PIP3) [1]. This process is negatively regulated by the phosphatase and tensin homologue (PTEN) [5]. Direct downstream mediators AKT and mTOR become activated via phosphorylation leading to transcription events that promote growth, invasion, metastases, and cell survival.

There are many underlying mechanisms for PI3K pathway activation in cancer, including receptor tyrosine kinase activation or amplification, mutation, deletion, silencing of negative regulators of the PI3K pathway, and activation or amplification of downstream kinase mediators [4]. Correlative investigations have demonstrated a significant prevalence of gain of function mutations in the *PIK3CA* gene in breast, colon, pancreatic, brain, ovary, and, recently, high-risk endometrial cancers [2,6–12].

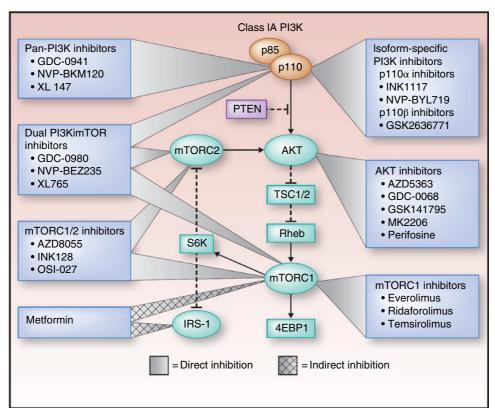


Figure 1.1 PI3K signaling cascade. Schematic showing the PI3K/AKT/mTOR and MAPK pathways and the current agents in development for targeting this cascade.

Recent reports have suggested that gene amplification affects approximately 20–40% of ovarian, endometrial, and cervical carcinomas across all subtypes, while gain of function mutations occur more commonly in endometrioid endometrial cancer and in clear-cell and endometrioid ovarian tumors at approximately a 20% rate [13–18]. Additionally, PI3K activation via these mechanisms was associated with chemoresistance and worsened survival, suggesting targeted inhibition could potentiate conventional platinum-based chemotherapy [19–24].

Given the high prevalence of PI3K pathway activation in gynecologic cancer, targeted strategies inhibiting this cascade could hold tremendous potential to benefit patients with ovarian, endometrial, and cervical cancer [11,25]. Multiple phase I and II clinical trials in endometrial and ovarian cancer have tested agents that target the PI3K pathway [26–28]. Reports from phase II trials of rapalogs inhibiting the mammalian target of rapamycin

(mTOR), a downstream mediator of the PI3K pathway, have revealed both objective responses as well as clinically significant disease stabilization [27,29,30]. In addition to the rapalogs, several other classes of PI3K pathway inhibitors including direct PI3K inhibitors, PI3K/mTOR dual inhibitors, and AKT inhibitors are in development for treating ovarian and endometrial cancer [5,28,31]. Early reports from clinical trials suggest that responses to single-agent blockade have an approximately 30% prevalence, occur with or without gain of function mutations in PIK3CA, and manifest limited response durability resulting in treatment resistance [26, 32-35]. This observation has resulted in the hypothesis that targeted blockade of one overactive protein in a fundamental pathway, such as PI3K, AKT, or mTOR, may not result in significant clinical response. Understanding resistance mechanisms will be critical in the clinical implementation of targeted therapies.

The identification of a biomarker associated with response will be crucial to the success of targeted therapy in general. For the PI3K pathway, PIK3CA gene amplification and gain-of-function mutation in both the catalytic subunit (PIK3CA) and the regulatory subunit (PIK3R1) have been described in ovarian, endometrial, and cervical cancer. Some preclinical and clinical data have suggested that those tumors harboring a mutation have increased sensitivity to PI3K pathway inhibition [24,32,36]. Of the gynecologic malignancies, endometrial cancer has the highest prevalence of these molecular alterations. These data have clear implications for selecting candidates for clinical trials so that accrual can enrich for those patients most likely to respond; however, responses to PI3K pathway have been observed in patients that harbor no mutation or amplification, suggesting that additional criteria need to be utilized to identify those women most likely to respond [37-40]. These observations have been confirmed in phase I trials of PI3K inhibitors, although the most robust responses were witnessed in those patients that carry a tumor with specific gain-of-function mutations [41-55]. Selection of endometrial cancer patients by loss of PTEN or gain-offunction mutation in PIK3CA for clinical trials of agents targeting PI3K or AKT is ongoing and it has yet to be determined whether or not these signatures confer sensitivity to directed therapy.

Mitogen-activated protein kinase (MAPK) pathway

The mitogen-activated protein kinase (MAPK) signaling pathway is another growth-signaling cascade associated with multiple cancers that is an attractive target for the development of targeted therapeutics [56–58]. The receptor tyrosine kinases (RTK) family is one of the more recognized kinase families. The MAPK kinase kinase (MAPKK) phosphorylates and activates MAPK kinase (MAPKK) that in turn can phosphorylate and activate MAPK by phosphorylation on the Thr and Tyr resides [59]. Members of the GTPase families, Ras and Rho, relay signals from the receptor complex to the MAPKKK. There are four major MAPK signaling pathways in mammals. These include extracellular signal-related kinase (ERK), ERK5, p38-MAPK 1 and 2, and c-jun N-terminal kinase (JNK)1, 2, and 3. Typically, the ERK pathways respond to growth

factor stimuli and p38 MAPK and c-jun are activated in response to stress stimuli such as UV irradiation and inflammatory cytokine [60,61]. There are, however, examples of growth factors activating the p38-MAPK and c-JNK via cross talk.

Ras proteins are integral intermediate modulators connecting the membrane receptors on the cell surface with their downstream effector MAPK signaling pathways. It has been reported that between 11.6% and 83% of endometrioid endometrial cancers harbor k-ras mutations [62–67]. Investigations utilizing endometrial cancer cell line models have suggested that MAPK/ERK1-2 is involved in promoting endometrial cancer cell proliferation in a number of *in vitro* studies utilizing *KRAS* mutant cell lines and MEK inhibitor [68–73].

A recent study further highlights the complexity of the interaction among the signaling pathways. Metformin, an oral biguanide commonly used for the treatment of type II diabetes, is thought to inhibit cell proliferation locally via activation of the AMPK signaling pathway, counteracting the growth-promoting effects of the PI3K/AKT/mTOR pathway. Recently, it was shown to be effective in down-regulating of ERK and AKT signaling and increasing cell death in endometrial cancer cells that constitutively expressed k-ras in endometrial cancer. Metformin resulted in concentration-dependent activation of AMPK in endometrial cancer cell lines [74,75]. The MAPK inhibitor, Selumetinib* (AZD-6244) is being tested in the recurrent endometrial cancer (NCT01011933) [75].

MAPK appears to play a significant role in the pathophysiology of low-grade serous cancers (LGSC) of the ovary, as well as serous borderline tumors. While approximately 85% of epithelial ovarian cancers are serous, only about 10% of these are LGSC [76–78]. LGSC rarely have p53 mutations but can have mutations of *KRAS* or *BRAF*. In contrast, high-grade serous cancers typically have evidence of p53 mutations and rarely *KRAS* and *BRAF* mutations.

An estimated 80% of ovarian LGSC have an active MAP kinase pathway [79]. Likewise, 78% of its putative precursor lesion, borderline tumors, also historically termed "low malignant potential (LMP) tumors," has been shown to have an active MAP kinase pathway [79,80]. While the LGSC and high-grade serous carcinomas have different clinical outcomes and molecular profiles, they are both treated with surgery followed by platinum and taxane-based therapy. The identification of

specific pathways may result in targeted treatments leading to better outcomes.

In a phase II Gynecologic Oncology Group (GOG) trial of MEK inhibitor Selumetinib* (AZD-6244), a 15% response rate was reported. There was a stable disease rate of 65% and a medium progression-free survival (PFS) rate of 11 months in patients with recurrent LGSC [81].

Human epidermal growth factor receptor 2 (HER2)

Human epidermal growth factor receptor 2 (HER2), also called HER2/neu or c-erbB2 is a growth factor receptor tyrosine kinase implicated in many cancers. Amplification of the HER2 (ERBB2) gene and overexpression of the HER2 protein have been described in breast, colon, gastric, esophageal, ovarian, and endometrial cancers [82-88]. The HER2gene encodes a 185-kDa transmembrane tyrosine kinase receptor and is located on chromosome 17q21. HER2 is a well-characterized member of the human epidermal growth factor receptor superfamily that consists of three other tyrosine kinase receptors (HER1/EGFR, HER3, and HER4) that when activated by ligand, can dimerize and induce signal transduction through the MAPK and PI3K/AKT/mTOR signaling pathways [89-96]. This downstream activation leads to induction of genes that promote oncogenic transformation via cell survival, proliferation, angiogenesis, and metastasis (Figure 1.1).

High-grade endometrial cancer, including grade 3 endometrioid, uterine serous carcinoma (USC) and carcinosarcoma, has a 10–30% rate of *HER2* gene amplification, with up to 70% of tumors exhibiting HER2 protein overexpression, which has been associated with decreased overall survival [97–103]. Despite promising preclinical data, the two phase II trials of anti-HER2 therapy in recurrent endometrial cancer showed poor responses [99, 100]. These trials suggest that single-agent therapies directed against HER2 may have limited activity, possibly due to innate or drug-induced resistance [104].

