Developments in Applied Phycology 13

Alfredo de Jesús Martínez-Roldán Editor

Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures



Developments in Applied Phycology

Volume 13

Series Editor

Michael A. Borowitzka, Algae R&D Centre, School of Veterinary and Life Sciences, Murdoch University, Murdoch, WA, Australia

Aims and Scope

Applied Phycology, the practical use of algae, encompasses a diverse range of fields including algal culture and seaweed farming, the use of algae to produce commercial products such as hydrocolloids, carotenoids and pharmaceuticals, algae as biofertilizers and soil conditioners, the application of algae in wastewater treatment, renewable energy production, algae as environmental indicators, environmental bioremediation and the management of algal blooms. The commercial production of seaweeds and microalgae and products derived there from is a large and well established industry and new algal species, products and processes are being continuously developed.

The aim of this book series, Developments in Applied Phycology, is to present state-of-theart syntheses of research and development in the field. Volumes of the series will consist of reference books, subject-specific monographs, peer reviewed contributions from conferences, comprehensive evaluations of large-scale projects, and other book-length contributions to the science and practice of applied phycology.

Prospective authors and/or editors should consult the Series Editor or Publishing Editor for more details. Series Editor: Michael A. Borowitzka - M.Borowitzka@murdoch.edu.au Publishing Editor: Éva Loerinczi - eva.loerinczi@springer.com

Alfredo de Jesús Martínez-Roldán Editor

Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures



Alfredo de Jesús Martínez-Roldán Investigador Nacional Nivel I, Catedrático CONAHCYT - TecNM/ITD Departamento de Ingenierías Química y Bioquímica División de Estudios de Posgrado Maestría en Sistemas Ambientales Durango, Mexico

ISSN 2543-0599 ISSN 2543-0602 (electronic) Developments in Applied Phycology ISBN 978-3-031-43968-1 ISBN 978-3-031-43969-8 (eBook) https://doi.org/10.1007/978-3-031-43969-8

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Switzerland AG 2024 This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Switzerland AG The registered company address is: Gewerbestrasse 11, 6330 Cham, Switzerland

Paper in this product is recyclable.

Editor

To Sofía, María Paula, Arya, and Jorge

Acknowledgement

Thanks to all those who at some point arrived and remained, to those who arrived and left, and to all those who will arrive.

Gracias a todos aquellos que en algún momento llegaron y permanecen, a los que llegaron y se marcharon y a todos aquellos que llegarán

Contents

1	Introduction to Environment-Friendly Bioprocesses by Microalgae and Cyanobacteria	1
2	CO₂ Bio-capture by Microalgae and Cyanobacteria Cultures Cigdem Demirkaya and Hector De la Hoz Siegler	5
3	Removal of Nitrogen and Phosphorus from DomesticWastewater by Microalgal CulturesMaría I. Ospina and Mohamed T. Darwich-Cedeño	19
4	Wastewater Treatment by Microalgae and Cyanobacteria.The Reduction of the Contaminant Potential of Real DomesticWastewater: A Case StudyAlfredo de Jesús Martínez-Roldán, Rebeca Paola Villanueva-Garcia,María Dolores Josefina Rodríguez Rosales, Sergio Valle Cervantes,and Hugo Virgilio Perales Vela	31
5	Phycoremediation of Industrial Wastewater	43
6	Treatment of Industrial Wastewater with Microalgae Rodríguez-Palacio Mónica Cristina, Lozano-Ramírez Cruz, and Martínez-Hernández Marisol	57
7	Biofertilization by Nitrogen-Fixing Cyanobacteria, Nutrient Supplementation, and Growth Promotion Rosa Olivia Cañizares-Villanueva, Citlally Ramírez-López, Pablo A. López-Pérez, and Dulce J. Hernández-Melchor	69
8	Biodiesel Production by Microalgal Biomass and Strategies to Improve Its Quality Martha Trinidad Arias Peñaranda	83
9	Dark Fermentation of Microalgae and Cyanobacteria for HydrogenProductionCigdem Demirkaya and Hector De la Hoz Siegler	99
10	Induction of Carotenoid Synthesis in Microalgae with Reference to Their Production Outdoors Cecilia Faraloni and Giuseppe Torzillo	113
11	Effect of Dietary Supplementation with Microalgae Biomass on Gastrointestinal Tract Health	125