In a trial of 800 ovarian carcinomas screened for membrane HER2 protein expression, 12% of tumors showed 2 or 3+ protein expression. Given the low prevalence of this event in the majority of ovarian cancers, its therapeutic value is likely limited [104–107].

Vascular endothelial growth factor (VEGF)

Angiogenesis is a key component of all tumor cell biology and has recently become a promising therapeutic target for women undergoing treatment for gynecologic cancers [108–123].

Most anti-angiogenic therapeutics target the VEGF signaling pathway. The VEGF family includes six related proteins. The most important member is VEGF-A, which was discovered first and was called simply "VEGF" before other variants (VEGF-B, C, and D) with more specialized functions involving embryonic and site-specific angiogenesis were described [124–128]. VEGF-A will be referenced as simply "VEGF" in this chapter.

Within the gynecologic malignancies, angiogenic signaling has been most studied in ovarian cancer [129]. Data suggest that angiogenesis plays a key role in metastatic spread of ovarian carcinoma, and increased angiogenic signaling has been shown to be a poor prognostic factor in ovarian cancer [122,130–137]. Analyses of ovarian cancer cell lines and human tumors have demonstrated higher levels of pro-angiogenic factors, such as hypoxia-inducible factor-1 (HIF-1), VEGF, and PDGF, as well as lower levels of anti-angiogenic factors, such as end-ostatins [136,137].

Retrospective studies in high- and low-grade ovarian cancer suggested benefit of utilizing bevacizumab, a humanized monoclonal antibody to circulating VEGF, in the recurrent, chemotherapy refractory setting [138-143]. Two subsequent phase II trials were published showing response rates as high as 15-21%, with a clinical benefit rate (stable disease rate + response rate) of greater than 60% [140,141]. Phase III trial data have documented the activity of bevacizumab in ovarian cancer [144-163]. In 2012, the OCEANS trial reported that combining bevacizumab with carboplatinum and gemcitabine significantly prolonged the PFS by 4 months in women with recurrent platinum-sensitive ovarian cancer compared to the combination without bevacizumab [144,145]. Notably, the experimental arm of this trial administered the bevacizumab to patients until progression of disease, raising the possibility that extended anti-angiogenic therapy also contributed to the survival benefit observed. The recent report of the AURELIA trial (NCT00976911) that randomized women with platinum-resistant ovarian cancer to liposomal doxorubicin, topotecan or weekly paclitaxel with or without bevacizumab suggested that adding anti-angiogenic therapy improved PFS when compared to use of single-agent chemotherapy alone. The most robust synergy was noted to be paclitaxel and bevacizumab with an almost 11-month PFS prolongation [146,147]. Use of bevacizumab in first-line and maintenance ovarian cancer has been evaluated via two phase III trials, GOG 218 and ICON 7, resulting in an improved PFS but no impact on overall survival [161–163].

A growing number of other anti-angiogenic agents have entered clinical trial that target VEGF or its receptor. A phase II trial using VEGF-Trap (aflibercept) with docetaxel demonstrated a 54% response rate in recurrent ovarian cancer [148]. In addition, numerous novel receptor tyrosine kinase (RTK) inhibitors directed against VEGF receptor have been tested. Agents such as sorafinib, sunitinib, and pazopanib interact with multiple additional RTKs in addition to the VEGF receptor including the PDGF receptor, epidermal growth factor receptor (EGFR), and c-Kit. Limited clinical trials in women with recurrent ovarian cancer have been performed using these multiple RTK inhibitors and these investigations have not revealed responses as robust as those observed with bevacizumab [149-153]. None of the trials testing angiogenic agents in gynecologic cancers have tested for heightened activation of the VEGF pathway and this type of approach may be required to enrich for relevant clinical responses for some of the multikinase inhibitors [151,154–163].

The role of angiogenesis in endometrial cancer is less well-understood. Immunohistochemical staining for mean vessel density (MVD) counts has been employed in the preclinical setting to investigate angiogenesis in endometrial hyperplasia and carcinoma. Abulafia et al. compared MVD counts in endometrial hyperplasia compared to Stage I endometrial carcinoma finding that as the histology progressed from simple hyperplasia to invasive carcinoma, so too did the MVD counts [164]. The authors concluded that higher tumor grade and depth of invasion were directly correlated with increasing angiogenic activation [164]. Kaku et al. confirmed these findings with their investigation of 85 specimens from patients with Stage I and II endometrial carcinoma, where MVD was strongly correlated with tumor grade, depth of myometrial invasion, as well as lymphovascular space invasion [165]. An additional preclinical study supported the finding that higher MVD counts are associated with worse PFS and overall survival (OS), confirming that angiogenesis appears to be a clinically relevant signature in endometrial cancer [166].

One phase II trial tested single-agent sorafenib, a multitarget tyrosine kinase inhibitor (TKI), finding a modest 5% response rate [167]. Another phase II trial investigated sunitinib in this same population and reported a 15% response rate with major toxicities limited to fatigue and hypertension [168]. Bevacizumab has been studied in a phase II trial (GOG 229E) demonstrating a 13.5% response rate which compared favorably with other single agents tested in the same setting. Tumor and serum VEGF levels were quantified by immunohistochemistry and ELISA and correlated with response and survival. High tumor VEGF staining correlated with improved survival, while elevated circulating serum VEGF levels correlated with decreased survival and treatment failure [169]. This study suggests that VEGF levels could serve as biomarkers to predict which patients are most likely to respond to VEGF directed therapies.

The development of abnormal vascularity has been described in cervical cancer [170]. Investigations examining the histology of early and late cervical cancers have demonstrated that almost 3% of patients with carcinoma in situ (CIS) produced abnormal vessels noted at the time of colposcopy compared to 50% of patients with microinvasive disease and 100% of patients with frankly invasive cancer [171]. Studies examining MVD and VEGF expression have inconsistently found that MVD counts in conjunction with depth of invasion, regional lymph node involvement, and lymphovascular invasion can offer independent prognostic information for women with cervical cancer [172-177]. Higher levels of VEGF protein expression have been correlated with tumor size, lymphovascular space invasion, and lymph node metastases, as well as a shorter disease-free interval [178-180].

Bevacizumab was tested in a phase II clinical trial for women with recurrent or persistent cervical cancer. In this trial (GOG227C), Monk and colleagues observed a 10.9% response rate, with a 24% stable disease rate at 6 months in a heavily pretreated and radiated population. The median PFS was 3.4 months, with responders manifesting a 6.2-month duration of response [181,182]. While additional trials have tested agents such as pazopanib and sunitinib (both targeting VEGFR, PGFR, c-Kit), response rates were limited with modest improvements in survival measured in weeks [181,183]. In the phase III trial,

GOG240 (NCT00803062) of advanced-stage, recurrent/ persistent cervical cancer, bevacizumab was evaluated in a four-by-four design with cisplatinum/paclitaxel, and topotecan/paclitaxel. Preliminary data confirmed a 3.7-month OS advantage with use of bevacizumab in combination with either cytotoxic doublet.

Perhaps one of the most promising targets in gynecologic cancers, VEGF has been the focus of many basic, translational, and clinical trial investigations, suggesting it to be a key promoter in the development and progression of epithelial ovarian, endometrial, and cervical cancer. Markers of altered angiogenesis, such as MVD or VEGF expression harbor prognostic value in these gynecologic malignancies, and clinical trials targeting mediators of neovascularization have been shown to have clear benefits in significant subsets of patients. Future challenges include the development of biomarkers that associate with response, managing the costs of expensive therapies, and vigilance in assessing whether or not anti-angiogenic therapies have unintended consequences, such as a rebound effect, which could render tumors more resistant to conventional chemotherapies.

Poly (adenosine) diphosphate [ADP]-ribose polymerase (PARP)

Perhaps one of the most notable successes of translational therapeutics has been the advent of poly (adenosine) diphosphate [ADP]-ribose polymerase (PARP) inhibition in women with a germ-line mutation in the *BRCA1* and *BRCA2* genes. PARP inhibition in patients with *BRCA* mutation has produced encouraging responses suggesting that selective targeting of a molecular finger-print can produce responses in the select population manifesting the signature.

A subset of 5–10% of patients with epithelial ovarian cancer has germ-line inactivating mutations in the *BRCA1* and *BRCA2* genes [184–187]. Approximately, 90% of hereditary ovarian cancer is *BRCA*-associated [188]. Both *BRCA1* and *BRCA2* are tumor suppressor proteins involved with homologous recombination (HR) required for repair of double-stranded DNA breaks [189,190]. A germ-line *BRCA* mutation constitutes the first "hit" that leads to a marked predisposition for patients to develop breast and ovarian cancer, in addition

to other cancers such as prostate and pancreatic [191]. These cancers are thought to arise as a result of inactivation of the other functioning BRCA allele. When both alleles are mutated, somatic cells utilize less meticulous repair DNA mechanisms intended for single-strand breaks, specifically base excision repair (BER), to compensate for accumulated DNA damage leading to oncogenic transformation [192]. The BER process is mediated by the PARP enzyme [193,194]. In cells lacking BRCAinduced HR, the inhibition of PARP would lead to apoptosis as a result of failed DNA repair. Researchers have coined the term "synthetic lethality" [195,196] to describe this BRCA-mutation-dependent cytotoxic effect because PARP inhibition in the setting of wild-type BRCA leads to minimal cellular toxicity. These characteristics make PARP inhibition an attractive therapeutic strategy.

Numerous inhibitors of PARP have been developed and a recent phase I trial evaluated olaparib (AZD 2281), an oral PARP, in 60 diverse cancer patients, finding that of the 15 women in this heavily pretreated cohort with ovarian cancer and a *BRCA* mutation, 8 patients responded, and 1 patient had stable disease [197,198]. Another phase I trial showed similar efficacy in platinum-resistant and sensitive ovarian cancer [199]. In a follow-up phase II study of women with ovarian cancer, Audeh and colleagues tested olaparib in recurrent, *BRCA*-mutated ovarian cancer and confirmed an overall response rate of 25% in 57 patients that had received a median 3–4 prior lines of therapy. At the higher dose 400 mg BID, the response rate was 33% with minimal toxicity [200].

Emerging data suggest that PARP inhibition not only offers clinical benefit to patients with *BRCA* gene mutation, but also appears to have activity in those who lack an identified mutation. In a trial of olaparib therapy in 91 women with recurrent ovarian cancer, a response rate of 41% was observed if a *BRCA* mutation, but interestingly a 24% response rate was manifest in the patients with sporadic ovarian cancer [201]. Researchers are optimistic that like in triple-negative breast cancer [202–204], responses to PARP inhibition will be observed in patients with ovarian cancer that lack *BRCA* mutation, likely secondary to epigenetic *BRCA* inactivation or "BRCAness" that has been observed in up to 31% of sporadic ovarian tumors [205–208].

PARP inhibition has also been evaluated in the maintenance setting. In randomized trial of patients with recurrent ovarian cancer to either olaparib or placebo, a significant PFS benefit (8.4 vs. 4.8 months, respectively, HR 0.35; 95% CI 0.25–0.49, P<0.001) was noted. No significant difference in overall survival has been reported.

Utilizing PARP inhibition in concert with cytotoxic chemotherapy has also been a rationale approach supported by preclinical data suggesting a synergy [209,210]. Simultaneous use of platinum agents with PARP inhibitors potentiates toxicity. Despite the potential for this toxicity, trials examining PARP inhibitors such as olaparib, veliparib (ABT-888), and iniparib (BSI-201) in combination with carboplatin-containing regimens for women with platinum-sensitive and resistant ovarian cancer are expected to reveal strong responses that justify the development of upfront trials examining the use of combined therapy (NCT01033123, NCT01033292, NCT01650376, NCT01081951, NCT01459380). Synergistic effects are anticipated, particularly in those patients with *BRCA* gene mutations, though this has yet to be reported.

PARP inhibition stands as a proof of concept that molecular signatures can perform as potent biomarkers to predict response to therapies that target that specific molecular alteration. While the most robust responses in ovarian cancer have been reported in those women with a germ-line *BRCA* gene mutation, emerging genomic and clinical data suggest that a significant proportion of women with ovarian cancer who lack mutation may benefit from PARP inhibition. It is currently unclear if PARP inhibition will offer the greatest benefit in the upfront, maintenance, or recurrent setting, but the next generation of trials is expected to provide more guidance on how best to incorporate PARP inhibition into the clinical care of women with ovarian cancer.

References

- 1 Engelman, J.A., Luo, J. and Cantley, L.C. (2006) The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nature Reviews Genetics*, 7, 606–619.
- 2 Samuels, Y., Wang, Z., Bardelli, A. *et al.* (2004) High frequency of mutations of the PIK3CA gene in human cancers. *Science* (*New York, NY*), **304**, 554.
- 3 Courtney, K.D., Corcoran, R.B. and Engelman, J.A. (2010) The PI3K pathway as drug target in human cancer. *Journal of Clinical Oncology*, 28, 1075–1083.

- 4 Yuan, T.L. and Cantley, L.C. (2008) PI3K pathway alterations in cancer: variations on a theme. *Oncogene*, **27**, 5497–5510.
- 5 Courtney, K.D., Corcoran, R.B. and Engelman, J.A. (2010) The PI3K pathway as drug target in human cancer. *Journal of Clinical Oncology*, 28, 1075–1083.
- 6 Samuels, Y. and Velculescu, V.E. (2004) Oncogenic mutations of PIK3CA in human cancers. Cell Cycle, 3, 1221–1224.
- 7 Abubaker, J., Bavi, P., Al-Haqawi, W. et al. (2009) PIK3CA alterations in Middle Eastern ovarian cancers. *Molecular Cancer*, 8, 51.
- 8 Souglakos, J., Philips, J., Wang, R. et al. (2009) Prognostic and predictive value of common mutations for treatment response and survival in patients with metastatic colorectal cancer. British Journal of Cancer, 101, 465–472.
- 9 Kalinsky, K., Jacks, L.M., Heguy, A. et al. (2009) PIK3CA mutation associates with improved outcome in breast cancer. Clinical Cancer Research, 15, 5049–5059.
- 10 Brugge, J., Hung, M.C. and Mills, G.B. (2007) A new mutational AKTivation in the PI3K pathway. *Cancer Cell*, 12, 104–107.
- 11 Salvesen, H.B., Carter, S.L., Mannelqvist, M. et al. (2009) Integrated genomic profiling of endometrial carcinoma associates aggressive tumors with indicators of PI3 kinase activation. Proceedings of National Academy of Sciences U S A, 106, 4834–4839.
- 12 Dobbin, Z.C. and Landen, C.N. (2013) The importance of the PI3K/AKT/MTOR pathway in the progression of ovarian cancer. *International Journal of Molecular Sciences*, 14, 8213–8227.
- 13 Campbell, I.G., Russell, S.E., Choong, D.Y. et al. (2004) Mutation of the PIK3CA gene in ovarian and breast cancer. Cancer Research, 64, 7678–7681.
- 14 Kolasa, I.K., Rembiszewska, A., Felisiak, A. et al. (2009) PIK3CA amplification associates with resistance to chemotherapy in ovarian cancer patients. Cancer Biology & Therapy, 8, 21–26.
- 15 Kuo, K.T., Mao, T.L., Jones, S. et al. (2009) Frequent activating mutations of PIK3CA in ovarian clear cell carcinoma. The American Journal of Pathology, 174, 1597–1601.
- 16 Dedes, K.J., Wetterskog, D., Ashworth, A. et al. (2011) Emerging therapeutic targets in endometrial cancer. Nature Reviews Clinical Oncology, 8, 261–271.
- 17 De Marco, C., Rinaldo, N., Bruni, P. et al. (2013) Multiple genetic alterations within the PI3K pathway are responsible for AKT activation in patients with ovarian carcinoma. PLoS One, 8, e55362.
- 18 Bertelsen, B.I., Steine, S.J., Sandvei, R. et al. (2006) Molecular analysis of the PI3K-AKT pathway in uterine cervical neoplasia: frequent PIK3CA amplification and AKT phosphorylation. *International Journal of Cancer*, 118, 1877–1883.

- 19 Dent, P., Grant, S., Fisher, P.B. and Curiel, D.T. (2009) PI3K: a rational target for ovarian cancer therapy? *Cancer Biology & Therapy*, 8, 27–30.
- 20 Lee, S., Choi, E.J., Jin, C. and Kim, D.H. (2005) Activation of PI3K/Akt pathway by PTEN reduction and PIK3CA mRNA amplification contributes to cisplatin resistance in an ovarian cancer cell line. *Gynecologic Oncology*, 97, 26–34.
- 21 Schwarz, J.K., Payton, J.E., Rashmi, R. et al. (2012) Pathway-specific analysis of gene expression data identifies the PI3K/Akt pathway as a novel therapeutic target in cervical cancer. Clinical Cancer Research, 18, 1464–1471.
- 22 Carracedo, A. and Pandolfi, P.P. (2008) The PTEN-PI3K pathway: of feedbacks and cross-talks. Oncogene, 27, 5527–5541.
- 23 Sheppard, K.E., Cullinane, C., Hannan, K.M. et al. (2013) Synergistic inhibition of ovarian cancer cell growth by combining selective PI3K/mTOR and RAS/ERK pathway inhibitors. European Journal of Cancer, 49, 3936–3944.
- 24 Sos, M.L., Fischer, S., Ullrich, R. et al. (2009) Identifying genotype-dependent efficacy of single and combined PI3K- and MAPK-pathway inhibition in cancer. Proceedings of National Academy of Sciences U S A, 106, 18351–18356.
- 25 Glaysher, S., Bolton, L.M., Johnson, P. et al. (2013) Targeting EGFR and PI3K pathways in ovarian cancer. British Journal of Cancer, 109, 1786–1794.
- 26 Janku, F., Wheler, J.J., Westin, S.N. et al. (2012) PI3K/AKT/ mTOR inhibitors in patients with breast and gynecologic malignancies harboring PIK3CA mutations. *Journal of Clinical Oncology*, 30, 777–782.
- 27 Behbakht, K., Sill, M.W., Darcy, K.M. et al. (2011) Phase II trial of the mTOR inhibitor, temsirolimus and evaluation of circulating tumor cells and tumor biomarkers in persistent and recurrent epithelial ovarian and primary peritoneal malignancies: a Gynecologic Oncology Group study. Gynecologic Oncology, 123, 19–26.
- 28 Polivka, J., Jr and Janku, F. (2013) Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. *Pharmacology & Therapeutics*, **142** (2), 164–175.
- 29 Oza, A.M., Elit, L., Tsao, M.S. et al. (2011) Phase II study of temsirolimus in women with recurrent or metastatic endometrial cancer: a trial of the NCIC Clinical Trials Group. *Journal of Clinical Oncology*, 29, 3278–3285.
- 30 Slomovitz, B.M., Lu, K.H., Johnston, T. et al. (2010) A phase 2 study of the oral mammalian target of rapamycin inhibitor, everolimus, in patients with recurrent endometrial carcinoma. Cancer, 116, 5415–5419.
- 31 Slomovitz, B.M. and Coleman, R.L. (2012) The PI3K/AKT/ mTOR pathway as a therapeutic target in endometrial cancer. Clinical Cancer Research, 18, 5856–5864.
- 32 Janku, F., Wheler, J.J., Naing, A. et al. (2013) PIK3CA mutation H1047R is associated with response to PI3K/AKT/mTOR signaling pathway inhibitors in early phase clinical trials. Cancer Research, 73, 276–284.