12	Sustainable Production of Diatom-Based Omega-3 Fatty Acids Aishwarya Mogal, Sarvjeet Kukreja, and Shristy Gautam	131
13	Metabolites from Microalgal Cultures as Potential Sources for thePharmaceutical IndustryMaría Luján Flores, Mariana Jiménez-Veuthey, and Osvaldo León Córdoba	139
14	Microalgae and Cyanobacteria Antioxidant Capacity under Stress Conditions	169
15	Biogas Obtained from the Anaerobic Digestion of Microalgal Biomass Alfredo de Jesús Martínez-Roldán, M. D. J. Rodríguez Rosales, and S. Valle Cervantes	181
16	Microalgal Biomass-Derived Biochars Eduardo Fuentes-Quezada, Alfredo de Jesús Martínez-Roldán, and Diana Cristina Martínez-Casillas	195
17	Process Integration via a Sustainable Biorefinery Approach Using Agro-industrial Residues and Photosynthetic Consortia Pablo A. López-Pérez, Dulce J. Hernández-Melchor, Lizeth Vanessa Hernández Quijano, Mónica Ivette Sánchez Contreras, and R. Icela Beltrán-Hernández	213
18	Attached Biofilm Cultivation (ABC) of Mixotrophic Microalgae for the Sustainable Supply of Innovative New Bioproducts Linda O'Higgins and Imen Hamed	229
19	Interactions Between Organizations Regarding Managementof River Water Basins at a Socially Vulnerable ContextPatricia L. Marconi and Laura I. de Cabo	245

х



Introduction to Environment-Friendly Bioprocesses by Microalgae and Cyanobacteria

Alfredo de Jesús Martínez-Roldán

Abstract

Microalgae include diverse organisms with different cellular structures (prokaryotic and eukaryotic), the capability to grow in diverse ecosystems (sea, rivers, lakes, lagoons, soil, etc.), and the possibility of performing autotrophic, mixotrophic, and heterotrophic metabolism. This diversity is the reason for their ability to produce and accumulate different compounds, many of which have the potential to be used in industrial processes. These compounds include lipids, proteins, amino acids, carotenoids, biofuels, adsorbents (carbons), polyunsaturated fatty acids, and animal feeds. In addition, processes based on microalgae can capture carbon dioxide (CO_2) , eliminate pollutants (such as nitrogen, phosphorous, and heavy metals), or even utilize biomass as feed for livestock. Nevertheless, the growth conditions, induction process, and extraction and purification strategies are specific to every strain. This book aims to include recent developments in environment-friendly processes derived from microalgae and cyanobacteria.

Keywords

Microalgae · Cyanobacteria · High-value bioproducts · Biotechnology

1.1 Introduction

Microalgae are photosynthetic microorganisms with many metabolic pathways very similar to superior plants; nevertheless, they have several advantages compared with terrestrial plants, such as the possibility to be cultivated and reaching

e-mail: adjmartinezro@conacyt.mx

massive cultures in photobioreactors and the fact that the development of fruits, seeds, or a specific tissue is not necessary, as in the case of vegetable crops, to obtain a high-value product (Barsanti and Gualtieri 2014). Historically, microalgae have been used in experimental studies to elucidate metabolic pathways, specifically for the description of oxygenic photosynthesis. The experiments performed to describe the route of carbon fixation in oxygenic photosynthesis, commonly known as the Calvin–Benson–Bassham cycle, were developed using *Chlorella* cultures exposed to light for small periods and subsequently inactivated by dropping in hot methanol. This experiment allowed us to determine all the molecules produced in the Calvin–Benson cycle and describe all the chemical reactions involved (Biel and Fomina 2015).

Since then, microalgae and cyanobacteria have been proposed to develop diverse bioprocesses with two main objectives: eliminating different pollutants from liquid and gaseous effluents and producing high-value products by taking advantage of specific metabolic pathways (Borowitzka 2013). Recently, genetic modifications of microalgae and cyanobacteria were carried out to either increase the amount of a specific metabolite or improve the performance of the culture under special operational conditions (Barati et al. 2021; Beacham et al. 2017).

Around the 1950s, the majority of the research related to microalgae and cyanobacteria focused on their role in the facultative lagoon and the tertiary treatment of wastewaters, as symbiosis was observed between aerobic bacteria and photosynthetic microorganisms and an increase in the efficiency of organic matter removal was observed in the presence of microalgae (Oswald et al. 1957; Oswald and Golueke 1960; Oswald and Gotaas 1957).