- 33 Janku, F., Wheler, J.J., Naing, A. et al. (2012) PIK3CA mutations in advanced cancers: characteristics and outcomes. Oncotarget, 3, 1566–1575.
- 34 Squillace, R.M., Miller, D., Cookson, M. et al. (2011) Antitumor activity of ridaforolimus and potential cellcycle determinants of sensitivity in sarcoma and endometrial cancer models. Molecular Cancer Therapeutics, 10, 1959–1968.
- 35 Shoji, K., Oda, K., Kashiyama, T. et al. (2012) Genotype-dependent efficacy of a dual PI3K/mTOR inhibitor, NVP-BEZ235, and an mTOR inhibitor, RAD001, in endometrial carcinomas. PLoS One, 7, e37431.
- 36 O'Brien, C., Wallin, J.J., Sampath, D. et al. (2011) Predictive biomarkers of sensitivity to the phosphatidylinositol 3' kinase inhibitor GDC-0941 in breast cancer preclinical models. Clinical Cancer Research, 16, 3670–3683.
- 37 Dan, S., Okamura, M., Seki, M. et al. (2010) Correlating phosphatidylinositol 3-kinase inhibitor efficacy with signaling pathway status: in silico and biological evaluations. Cancer Research, 70, 4982–4994.
- 38 O'Brien, C., Wallin, J.J., Sampath, D. et al. (2010) Predictive biomarkers of sensitivity to the phosphatidylinositol 3' kinase inhibitor GDC-0941 in breast cancer preclinical models. Clinical Cancer Research, 16, 3670–3683.
- 39 Wallin, J.J., Edgar, K.A., Guan, J. et al. (2011) GDC-0980 is a novel class I PI3K/mTOR kinase inhibitor with robust activity in cancer models driven by the PI3K pathway. Molecular Cancer Therapeutics, 10, 2426–2436.
- 40 Engelman, J.A., Chen, L., Tan, X. et al. (2008) Effective use of PI3K and MEK inhibitors to treat mutant Kras G12D and PIK3CA H1047R murine lung cancers. *Nature Medicine*, 14, 1351–1356.
- 41 Janku, F., Wheler, J.J., Naing, A. *et al.* (2012) PIK3CA mutations in advanced cancers: characteristics and outcomes. *Oncotarget*, **3**, 1566–1575.
- 42 Janku, F., Wheler, J.J., Naing, A. et al. (2013) PIK3CA mutation H1047R is associated with response to PI3K/AKT/mTOR signaling pathway inhibitors in early-phase clinical trials. Cancer Research, 73, 276–284.
- 43 McCubrey, J.A., Steelman, L.S., Chappell, W.H. et al. (2012) Ras/Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR cascade inhibitors: how mutations can result in therapy resistance and how to overcome resistance. Oncotarget, 3, 1068–1111.
- 44 Janku, F., Lee, J.J., Tsimberidou, A.M. et al. (2011) PIK3CA mutations frequently coexist with RAS and BRAF mutations in patients with advanced cancers. PLoS One, 6, e22769.
- 45 Janku, F., Wheler, J.J., Westin, S.N. et al. (2012) PI3K/AKT/ mTOR inhibitors in patients with breast and gynecologic malignancies harboring PIK3CA mutations. *Journal of Clinical Oncology*, 30, 777–782.
- 46 O'Reilly, K.E., Rojo, F., She, Q.B. et al. (2006) mTOR inhibition induces upstream receptor tyrosine kinase signaling and activates Akt. Cancer Research, 66, 1500–1508.

- 47 Slomovitz, B.M. and Coleman, R.L. (2012) The PI3K/AKT/ mTOR pathway as a therapeutic target in endometrial cancer. Clinical Cancer Research, 18, 5856–5864.
- 48 Shoji, K., Oda, K., Kashiyama, T. et al. (2012) Genotype-dependent efficacy of a dual PI3K/mTOR inhibitor, NVP-BEZ235, and an mTOR inhibitor, RAD001, in endometrial carcinomas. PLoS One, 7, e37431.
- 49 Muellner, M.K., Uras, I.Z., Gapp, B.V. et al. (2011) A chemical-genetic screen reveals a mechanism of resistance to PI3K inhibitors in cancer. *Nature Chemical Biology*, 7, 787–793.
- 50 Ilic, N., Utermark, T., Widlund, H.R. and Roberts, T.M. (2011) PI3K-targeted therapy can be evaded by gene amplification along the MYC-eukaryotic translation initiation factor 4E (eIF4E) axis. Proceedings of National Academy of Sciences U S A, 108, E699–E708.
- 51 Ibrahim, Y.H., Garcia-Garcia, C., Serra, V. et al. (2012) PI3K inhibition impairs BRCA1/2 expression and sensitizes BRCA-proficient triple-negative breast cancer to PARP inhibition. Cancer Discovery, 2, 1036–1047.
- 52 Juvekar, A., Burga, L.N., Hu, H. et al. (2012) Combining a PI3K inhibitor with a PARP inhibitor provides an effective therapy for BRCA1-related breast cancer. Cancer Discovery, 2, 1048–1063.
- 53 Forster, M.D., Dedes, K.J., Sandhu, S. et al. (2011) Treatment with olaparib in a patient with PTEN-deficient endometrioid endometrial cancer. *Nature Reviews Clinical Oncology*, 8, 302–306.
- 54 Shaheen, M., Allen, C., Nickoloff, J.A. and Hromas, R. (2011) Synthetic lethality: exploiting the addiction of cancer to DNA repair. *Blood*, 117, 6074–6082.
- 55 Leung, M., Rosen, D., Fields, S. et al. (2011) Poly(ADPribose) polymerase-1 inhibition: preclinical and clinical development of synthetic lethality. Molecular Medicine, 17, 854–862.
- 56 Shields, J.M., Pruitt, K., McFall, A. et al. (2000) Understanding Ras: 'it ain't over 'til it's over'. Trends in Cell Biology, 10, 147–154.
- 57 Flaherty, K.T., Robert, C., Hersey, P. et al. (2012) Improved survival with MEK inhibition in BRAF-mutated melanoma. The New England Journal of Medicine, 367, 107–114.
- 58 Chapman, P.B., Hauschild, A., Robert, C. et al. (2011) Improved survival with vemurafenib in melanoma with BRAF V600E mutation. The New England Journal of Medicine, 364, 2507–2516.
- 59 Krishna, M. and Narang, H. (2008) The complexity of mitogen-activated protein kinases (MAPKs) made simple. Cellular and Molecular Life Sciences: CMLS, 65, 3525–3544.
- 60 Narang, H. and Krishna, M. (2008) Effect of nitric oxide donor and gamma irradiation on MAPK signaling in murine peritoneal macrophages. *Journal of Cellular Biochemistry*, 103, 576–587.