Later, the development of technological devices for microalgal culture started; their containers were called photobioreactors and today their variety is huge (Martínez-Roldán and Cañizares-Villanueva 2015). Some photobioreactors include configurations, such as fermenters,

A. de Jesús Martínez-Roldán (🖂)

Investigador Nacional Nivel I, Catedrático CONAHCYT - TecNM/ITD, Departamento de Ingenierías Química y Bioquímica, División de Estudios de Posgrado, Maestría en Sistemas Ambientales, Durango, Mexico

[©] The Author(s), under exclusive license to Springer Nature Switzerland AG 2024

A. de Jesús Martínez-Roldán (ed.), *Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures*, Developments in Applied Phycology 13, https://doi.org/10.1007/978-3-031-43969-8_1

tubular horizontal and/or vertical airlift mixing, columns, flat panels, and thin layers, all of which satisfy specific culturing requirements of the microorganisms, such as mixing, shear stress, and light supply. In all configurations, the main objective is to maximize biomass production, biomass productivity, or even the production of a specific metabolite or a highvalue product (Acién et al. 2017; Chini Zittelli et al. 2013; Torzillo and Chini Zittelli 2015). Recently, microalgal biotechnology has focused on environmental applications or the production of high-value bioproducts, but always from a sustainability perspective.

Some environmental applications take advantage of the diverse qualities of microalgal cultures, e.g., their capability to fix carbon dioxide (CO₂), which is useful for the developing processes to capture CO₂ or to reduce the CO₂ concentration in fuel gases from diverse industrial processes. However, there are no technological developments at the commercial scale, because there are numerous obstacles related to the engendering of the process (Solovchenko and Khozin-Goldberg 2013; Wang et al. 2008; Zhou et al. 2017). Another characteristic used for environmental applications of microalgae is the fast consumption of nutrients from its culture media (nitrogen and phosphorus), which has been fully studied, allowing us to describe the role of microalgae and cyanobacteria in the stabilization of lagoons or even the use of microalgal cultures for the tertiary treatment of domeswastewater. The inclusion of microalgae tic and cyanobacteria in wastewater treatment permits the elimination of nitrogen and phosphorus, which cannot be eliminated by aerobic and anaerobic processes for organic matter elimination (Martínez-Roldán and Ibarra-Berumen 2019; Olguin 2003).

With regard to the use of microalgae and cyanobacteria in wastewater treatment, the proposal is to eliminate specific contaminants from water, some of which are heavy metals and semimetals. Microalgal biomass has a huge potential for the removal of ions because it is possible to use both live and dead biomass as adsorbents. The use and process of microalgal biomass as ion adsorbents is very efficient, and recovery of the removed ions is quite simple (Cañizares-Villanueva 2000; Perales-Vela et al. 2006). Owing to their capability to remove pollutants from the culture medium, the microalgae are proposed to eliminate specific pollutants recently detected in urban wastewaters and denominated as emerging contaminants; the major problem with this type of compound is its wide variety because the sources are very diverse (Peña-Guzmán et al. 2019).

Some emerging pollutants have actually reached high concentrations and are further increasing, causing concern to the scientific community. Therefore, many studies have focused on the development of processes to eliminate them. The emerging contaminants include colorants, drugs, hormones, healthcare products, cosmetics, and antibiotics. Microalgae have proven to eliminate the contaminants by the process of adsorption/absorption or even biotransformation; however, in this case, it is possible to obtain subproducts with higher toxicity than the original ones (Geissen et al. 2015; Jain et al. 2022; Keen et al. 2014; Peña-Guzmán et al. 2019). Therefore, regardless of the potential of microalgae and cyanobacteria to eliminate these contaminants, there are no commercial-scale treatment processes, and there is an unknown economic cost and real efficiency.

The potential environmental application of microalgae is not only the elimination of pollutants from liquid and gaseous effluents. Since the biomass is a source of a large number of different molecules, several of them have high market value (Borowitzka 2013). Some of these bioproducts include pigments, antioxidants, fatty acids, oils, polyunsaturated fatty acids, and the lipid fraction of the biomass, which can be converted into liquid fuels, such as biodiesel or jet fuel (Cañizares-Villanueva et al. 2022).

The high-value bioproducts are very diverse, but some of them have higher potential for pigment production because there are strains with the capability to produce high amounts of different carotenoids or xanthophylls, such as betacarotene. astaxanthin. lutein. violaxanthin. and antheraxanthin, as well as many other molecules with similar chemical properties. In addition, it is possible to obtain molecules with antioxidant properties, such as polyphenols, tocopherols, and ascorbic acid (Safafar et al. 2015). The lipid fraction of the biomass can be used to obtain a specific fatty acid (oleic, linoleic, arachidonic, etc.), or subjected to a chemical process to obtain a specific type of fuel, such as biodiesel or jet fuel (Rodolfi et al. 2009). Nevertheless, the number of processes at the production scale is small, and economic feasibility has not been proven.