- 61 McCubrey, J.A., Steelman, L.S., Chappell, W.H. et al. (2012) Mutations and deregulation of Ras/Raf/MEK/ERK and PI3K/PTEN/Akt/mTOR cascades which alter therapy response. Oncotarget, 3, 954–987.
- 62 Dedes, K.J., Wetterskog, D., Ashworth, A. et al. (2011) Emerging therapeutic targets in endometrial cancer. Nature Reviews Clinical Oncology, 8, 261–271.
- 63 Duggan, B.D., Felix, J.C., Muderspach, L.I. et al. (1994) Early mutational activation of the c-Ki-ras oncogene in endometrial carcinoma. Cancer Research, 54, 1604–1607.
- 64 Caduff, R.F., Johnston, C.M. and Frank, T.S. (1995) Mutations of the Ki-ras oncogene in carcinoma of the endometrium. *The American Journal of Pathology*, 146, 182–188.
- 65 Lax, S.F., Kendall, B., Tashiro, H. et al. (2000) The frequency of p53, K-ras mutations, and microsatellite instability differs in uterine endometrioid and serous carcinoma: evidence of distinct molecular genetic pathways. *Cancer*, 88, 814–824.
- 66 Enomoto, T., Fujita, M., Inoue, M. et al. (1993) Alterations of the p53 tumor suppressor gene and its association with activation of the c-K-ras-2 protooncogene in premalignant and malignant lesions of the human uterine endometrium. Cancer Research, 53, 1883–1888.
- 67 Wang, L.E., Ma, H., Hale, K.S. et al. (2012) Roles of genetic variants in the PI3K and RAS/RAF pathways in susceptibility to endometrial cancer and clinical outcomes. *Journal of Cancer Research and Clinical Oncology*, 138, 377–385.
- 68 Ninomiya, Y., Kato, K., Takahashi, A. et al. (2004) K-Ras and H-Ras activation promote distinct consequences on endometrial cell survival. Cancer Research, 64, 2759–2765.
- 69 Suga, S., Kato, K., Ohgami, T. *et al.* (2007) An inhibitory effect on cell proliferation by blockage of the MAPK/estrogen receptor/MDM2 signal pathway in gynecologic cancer. *Gynecologic Oncology*, **105**, 341–350.
- 70 Weigelt, B., Warne, P.H., Lambros, M.B. et al. (2013) PI3K pathway dependencies in endometrioid endometrial cancer cell lines. Clinical Cancer Research, 19, 3533–3544.
- 71 Oda, K., Stokoe, D., Taketani, Y. and McCormick, F. (2005) High frequency of coexistent mutations of PIK3CA and PTEN genes in endometrial carcinoma. *Cancer Research*, 65, 10669–10673.
- 72 Kandoth, C., Schultz, N., Cherniack, A.D. et al. (2013) Integrated genomic characterization of endometrial carcinoma. *Nature*, 497, 67–73.
- 73 Hoeflich, K.P., O'Brien, C., Boyd, Z. et al. (2009) In vivo antitumor activity of MEK and phosphatidylinositol 3-kinase inhibitors in basal-like breast cancer models. Clinical Cancer Research, 15, 4649–4664.
- 74 Iglesias, D.A., Yates, M.S., van der Hoeven, D. et al. (2013) Another surprise from metformin: novel mechanism of action via k-ras influences endometrial cancer response to therapy. Molecular Cancer Therapeutics, 12, 2847–2856.

- 75 Epstein, R.J. (2013) The unpluggable in pursuit of the undruggable: tackling the dark matter of the cancer therapeutics universe. *Frontiers in Oncology*, **3**, 304.
- 76 Seidman, J.D., Horkayne-Szakaly, I., Haiba, M. et al. (2004) The histologic type and stage distribution of ovarian carcinomas of surface epithelial origin. *International Journal of Gynecological Pathology*, 23, 41–44.
- 77 Bodurka, D.C., Deavers, M.T., Tian, C. et al. (2012) Reclassification of serous ovarian carcinoma by a 2-tier system: a Gynecologic Oncology Group Study. Cancer, 118, 3087–3094.
- 78 Romero, I., Sun, C.C., Wong, K.K. et al. (2013) Low-grade serous carcinoma: new concepts and emerging therapies. Gynecologic Oncology, 130, 660–666.
- 79 Hsu, C.Y., Bristow, R., Cha, M.S. et al. (2004) Characterization of active mitogen-activated protein kinase in ovarian serous carcinomas. Clinical Cancer Research, 10, 6432–6436.
- 80 Pritchard, A.L. and Hayward, N.K. (2013) Molecular pathways: mitogen-activated protein kinase pathway mutations and drug resistance. Clinical Cancer Research, 19, 2301–2309.
- 81 Farley, J., Brady, W.E., Vathipadiekal, V. et al. (2013) Selumetinib in women with recurrent low-grade serous carcinoma of the ovary or peritoneum: an open-label, singlearm, phase 2 study. The Lancet Oncology, 14, 134–140.
- 82 Khasraw, M. and Bell, R. (2012) Primary systemic therapy in HER2-amplified breast cancer: a clinical review. *Expert Review of Anticancer Therapy*, **12**, 1005–1013.
- 83 Chan, D.S., Twine, C.P. and Lewis, W.G. (2012) Systematic review and meta-analysis of the influence of HER2 expression and amplification in operable oesophageal cancer. *Journal of Gastrointestinal Surgery*, **16**, 1821–1829.
- 84 Hechtman, J.F. and Polydorides, A.D. (2012) HER2/neu gene amplification and protein overexpression in gastric and gastroesophageal junction adenocarcinoma: a review of histopathology, diagnostic testing, and clinical implications. Archives of Pathology & Laboratory Medicine, 136, 691–697.
- 85 Spector, N.L. and Blackwell, K.L. (2009) Understanding the mechanisms behind trastuzumab therapy for human epidermal growth factor receptor 2-positive breast cancer. *Journal of Clinical Oncology*, 27, 5838–5847.
- 86 Bookman, M.A., Darcy, K.M., Clarke-Pearson, D. et al. (2003) Evaluation of monoclonal humanized anti-HER2 antibody, trastuzumab, in patients with recurrent or refractory ovarian or primary peritoneal carcinoma with overexpression of HER2: a phase II trial of the Gynecologic Oncology Group. *Journal of Clinical Oncology*, 21, 283–290.
- 87 Berchuck, A., Kamel, A., Whitaker, R. et al. (1990) Overexpression of HER-2/neu is associated with poor survival in advanced epithelial ovarian cancer. Cancer Research, 50, 4087–4091.

- 88 Rubin, S.C., Finstad, C.L., Wong, G.Y. et al. (1993) Prognostic significance of HER-2/neu expression in advanced epithelial ovarian cancer: a multivariate analysis. American Journal of Obstetrics and Gynecology, 168, 162–169.
- 89 Citri, A. and Yarden, Y. (2006) EGF-ERBB signalling: towards the systems level. *Nature Reviews Molecular Cell Biology*, 7, 505–516.
- 90 Yarden, Y. and Sliwkowski, M.X. (2001) Untangling the ErbB signalling network. *Nature Reviews Molecular Cell Biology*, 2, 127–137.
- 91 Baselga, J. (2010) Treatment of HER2-overexpressing breast cancer. Annals of Oncology, 21 Suppl 7, vii36–vii40.
- 92 Baselga, J. (2010) Treatment of HER2-overexpressing breast cancer. *Annals of Oncology*, **21** Suppl 7, vii36–vii40.
- 93 Baselga, J., Gelmon, K.A., Verma, S. et al. (2010) Phase II trial of pertuzumab and trastuzumab in patients with human epidermal growth factor receptor 2-positive metastatic breast cancer that progressed during prior trastuzumab therapy. *Journal of Clinical Oncology*, 28, 1138–1144.
- 94 Blackwell, K.L., Burstein, H.J., Storniolo, A.M. et al. (2012) Overall survival benefit with lapatinib in combination with trastuzumab for patients with human epidermal growth factor receptor 2-positive metastatic breast cancer: final results from the EGF104900 Study. Journal of Clinical Oncology, 30, 2585–2592.
- 95 Swain, S.M., Kim, S.B., Cortes, J. et al. (2013) Pertuzumab, trastuzumab, and docetaxel for HER2-positive metastatic breast cancer (CLEOPATRA study): overall survival results from a randomised, double-blind, placebo-controlled, phase 3 study. The Lancet Oncology, 14, 461–471.
- 96 Baselga, J., Bradbury, I., Eidtmann, H. et al. (2012) Lapatinib with trastuzumab for HER2-positive early breast cancer (NeoALTTO): a randomised, open-label, multicentre, phase 3 trial. Lancet, 379, 633–640.
- 97 Slomovitz, B.M., Broaddus, R.R., Burke, T.W. *et al.* (2004) Her-2/neu overexpression and amplification in uterine papillary serous carcinoma. *Journal of Clinical Oncology*, **22**, 3126–3132.
- 98 El-Sahwi, K., Bellone, S., Cocco, E. et al. (2010) In vitro activity of pertuzumab in combination with trastuzumab in uterine serous papillary adenocarcinoma. British Journal of Cancer, 102, 134–143.
- 99 Galsky, M.D., Von Hoff, D.D., Neubauer, M. et al. (2010) Target-specific, histology-independent, randomized discontinuation study of lapatinib in patients with HER2-amplified solid tumors. *Investigational New Drugs*, 30 (2), 695–701.
- 100 Leslie, K.K., Sill, M.W., Lankes, H.A. et al. (2012) Lapatinib and potential prognostic value of EGFR mutations in a Gynecologic Oncology Group phase II trial of persistent or recurrent endometrial cancer. Gynecologic Oncology, 127, 345–350.