There are many possible applications of microalgae and cyanobacteria, but it is necessary to develop processes from the perspective of sustainability. This has led to an increase in the proposal of processes based on the use of wastewater as a nutrient source and the complete exploitation of the biomass in the biorefinery concept (because of its similarity with oil refinery). The biorefinery processes propose to reduce the effect of the processes on the environment and reduce the generation of residues and reach a positive life-cycle assessment.

This book analyzes many examples of biotechnological applications of microalgae and cyanobacteria cultures, some of them with experimental data, and other chapters that include reviews with a general overview of innovative and promising applications.

References

- Acién FG, Molina E, Reis A, Torzillo G, Zittelli GC, Sepúlveda C, Masojídek J (2017) Photobioreactors for the production of microalgae. In: Muñoz R, Gonzalez-Fernandez C (eds) Microalgae-based biofuels and bioproducts: from feedstock cultivation to end-products. Elsevier, pp 1–44
- Barati B, Zeng K, Baeyens J, Wang S, Addy M, Gan SY, El-Fatah Abomohra A (2021) Recent progress in genetically modified microalgae for enhanced carbon dioxide sequestration. Biomass Bioenergy 145:105927
- Barsanti L, Gualtieri P (2014) Algae: anatomy, biochemistry, and biotechnology, 2nd edn. CRC Press, p 362
- Beacham TA, Sweet JB, Allen MJ (2017) Large scale cultivation of genetically modified microalgae: a new era for environmental risk assessment. Algal Res 25:90–100
- Biel K, Fomina I (2015) Benson-Bassham-Calvin cycle contribution to the organic life on our planet. Photosynthetica 53:161–167
- Borowitzka MA (2013) High-value products from microalgae-their development and commercialisation. J Appl Phycol 25:743–756
- Cañizares-Villanueva RO (2000) Biosorption of heavy metals by microorganisms. Rev Latinoam Microbiol 42:131–143
- Cañizares-Villanueva RO, Rojo-Gómez E, Arroyo-Sánchez BI, Martínez-Roldán AJ (2022) *Chlorella* as a source of antioxidant compounds. In: Marutholi M (ed) *Chlorella* and its health benefits. Nova Science Publishers
- Chini Zittelli G, Rodolfi L, Bassi N, Biondi N, Tredici MR (2013) Photobioreactors for microalgal biofuel production. In: Borowitzka MA, Moheimani NR (eds) Algae for biofuels and energy. Springer, Dordrecht, pp 115–131
- Geissen V, Mol H, Klumpp E, Umlauf G, Nadal M, van der Ploeg M, van de Zee SEATM, Ritsema CJ (2015) Emerging pollutants in the environment: a challenge for water resource management. Int Soil Water Conserv Res 3:57–65
- Jain M, Khan SA, Sharma K, Jadhao PR, Pant KK, Ziora ZM, Blaskovich MAT (2022) Current perspective of innovative strategies for bioremediation of organic pollutants from wastewater. Bioresour Technol 344:126305
- Keen OS, Bell KY, Cherchi C, Finnegan BJ, Mauter MS, Parker AM, Rosenblum JS, Stretz HA (2014) Emerging pollutants – Part II: Treatment. Water Environ Res 86:2036–2096
- Martínez-Roldán AJ, Cañizares-Villanueva RO (2015) Photobioreactors: improving the biomass productivity. In: Torres-

Bustillos L (ed) Microalgae and other phototrophic bacteria: culture, processing, recovery and new products. Nova Science Publishers, pp 145–170

- Martínez-Roldán AJ, Ibarra-Berumen J (2019) Employment of wastewater to produce microalgal biomass as a biorefinery concept. In: Alam A, Wang Z (eds) Microalgae biotechnology for development of biofuel and wastewater treatment. Springer, Singapore, pp 487–504
- Olguin E (2003) Phycoremediation: key issues for cost-effective nutrient removal processes. Biotechnol Adv 22:81–91
- Oswald WJ, Golueke CG (1960) Biological transformation of solar energy. Adv Appl Microbiol 2:223–262
- Oswald WJ, Gotaas HB (1957) Photosynthesis in sewage treatment. Trans Am Soc Civil Eng 122:73–105
- Oswald WJ, Gotaas HB, Golueke CG, Kellen WR, Gloyna EF, Hermann ER (1957) Algae in waste treatment. Sewage Ind Wastes 29:437–457
- Peña-Guzmán C, Ulloa-Sánchez S, Mora K, Helena-Bustos R, Lopez-Barrera E, Alvarez J, Rodriguez-Pinzón M (2019) Emerging pollutants in the urban water cycle in Latin America: a review of the current literature. J Environ Manag 237:408–423
- Perales-Vela HV, Peña-Castro JM, Cañizares-Villanueva RO (2006) Heavy metal detoxification in eukaryotic microalgae. Chemosphere 64:1–10
- Rodolfi L, Chini Zittelli G, Bassi N, Padovani G, Biondi N, Bonini G, Tredici MR (2009) Microalgae for oil: strain selection, induction of lipid synthesis and outdoor mass cultivation in a low-cost photobioreactor. Biotechnol Bioeng 102:100–112
- Safafar H, Van Wagenen J, Møller P, Jacobsen C (2015) Carotenoids, phenolic compounds and tocopherols contribute to the antioxidative properties of some microalgae species grown on industrial wastewater. Mar Drugs 13:7339–7356
- Solovchenko A, Khozin-Goldberg I (2013) High-CO₂ tolerance in microalgae: possible mechanisms and implications for biotechnology and bioremediation. Biotechnol Lett 35:1745–1752
- Torzillo G, Chini Zittelli G (2015) Tubular photobioreactors. In: Prokop A, Bajpai RK, Zappi ME (eds) Algal biorefineries: Volume 2: Products and refinery design. Springer, Cham, pp 187–212
- Wang B, Li Y, Wu N, Lan CQ (2008) CO₂ bio-mitigation using microalgae. Appl Microbiol Biotechnol 79:707–718
- Zhou W, Wang J, Chen P, Ji C, Kang Q, Lu B, Li K, Liu J, Ruan R (2017) Bio-mitigation of carbon dioxide using microalgal systems: advances and perspectives. Renew Sust Energ Rev 76:1163–1175



Cigdem Demirkaya and Hector De la Hoz Siegler

Abstract

Climate change is a global problem caused by the rise of carbon dioxide (CO_2) concentration in the atmosphere. To limit global warming to less than 2 °C, large-scale deployment of technologies to remove CO₂ from the air will be needed. As highly efficient, photosynthetic, single-cell factories, microalgae and cyanobacteria can play a critical role among carbon-negative technologies. Bio-capture of CO₂ using photosynthetic microbes is a viable method for recycling CO₂ into biomass, which can subsequently be utilized to produce bioenergy, fertilizers, biomaterials, and other high-value products. This chapter provides an overview of the different strategies for utilizing microalgae and cyanobacteria for CO₂ capture directly from the atmosphere or stationary point sources with minimal environmental impacts. Challenges, research needs, and opportunities for the integration of CO₂ bio-capture within a biorefinery perspective are discussed.

Keywords

 $Carbon\ capture\ \cdot\ Microalgae\ \cdot\ Cyanobacteria\ \cdot\ Photosynthesis\ \cdot\ Bioconversion\ \cdot\ Biorefinery\ \cdot\ Climate\ change$

2.1 Introduction

Due to the role of carbon dioxide (CO_2) in driving global climate change, there is an increasing global pressure to limit CO_2 emissions, particularly at large-emission source points. In 2015, with the signing of the Paris Agreement, nations committed to reduce global emissions annually by 3% to avoid a global climate catastrophe. However, this has not been achieved and the path that is being followed, which

Department of Chemical and Petroleum Engineering, University of Calgary, Calgary, AB, Canada e-mail: h.siegler@ucalgary.ca includes mainly treatment of point sources, such as flue gas, is not enough to meet the target of limiting global average temperature below a 2 °C increase. Thus, there is a rising urgency for innovative methods to mitigate new emissions and to remove the CO_2 already in the atmosphere.

Photosynthetic microbes, including microalgae, cyanobacteria, and diatoms have a great potential for mitigating and abating CO_2 emissions, while producing valuable products (Moreira and Pires 2016; Vale et al. 2020) and fostering a more sustainable bio-economy. If part of the CO_2 captured in the biomass is used to make products with relatively long life (i.e., years), or if they are permanently stored, then the cultivation of microalgae and cyanobacteria can become a key carbonnegative technology to address the climate change crisis.

Biological carbon capture is an effective and simple approach with potentially much lower energy needs compared to physical and chemical carbon capture methods. For instance, the standard technology for carbon capture in postcombustion processes is amine scrubbing, using primary or secondary amines. This technology is the basis of several megaton-scale carbon-capture projects (Feron et al. 2020). The regeneration of the amine, however, is very energyintensive, introducing a significant energy penalty and reducing the overall mitigation potential of this method (Alesi and Kitchin 2012; Stern et al. 2013).