- 101 Fleming, G.F., Sill, M.W., Darcy, K.M. et al. (2010) Phase II trial of trastuzumab in women with advanced or recurrent, HER2-positive endometrial carcinoma: a Gynecologic Oncology Group study. Gynecologic Oncology, 116, 15–20.
- 102 Higgins, M.J. and Baselga, J. (2011) Targeted therapies for breast cancer. The Journal of Clinical Investigation, 121, 3797–3803.
- 103 Pazo Cid, R.A. and Anton, A. (2013) Advanced HER2positive gastric cancer: current and future targeted therapies. Critical Reviews in Oncology/Hematology, 85, 350-362.
- 104 Santin, A.D. (2010) Letter to the Editor referring to the manuscript entitled: "Phase II trial of trastuzumab in women with advanced or recurrent HER-positive endometrial carcinoma: a Gynecologic Oncology Group study" recently reported by Fleming et al., (Gynecologic Oncology, 116;15–20;2010). Gynecologic Oncology, 118, 95–96; author reply 6–7.
- 105 Chay, W.Y., Chew, S.H., Ong, W.S. et al. (2013) HER2 amplification and clinicopathological characteristics in a large Asian cohort of rare mucinous ovarian cancer. PLoS One, 8, e61565.
- 106 Lin, W.L., Kuo, W.H., Chen, F.L. et al. (2011) Identification of the coexisting HER2 gene amplification and novel mutations in the HER2 protein-overexpressed mucinous epithelial ovarian cancer. Annals of Surgical Oncology, 18, 2388–2394.
- 107 Prat, A. and Baselga, J. (2013) Dual human epidermal growth factor receptor 2 (HER2) blockade and hormonal therapy for the treatment of primary HER2-positive breast cancer: one more step toward chemotherapy-free therapy. *Journal of Clinical Oncology*, **31**, 1703–1706.
- 108 Folkman, J. (1971) Tumor angiogenesis: therapeutic implications. The New England Journal of Medicine, 285, 1182– 1186.
- 109 Hurwitz, H., Fehrenbacher, L., Novotny, W. et al. (2004) Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. The New England Journal of Medicine, 350, 2335–2342.
- 110 Sandler, A., Gray, R., Perry, M.C. et al. (2006) Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. The New England Journal of Medicine, 355, 2542–2550.
- 111 Reck, M., von Pawel, J., Zatloukal, P. et al. (2009) Phase III trial of cisplatin plus gemcitabine with either placebo or bevacizumab as first-line therapy for nonsquamous non-small-cell lung cancer: AVAil. Journal of Clinical Oncology, 27, 1227–1234.
- 112 Cortes-Funes, H. (2009) The role of antiangiogenesis therapy: bevacizumab and beyond. Clinical and Translational Oncology, 11, 349–355.

- 113 Giantonio, B.J., Catalano, P.J., Meropol, N.J. et al. (2007) Bevacizumab in combination with oxaliplatin, fluorouracil, and leucovorin (FOLFOX4) for previously treated metastatic colorectal cancer: results from the Eastern Cooperative Oncology Group Study E3200. Journal of Clinical Oncology, 25, 1539–1544.
- 114 Yang, J.C., Haworth, L., Sherry, R.M. et al. (2003) A randomized trial of bevacizumab, an anti-vascular endothelial growth factor antibody, for metastatic renal cancer. The New England Journal of Medicine, 349, 427–434.
- 115 Escudier, B., Pluzanska, A., Koralewski, P. et al. (2007) Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, double-blind phase III trial. *Lancet*, 370, 2103–2111.
- 116 Escudier, B., Eisen, T., Stadler, W.M. et al. (2007) Sorafenib in advanced clear-cell renal-cell carcinoma. The New England Journal of Medicine, 356, 125–134.
- 117 Motzer, R.J., Rini, B.I., Bukowski, R.M. et al. Sunitinib in patients with metastatic renal cell carcinoma. *JAMA*, 295, 2516–2524.
- 118 Motzer, R.J., Hutson, T.E., Tomczak, P. et al. (2007) Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. The New England Journal of Medicine, 356, 115–124.
- 119 Vredenburgh, J.J., Desjardins, A., Herndon, J.E., 2nd et al. (2007) Bevacizumab plus irinotecan in recurrent glio-blastoma multiforme. Journal of Clinical Oncology, 25, 4722–4729.
- 120 Norden, A.D., Young, G.S., Setayesh, K. et al. (2008) Bevacizumab for recurrent malignant gliomas: efficacy, toxicity, and patterns of recurrence. Neurology, 70, 779–787.
- 121 Ranieri, G., Patruno, R., Ruggieri, E. et al. (2006) Vascular endothelial growth factor (VEGF) as a target of bevacizumab in cancer: from the biology to the clinic. Current Medicinal Chemistry, 13, 1845–1857.
- 122 Rasila, K.K., Burger, R.A., Smith, H. et al. (2005) Angiogenesis in gynecological oncology-mechanism of tumor progression and therapeutic targets. *International Journal of Gynecological Cancer*, 15, 710–726.
- 123 Spannuth, W.A., Sood, A.K. and Coleman, R.L. (2008) Angiogenesis as a strategic target for ovarian cancer therapy. Nature Clinical Practice Oncology, 5, 194–204.
- 124 Li, X. and Eriksson, U. (2001) Novel VEGF family members: VEGF-B, VEGF-C and VEGF-D. International Journal of Biochemistry & Cell Biology, 33, 421–426.
- 125 Kerbel, R.S. (2008) Tumor angiogenesis. The New England Journal of Medicine, 358, 2039–2049.
- 126 Fauconnet, S., Bernardini, S., Lascombe, I. et al. (2009) Expression analysis of VEGF-A and VEGF-B: relationship with clinicopathological parameters in bladder cancer. Oncology Reports, 21, 1495–1504.

- 127 Hopfl, G., Ogunshola, O., and Gassmann, M. (2004) HIFs and tumors–causes and consequences. *American Journal of Physiology-Regulatory Integrative and Comparative Physiology*, **286**, R608–R623.
- 128 Monk, B.J., Willmott, L.J. and Sumner, D.A. (2010) Antiangiogenesis agents in metastatic or recurrent cervical cancer. *Gynecologic Oncology*, 116, 181–186.
- 129 Kumaran, G.C., Jayson, G.C. and Clamp, A.R. (2009) Antiangiogenic drugs in ovarian cancer. *British Journal of Cancer*, 100, 1–7.
- 130 Alvarez, A.A., Krigman, H.R., Whitaker, R.S. et al. (1999) The prognostic significance of angiogenesis in epithelial ovarian carcinoma. Clinical Cancer Research, 5, 587–591.
- 131 Goodheart, M.J., Vasef, M.A., Sood, A.K. et al. (2002) Ovarian cancer p53 mutation is associated with tumor microvessel density. Gynecologic Oncology, 86, 85–90.
- 132 Abulafia, O., Triest, W.E. and Sherer, D.M. (1997) Angiogenesis in primary and metastatic epithelial ovarian carcinoma. American Journal of Obstetrics and Gynecology, 177, 541–547.
- 133 Gadducci, A., Viacava, P., Cosio, S. et al. (2003) Intratumoral microvessel density, response to chemotherapy and clinical outcome of patients with advanced ovarian carcinoma. Anticancer Research, 23, 549–556.
- 134 Hollingsworth, H.C., Kohn, E.C., Steinberg, S.M. et al. (1995) Tumor angiogenesis in advanced stage ovarian carcinoma. The American Journal of Pathology, 147, 33–41.
- 135 Rubatt, J.M., Darcy, K.M., Hutson, A. et al. (2009) Independent prognostic relevance of microvessel density in advanced epithelial ovarian cancer and associations between CD31, CD105, p53 status, and angiogenic marker expression: a Gynecologic Oncology Group study. Gynecologic Oncology, 112, 469–474.
- 136 Papetti, M. and Herman, I.M. (2002) Mechanisms of normal and tumor-derived angiogenesis. *American Journal* of Physiology Cell Physiology, 282, C947–C970.
- 137 Duhoux, F.P. and Machiels, J.P. (2010) Antivascular therapy for epithelial ovarian cancer. *Journal of Oncology*, 2010, 372547.
- 138 Monk, B.J., Choi, D.C., Pugmire, G. and Burger, R.A. (2005) Activity of bevacizumab (rhuMAB VEGF) in advanced refractory epithelial ovarian cancer. *Gynecologic Oncology*, 96, 902–905.
- 139 Bidus, M.A., Webb, J.C., Seidman, J.D. et al. (2006) Sustained response to bevacizumab in refractory well-differentiated ovarian neoplasms. Gynecologic Oncology, 102, 5–7.
- 140 Cannistra, S.A., Matulonis, U.A., Penson, R.T. et al. (2007) Phase II study of bevacizumab in patients with platinumresistant ovarian cancer or peritoneal serous cancer. *Journal* of Clinical Oncology, 25, 5180–5186.

- 141 Burger, R.A., Sill, M.W., Monk, B.J. et al. (2007) Phase II trial of bevacizumab in persistent or recurrent epithelial ovarian cancer or primary peritoneal cancer: a Gynecologic Oncology Group Study. Journal of Clinical Oncology, 25, 5165–5171.
- 142 Nimeiri, H.S., Oza, A.M., Morgan, R.J. et al. (2008) Efficacy and safety of bevacizumab plus erlotinib for patients with recurrent ovarian, primary peritoneal, and fallopian tube cancer: a trial of the Chicago, PMH, and California Phase II Consortia. Gynecologic Oncology, 110, 49–55.
- 143 Garcia, A.A., Hirte, H., Fleming, G. et al. (2008) Phase II clinical trial of bevacizumab and low-dose metronomic oral cyclophosphamide in recurrent ovarian cancer: a trial of the California, Chicago, and Princess Margaret Hospital phase II consortia. Journal of Clinical Oncology, 26, 76–82.
- 144 Aghajanian, C., Blank, S.V., Goff, B.A. *et al.* (2012) OCEANS: a randomized, double-blind, placebo-controlled phase iii trial of chemotherapy with or without bevacizumab in patients with platinum-sensitive recurrent epithelial ovarian, primary peritoneal, or fallopian tube cancer. *Journal of Clinical Oncology*, **31**, 166–167.
- 145 Aghajanian, C.A., Finkler, N.J., Rutherford, T. et al. (2011) OCEANS: a randomized, double-blinded, placebocontrolled phase III trial of chemotherapy with or without bevacizumab (BEV) in patients with platinum-sensitive recurrent epithelial ovarian (EOC), primary peritoneal (PPC), or fallopian tube cancer (FTC). Journal of Clinical Oncology, 29.
- 146 Monk, B.J., Pujade-Lauraine, E. and Burger, R.A. (2013) Integrating bevacizumab into the management of epithelial ovarian cancer: the controversy of front-line versus recurrent disease. *Annals of Oncology*, 24 (Suppl 10), x53–x58.
- 147 Burger, R.A. (2007) Experience with bevacizumab in the management of epithelial ovarian cancer. *Journal of Clinical Oncology*, 25, 2902–2908.
- 148 Coleman, R.L., Duska, L.R., Ramirez, P.T. et al. (2011) Phase 1-2 study of docetaxel plus aflibercept in patients with recurrent ovarian, primary peritoneal, or fallopian tube cancer. The Lancet Oncology, 12, 1109–1117.
- 149 Bodnar, L., Gornas, M. and Szczylik, C. (2011) Sorafenib as a third line therapy in patients with epithelial ovarian cancer or primary peritoneal cancer: a phase II study. *Gynecologic Oncology*, 123, 33–36.
- 150 Matei, D., Sill, M.W., Lankes, H.A. et al. (2011) Activity of sorafenib in recurrent ovarian cancer and primary peritoneal carcinomatosis: a gynecologic oncology group trial. *Journal of Clinical Oncology*, 29, 69–75.
- 151 Burger, R.A. (2011) Overview of anti-angiogenic agents in development for ovarian cancer. *Gynecologic Oncology*, 121, 230–238.