Photosynthesis is nature's carbon capture solution. Photosynthetic organisms utilize the energy from light to drive the reaction of CO_2 and water and form biomolecules. In this way, carbon is removed from the atmosphere and stored in biomass. Gross primary production (GPP) refers to the amount of CO_2 removed from the atmosphere by photosynthesis. This is known to be one of the main fluxes controlling the carbon balance in the atmosphere and has a significant potential to offset anthropogenic carbon emissions (Beer et al. 2010). Terrestrial GPP is estimated at about 120 Pg of carbon per year (Beer et al. 2010), while marine phytoplankton are estimated to account for an additional 50 Pg of carbon per year (Yang et al. 2020). Global anthropogenic energy-

C. Demirkaya · H. De la Hoz Siegler (⊠)

[©] The Author(s), under exclusive license to Springer Nature Switzerland AG 2024

A. de Jesús Martínez-Roldán (ed.), *Biotechnological Processes for Green Energy, and High Value Bioproducts by Microalgae, and Cyanobacteria Cultures*, Developments in Applied Phycology 13, https://doi.org/10.1007/978-3-031-43969-8_2

related CO₂ emissions in 2020 were estimated at 8.6 Pg of Carbon (IEA 2021), or roughly 5% of the carbon naturally fixed by photosynthesis. Thus, it is conceivable that technological solutions based on photosynthesis will be able to offset anthropogenic carbon emissions.

Microalgae and cyanobacteria are rapidly growing microorganisms able to fix CO₂ with efficiency 10 to 50 times higher than that of terrestrial plants (Cheah et al. 2015; Raheem et al. 2018; Zhang and Liu 2021); they have high areal productivity, and high lipid and/or carbohydrate content. They are able to grow in nonarable land, with minimal nutrient inputs, and in wastewater, saline, brines, or haloalkaline waters (Moreira and Pires 2016). Thanks to their high areal productivity and ability to grow in hostile environments, photosynthetic microbes are more suitable for biological carbon capture technologies than terrestrial plants. The use of dedicated crops for industrial purposes has previously resulting in the diversion of arable lands away from traditional food crops, creating unintended impacts on food cost and supply, resulting in the wellknown food versus fuel dilemma (Darnoko and Cheryan 2000; Issariyakul and Dalai 2012). This dilemma is avoided when using photosynthetic microbes.

In addition to their role of fixing CO₂ emissions, microalgae and cyanobacteria can be used to remove nitrogen and phosphorous from agricultural and industrial effluents, reducing eutrophication of receiving water bodies (Fal et al. 2021; Guo et al. 2018; W. Zhang et al. 2020). The microbial biomass produced can be used for several applications including the production of biofuels, bioplastics, food supplements, animal feed, cosmetic additives, pharmaceutical products, and building materials (Venkata Mohan et al. 2016; Singh and Dhar 2019; Daneshvar et al. 2022). Thus, biological carbon capture with microalgae and cyanobacteria offers a wide range of opportunities for building sustainable integrated processes to support a bioresource-based circular economy (Venkata Mohan et al. 2016; Hemalatha et al. 2019; Vale et al. 2020).

This chapter presents an overview of the factors affecting the performance of carbon capture using photosynthetic microorganisms and the different strategies for utilizing microalgae and cyanobacteria for CO_2 capture directly from the atmosphere or stationary point sources with minimal environmental impacts. The challenges, research needs, and opportunities for the integration of CO_2 bio-capture from a biorefinery perspective are discussed.

2.2 Photosynthesis: Natural Carbon Capture

Photosynthesis is a natural way of capturing CO_2 and is the process responsible for transforming Earth's atmosphere from CO_2 -rich, more than 2 billion years ago when CO_2

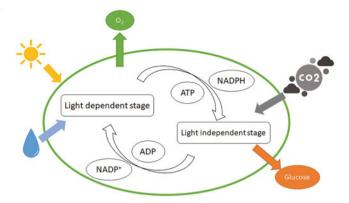


Fig. 2.1 Light-dependent and light-independent stage during photosynthesis (Adapted from Cheah et al. 2015)

atmospheric concentration was about 10 to 200 times the present level, to a relatively CO_2 -depleted one (Kaufman and Xiao 2003). Photosynthesis is carried out in two phases, see Fig. 2.1. In the first phase, light-dependent reactions capture light energy and convert it into chemical energy that is ultimately stored within nicotinamide adenine dinucleotide phosphate (NADPH) and adenosine triphosphate (ATP). The light reactions occur in the photosynthetic unit (PSU), a light-harvesting complex and reaction center located within the thylakoid membrane. The NADPH and ATP energetic molecules are then consumed in the second phase, where light-independent reactions are used to convert CO_2 into sugars (Barsanti and Gualtieri 2005; Jensen et al. 2017; Sánchez-Baracaldo and Cardona 2020).

 $2NADP + 2H_2O + 2ADP + 2P_i \rightarrow 2NADPH_2 + 2ATP + O_2 Light - dependent reaction$

$$CO_2 + 4H^+ + 4e^- \rightarrow CH_2O + H_2O$$
 Light
- independent reaction

Although photosynthesis originated in an environment with much higher CO_2 concentrations, microalgae and cyanobacteria cells have developed biological adaptations to survive under low CO_2 concentrations. The carbon concentrating mechanism (CCM) allows to increase the concentration of CO_2 within the cells relative to the normal CO_2 concentration in the air (300–400 ppm). The CCM improves photosynthetic efficiency by increasing the available CO_2 for ribulose bisphosphate carboxylase-oxygenase (RuBisCO). RuBisCO is an important enzyme that converts CO_2 into organic carbon (Gruber and Feiz 2018). Another important enzyme in the CCM is carbonic anhydrase (CA), which catalyzes the reversible conversion of CO_2 into HCO₃⁻ (DiMario et al. 2018). Several different CCMs have been identified in microalgae and cyanobacteria. In cyanobacteria, there are two types of carboxysomes (α -type and β -type), which are specialized compartments for the accumulation of HCO₃⁻. The accumulated HCO₃⁻ is then converted into CO₂ by the action of carboxysomal CA (Moroney and Ynalvez 2007). In the case of microalgae, *Chlamydomonas reinhardtii* has been studied as the model organism to understand the action of the CCM. In this microalga, the CCM can be divided into two phases. In the first phase, inorganic carbon is gathered from the environment in the form of CO₂ and HCO₃⁻ by transporter proteins. In the second phase, as the concentration of HCO₃⁻ increases in the chloroplast, HCO₃⁻ is converted into CO₂ by the action of CA (Wang et al. 2015).

Although microalgae and cyanobacteria have in general faster growth rate and higher light conversion efficiency than plants, the efficiency of large cultivation is hindered by the low CO_2 gas–liquid mass transfer rate and reduced light penetration and shading. In a fast-growing culture, the CO_2 transfer rate between the gas phase (i.e., air above culture or bubbles sparged) and the liquid medium phase media is too low to compensate for the CO_2 uptake by the cells (Zuccaro et al. 2020), resulting in carbon limitation and slower photosynthetic rate.

Light limitation also affects photosynthetic efficiency negatively (Brennan and Owende 2010). Although theoretical photosynthetic efficiency ranges between 8% and 12%, practical photosynthesis efficiency is rarely above 1.5–2%. This efficiency loss is primarily caused by light scattering and nonproductive absorption, which causes light to be exponentially attenuated as it travels along the optical path (Nwoba et al. 2019). During photosynthesis, the PSU can be in either a resting or nonactivated state or an activated state. A resting PSU is activated by the absorption of a photon. The absorption of excess photons converts functional PSUs into nonfunctional PSUs, resulting in photoinhibition (Camacho-Rubio et al. 2003). In a culture, the cells closer to the light source are more prone to experience photoinhibition as they are exposed to a higher light intensity, while the cells further down the optical path may not receive enough light. Thus, overall photosynthetic efficiency is affected by both light attenuation and scattering and photoinhibition.

2.3 CO₂ Bio-capture from Different Sources

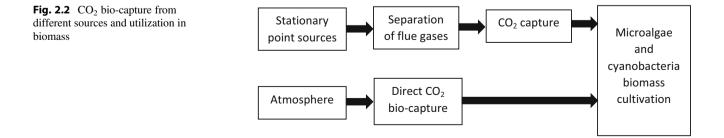
Microalgae and cyanobacteria can capture CO_2 from stationary point emission sources, such as power plants or other carbon-intensive industrial processes, or directly from the atmosphere. The CO₂ concentration in the atmosphere is 0.03-0.06% (v/v), while for stationary point sources the CO₂ concentration can vary between 6 and 15% (v/v) (Rahaman et al. 2011). The cost and energy needed for capturing CO₂ is inversely proportional to concentration, the lower the concentration of CO₂ in a given source, the more expensive the capture process. Thus, capture from large-point sources is one of the best and more efficient options to abate CO₂ emissions, as the effluent streams from combustion and industrial processes have higher CO₂ concentrations.