- 152 Biagi, J.J., Oza, A.M., Chalchal, H.I. et al. (2011) A phase II study of sunitinib in patients with recurrent epithelial ovarian and primary peritoneal carcinoma: an NCIC Clinical Trials Group Study. Annals of Oncology, 22, 335–340.
- 153 Friedlander, M., Hancock, K.C., Rischin, D. et al. (2010) A Phase II, open-label study evaluating pazopanib in patients with recurrent ovarian cancer. Gynecologic Oncology, 119, 32–37.
- 154 Merritt, W.M., Nick, A.M., Carroll, A.R. et al. (2010) Bridging the gap between cytotoxic and biologic therapy with metronomic topotecan and pazopanib in ovarian cancer. Molecular Cancer Therapeutics, 9, 985–995.
- 155 Kobold, S., Hegewisch-Becker, S., Oechsle, K. et al. (2009) Intraperitoneal VEGF inhibition using bevacizumab: a potential approach for the symptomatic treatment of malignant ascites? The Oncologist, 14, 1242–1251.
- 156 Numnum, T.M., Rocconi, R.P., Whitworth, J. and Barnes, M.N. (2006) The use of bevacizumab to palliate symptomatic ascites in patients with refractory ovarian carcinoma. *Gynecologic Oncology*, 102, 425–428.
- 157 Cohn, D.E., Valmadre, S., Resnick, K.E. et al. (0000) Bevacizumab and weekly taxane chemotherapy demonstrates activity in refractory ovarian cancer. Gynecologic Oncology, 102, 134–139.
- 158 Hamilton, C.A., Maxwell, G.L., Chernofsky, M.R. et al. (2008) Intraperitoneal bevacizumab for the palliation of malignant ascites in refractory ovarian cancer. Gynecologic Oncology, 111, 530–532.
- 159 Micha, J.P., Goldstein, B.H., Rettenmaier, M.A. et al. (2007) A phase II study of outpatient first-line paclitaxel, carboplatin, and bevacizumab for advanced-stage epithelial ovarian, peritoneal, and fallopian tube cancer. International Journal of Gynecological Cancer, 17, 771–776.
- 160 Penson, R.T., Dizon, D.S., Cannistra, S.A. et al. (2010) Phase II study of carboplatin, paclitaxel, and bevacizumab with maintenance bevacizumab as first-line chemotherapy for advanced mullerian tumors. *Journal of Clinical Oncology*, 28, 154–159.
- 161 Perren, T.J., Swart, A.M., Pfisterer, J. et al. (2011) A phase 3 trial of bevacizumab in ovarian cancer. The New England Journal of Medicine, 365, 2484–2496.
- 162 Burger, R.A., Brady, M.F., Bookman, M.A. et al. Incorporation of bevacizumab in the primary treatment of ovarian cancer. The New England Journal of Medicine, 365, 2473–2483.
- 163 Cohn, D.E., Kim, K.H., Resnick, K.E. et al. (2011) At what cost does a potential survival advantage of bevacizumab make sense for the primary treatment of ovarian cancer? A cost-effectiveness analysis. *Journal of Clinical Oncology*, 29, 1247–1251.

- 164 Abulafia, O., Triest, W.E., Sherer, D.M. *et al.* Angiogenesis in endometrial hyperplasia and stage I endometrial carcinoma. *Obstetrics & Gynecology*, **86**, 479–485.
- 165 Kaku, T., Kamura, T., Kinukawa, N. et al. (1997) Angiogenesis in endometrial carcinoma. Cancer, 80, 741–747.
- 166 Kirschner, C.V., Alanis-Amezcua, J.M., Martin, V.G. et al. (1996) Angiogenesis factor in endometrial carcinoma: a new prognostic indicator? American Journal of Obstetrics & Gynecology, 174, 1879–1882; discussion 82–84.
- 167 Nimeiri, H.S., Oza, A.M., Morgan, R.J. et al. (2010) A phase II study of sorafenib in advanced uterine carcinoma/carcinosarcoma: a trial of the Chicago, PMH, and California Phase II Consortia. Gynecologic Oncology, 117, 37–40.
- 168 Correa, R., Mackay, H., Hirte, H.W. et al. (2010) A phase II study of sunitinib in recurrent or metastatic endometrial carcinoma: a trial of the Princess Margaret Hospital, The University of Chicago, and California Cancer Phase II Consortia. Journal of Clinical Oncology, 28, 15s.
- 169 Aghajanian, C., Sill, M.W., Darcy, K.M. et al. (2011) Phase II trial of bevacizumab in recurrent or persistent endometrial cancer: a Gynecologic Oncology Group study. *Journal of Clinical Oncology*, 29, 2259–2265.
- 170 Stafl, A. and Mattingly, R.F. (1975) Angiogenesis of cervical neoplasia. American Journal of Obstetrics & Gynecology, 121, 845–852.
- 171 Sillman, F., Boyce, J. and Fruchter, R. (1981) The significance of atypical vessels and neovascularization in cervical neoplasia. American Journal of Obstetrics & Gynecology, 139, 154–159.
- 172 Willmott, L.J. and Monk, B.J. (2009) Cervical cancer therapy: current, future and anti-angiogensis targeted treatment. Expert Review of Anticancer Therapy, 9, 895–903.
- 173 Smith-McCune, K. (1997) Angiogenesis in squamous cell carcinoma in situ and microinvasive carcinoma of the uterine cervix. *Obstetrics & Gynecology*, **89**, 482–483.
- 174 Dellas, A., Moch, H., Schultheiss, E. et al. (1997) Angiogenesis in cervical neoplasia: microvessel quantitation in precancerous lesions and invasive carcinomas with clinicopathological correlations. Gynecologic Oncology, 67, 27–33.
- 175 Dinh, T.V., Hannigan, E.V., Smith, E.R. et al. (1996) Tumor angiogenesis as a predictor of recurrence in stage Ib squamous cell carcinoma of the cervix. Obstetrics & Gynecology, 87, 751–754.
- 176 Schlenger, K., Hockel, M., Mitze, M., et al. (1995) Tumor vascularity–a novel prognostic factor in advanced cervical carcinoma. Gynecologic Oncology, 59, 57–66.
- 177 Santin, A.D., Hermonat, P.L., Ravaggi, A. et al. (1999) Secretion of vascular endothelial growth factor in adenocarcinoma and squamous cell carcinoma of the uterine cervix. Obstetrics and Gynecology, 94, 78–82.

- 178 Loncaster, J.A., Cooper, R.A., Logue, J.P. et al. (2000) Vascular endothelial growth factor (VEGF) expression is a prognostic factor for radiotherapy outcome in advanced carcinoma of the cervix. British Journal of Cancer, 83, 620– 625.
- 179 Cheng, W.F., Chen, C.A., Lee, C.N. *et al.* (1999) Vascular endothelial growth factor in cervical carcinoma. *Obstetrics & Gynecology*, **93**, 761–765.
- 180 Cheng, W.F., Chen, C.A., Lee, C.N. *et al.* (2000) Vascular endothelial growth factor and prognosis of cervical carcinoma. *Obstetrics & Gynecology*, **96**, 721–726.
- 181 Monk, B.J., Mas Lopez, L., Zarba, J.J. et al. (2010) Phase II, open-label study of pazopanib or lapatinib monotherapy compared with pazopanib plus lapatinib combination therapy in patients with advanced and recurrent cervical cancer. *Journal of Clinical Oncology*, 28, 3562–3569.
- 182 Monk, B.J., Sill, M.W., Burger, R.A. et al. (2009) Phase II trial of bevacizumab in the treatment of persistent or recurrent squamous cell carcinoma of the cervix: a gynecologic oncology group study. Journal of Clinical Oncology, 27, 1069–1074.
- 183 Mackay, H.J., Tinker, A., Winquist, E. et al. (2010) A phase II study of sunitinib in patients with locally advanced or metastatic cervical carcinoma: NCIC CTG Trial IND.184. Gynecologic Oncology, 116, 163–167.
- 184 Risch, H.A., McLaughlin, J.R., Cole, D.E. et al. (2006) Population BRCA1 and BRCA2 mutation frequencies and cancer penetrances: a kin-cohort study in Ontario, Canada. *Journal of National Cancer Institute*, 98, 1694–1706.
- 185 Hennessy, B.T., Coleman, R.L. and Markman, M. (2009) Ovarian cancer. *Lancet*, 374, 1371–1382.
- 186 Berchuck, A., Cirisano, F., Lancaster, J.M. et al. (1996) Role of BRCA1 mutation screening in the management of familial ovarian cancer. American Journal of Obstetrics and Gynecology, 175, 738–746.
- 187 Narod, S.A. and Boyd, J. (2002) Current understanding of the epidemiology and clinical implications of BRCA1 and BRCA2 mutations for ovarian cancer. Current Opinion in Obstetrics & Gynecology, 14, 19–26.
- 188 Boyd, J., Sonoda, Y., Federici, M.G. et al. (2000) Clinicopathologic features of BRCA-linked and sporadic ovarian cancer. JAMA, 283, 2260–2265.
- 189 Moynahan, M.E., Cui, T.Y. and Jasin, M. (2001) Homology-directed dna repair, mitomycin-c resistance, and chromosome stability is restored with correction of a Brca1 mutation. Cancer Research, 61, 4842–4850.
- 190 Bryant, H.E., Schultz, N., Thomas, H.D. et al. (2005) Specific killing of BRCA2-deficient tumours with inhibitors of poly(ADP-ribose) polymerase. *Nature*, 434, 913–917.
- 191 Wooster, R. and Weber, B.L. (2003) Breast and ovarian cancer. The New England Journal of Medicine, 348, 2339–2347.

- 192 Farmer, H., McCabe, N., Lord, C.J. et al. (2005) Targeting the DNA repair defect in BRCA mutant cells as a therapeutic strategy. Nature, 434, 917–921.
- 193 Sandhu, S.K., Yap, T.A. and de Bono, J.S. (2010) Poly(ADPribose) polymerase inhibitors in cancer treatment: a clinical perspective. *European Journal of Cancer*, 46, 9–20.
- 194 Ashworth, A. (2008) A synthetic lethal therapeutic approach: poly(ADP) ribose polymerase inhibitors for the treatment of cancers deficient in DNA double-strand break repair. *Journal of Clinical Oncology*, 26, 3785–3790.
- 195 Dobzhansky, T. (1946) Genetics of natural populations. Xiii. Recombination and variability in populations of drosophila pseudoobscura. *Genetics*, 31, 269–290.
- 196 Hartwell, L.H., Szankasi, P., Roberts, C.J. et al. (1997) Integrating genetic approaches into the discovery of anticancer drugs. Science (New York, NY), 278, 1064–1068.
- 197 Clark, J.B., Ferris, G.M. and Pinder, S. (1971) Inhibition of nuclear NAD nucleosidase and poly ADP-ribose polymerase activity from rat liver by nicotinamide and 5'-methyl nicotinamide. *Biochimica et Biophysica Acta*, 238, 82–85.
- 198 Fong, P.C., Boss, D.S., Yap, T.A. et al. (2009) Inhibition of poly(ADP-ribose) polymerase in tumors from BRCA mutation carriers. The New England Journal of Medicine, 361, 123–134.
- 199 Fong, P.C., Yap, T.A., Boss, D.S. et al. (2010) Poly(ADP)ribose polymerase inhibition: frequent durable responses in BRCA carrier ovarian cancer correlating with platinumfree interval. *Journal of Clinical Oncology*, 28, 2512–2519.
- 200 Audeh, M.W., Carmichael, J., Penson, R.T. et al. (2010) Oral poly(ADP-ribose) polymerase inhibitor olaparib in patients with BRCA1 or BRCA2 mutations and recurrent ovarian cancer: a proof-of-concept trial. *Lancet*, 376, 245–251.
- 201 Gelmon, K.A., Tischkowitz, M., Mackay, H. et al. (2011) Olaparib in patients with recurrent high-grade serous or poorly differentiated ovarian carcinoma or triple-negative breast cancer: a phase 2, multicentre, open-label, nonrandomised study. The Lancet Oncology, 12, 852–861.
- 202 O'Shaughnessy, J.O. (2009) Efficacy of BSI-201, a poly (ADP-ribose) polymerase-1 (PARP1) inhibitor, in combination with gemcitabine/carboplatin (G/C) in patients with metastatic triple-negative breast cancer (TNBC): results of a randomized phase II trial. American Society for Clinical Oncology, 2009.
- 203 Dent, R., Trudeau, M., Pritchard, K.I. et al. (2007) Triplenegative breast cancer: clinical features and patterns of recurrence. Clinical Cancer Research, 13, 4429–4434.
- 204 Foulkes, W.D., Stefansson, I.M., Chappuis, P.O. et al. (2003) Germline BRCA1 mutations and a basal epithelial phenotype in breast cancer. *Journal of National Cancer Institute*, 95, 1482–1485.

- 205 Press, J.Z., De Luca, A., Boyd, N. et al. (2008) Ovarian carcinomas with genetic and epigenetic BRCA1 loss have distinct molecular abnormalities. BMC Cancer, 8, 17.
- 206 Konstantinopoulos, P.A., Spentzos, D., Karlan, B.Y. et al. (2010) Gene expression profile of BRCAness that correlates with responsiveness to chemotherapy and with outcome in patients with epithelial ovarian cancer. *Journal of Clinical Oncology*, 28, 3555–3561.
- 207 Turner, N., Tutt, A. and Ashworth, A. (2004) Hallmarks of 'BRCAness' in sporadic cancers. *Nature Reviews Cancer*, 4, 814–819.
- 208 Cancer Genome Atlas Research Network (2011) Integrated genomic analyses of ovarian carcinoma. *Nature*, 474, 609–615.
- 209 Lin, Z.P., Lee, Y., Lin, F. et al. (2011) Reduced level of ribonucleotide reductase R2 subunits increases dependence on homologous recombination repair of cisplatin-induced DNA damage. Molecular Pharmacology, 80, 1000–1012.
- 210 Zdraveski, Z.Z., Mello, J.A., Marinus, M.G. and Essigmann, J.M. (2000) Multiple pathways of recombination define cellular responses to cisplatin. *Chemisty & Biology*, 7, 39–50.

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CHAPTER 2

Imaging of Rare Gynecologic Tumors

Hima B. Prabhakar¹ and Dinushi S. Perera²

¹Perelman School of Medicine of the University of Pennsylvania, Division of Body Imaging, Pennsylvania Hospital, Philadelphia, PA, USA

Imaging guidelines

Imaging plays a central role in the evaluation of patients with symptoms and clinical presentations suggesting underlying pelvic pathology. As many patients present with nonspecific symptoms (i.e., pain, bloating, bleeding), any patient with concerning or unclear physical evaluation findings will typically undergo radiologic evaluation. In general, depending on the patient's symptoms, this initial radiologic evaluation will be performed with ultrasound. Occasionally, computed tomography (CT) may be a good initial choice in some patients, particularly if a large mass or large volume ascites is present on physical examination, for which evaluation by ultrasound can be limited. Magnetic resonance imaging (MRI) is typically utilized as a follow-up to ultrasound [1]. 18F-fluorodeoxyglucose positron emission tomography—CT (PET/CT) is typically reserved for patients with known malignancy—for initial staging, evaluation of treatment response, and evaluation for recurrent or metastatic disease [2-4].

Ultrasound is an optimal first step in the evaluation of patients with suspected pelvic pathology. It is quick to perform, and is without ionizing radiation; however, it is operator-dependent [5]. Ultrasound is particularly useful in evaluation of cystic and solid lesions of the pelvis, as it is able to demonstrate any complexity or nodularity of cystic lesions, as well as evaluate for vascular flow within any solid components. Ultrasound of the pelvis should

optimally be performed with transabdominal and transvaginal technique, unless the patient is unable to tolerate the endovaginal probe [6]. Potential drawbacks of ultrasound include suboptimal evaluation of the collapsed vagina and vulva, though these can sometimes be evaluated using translabial ultrasound [7]. Additionally, in patients with large adnexal or pelvic masses, ultrasound may be limited in determining the organ of origin, given the limited scope of imaging field. Once the patient has a known malignancy, ultrasound is insensitive in staging, and CT or MRI should be utilized [3].

CT is an excellent initial imaging modality in any patient who presents with physical examination findings suggesting large abdominopelvic mass, large volume ascites, or signs of pathology not confined to the pelvis. It is easily available and scanning is quick (usually <1 min) [5]. In these patients, CT should be performed with both intravenous and oral contrast if the patient's renal function allows. In particular, the use of intravenous contrast allows for evaluation of abdominal/pelvic parenchymal organ enhancement, as well as the assessment of enhancing masses. Oral contrast agents allow for delineation of the bowel; this is particularly useful in allowing for any potential cystic lesions to be differentiated from bowel, as particularly small bowel can lie within the pelvis and occasionally demonstrate fluid distention, making such distinction difficult. CT utilizes ionizing radiation, which can be a concern particularly in younger patients [1].

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²Pennsylvania Hospital, University of Pennsylvania Health System, Philadelphia, PA, USA