Figure 2.2 shows how different CO_2 sources can be integrated with microalgae and cyanobacteria biomass cultivation for CO_2 bio-capture. The following sections present an overview of technologies for CO_2 bio-capture and a discussion of the efficiency of these bio-capture methods.

2.3.1 Bio-capture of CO₂ from Stationary Point Sources

Flue or stack gases released by various stationary point sources, such as industrial complexes and power plants, have relatively high CO₂ concentrations ranging from 6 to 15% (v/v) (Thomas et al. 2016). These flue gases can be used to boost the productivity of microalgal and cyanobacteria cultures. The high concentration of CO₂ in the flue gas allows for a faster mass transfer rate, higher photosynthetic efficiency, and support a higher final cell density in the cultures.

Because of the low CO_2 solubility, the flue gas needs to be injected or bubbled directly into the cultivation medium, adding to electricity demands. The energy spent in bubbling and mixing the CO_2 in the media represents up to 27% of the overall production cost; at the same time, typically between 55 and 90% of the CO_2 injected in the culture is lost to the atmosphere (Markou et al. 2014; Caia et al. 2018). Consequently, significant research efforts have been dedicated to



improving CO_2 diffusion rates (see Sect. 2.5) and increasing CO_2 utilization efficiency.

Microalgae and cyanobacteria strains with high CO₂ uptake rate and high biomass productivity are desirable to ensure an efficiency CO₂ capture process. Sepulveda et al. (2019) assessed the ability of 11 different microalgae and cyanobacteria strains to capture CO₂ and produce high biomass productivity. They reported that Scenedesmus almeriensis and Neochloris oleoabundans were the most productive strains when used in CO₂ capture processes compared to the cyanobacteria strains. Park et al. (2021) investigated CO₂ fixation at a CO₂ concentration ranging from 5 to 40% from biogas in five pure microalgal cultures and a mixed microalgal culture, including Chlorella sp., Anabaena variabilis. Chlamvdomonas ivengarii. Chlorella vulgaris, and Chlorella sorokiniana. The highest CO₂ fixation rate was reported for *Chlorella sp.* at 1.785 g $L^{-1}d^{-1}$ at a CO₂ concentration of 15%. Additional studies on CO₂ capture and uptake by microalgae and cyanobacteria are summarized in Table 2.1.

Although the high CO_2 level in flue gas is beneficial for microalgae and cyanobacteria growth, these gases usually contain substances that can be inhibitory (Lam et al. 2012; Vale et al. 2020). In particular, unfiltered flue gas from coal combustion can have high concentration of SO_x and NO_x, microparticles, and heavy metals, such as mercury, which can present a challenge to biomass growth (Napan et al. 2015; Thomas et al. 2016). As the concentration of SO_x and NO_x increases, the acidity of the culture medium increases and this lowers the pH (Vale et al. 2020). Low pH values may inhibit microalgal growth or even result in cell death. Duarte et al. (2016) evaluated the tolerance of microalgae and cyanobacteria to the presence of NO_x and SO_x and found that strains were able to tolerate those gases at concentration of up to 400 ppm. Aslam et al. (2017) demonstrated the adaptation of mixed microalgal communities to growth in unfiltered flue gas from coal combustion. This microalgal community was dominated by Desmodesmus spp., which was the most resilient species. Radmann et al. (2011) evaluated the NO_x and SO_x tolerance of C. vulgaris,

 Table 2.1 Application of microalgae in CO₂ capture from the atmosphere and CO₂ reach sources

CO ₂ source	Microorganism	CO ₂ % (v/v)	$ \begin{array}{c} \text{CO}_2 \\ \text{fixation rate} \\ (\text{g } \text{L}^{-1} \text{d}^{-1}) \end{array} $	Culture conditions	Reference
Atmospheric CO ₂	Chlorella vulgaris and Pseudokirchneriella subcapitata	Air	0.305	OECD medium, $T = 22$ °C, different dark/light cycles at 126 µmol photons m ⁻² s ⁻¹	Pires et al. (2014)
	Dunaliella tertiolecta	0.04	0.07	Artificial sea water, $T = 26$ °C, continuous illumination at 350 µmol photons m ⁻² s ⁻¹	Hulatt and Thomas (2011)
	C. vulgaris	0.09	3.45	Artificial sea water, $T = 25$ °C, continuous illumination at ~50 µmol photons m ⁻² s ⁻¹	Fan et al. (2008)
	Anabaena sp.	0.03	1.45	Allen and Arnon medium, $T = 23$ °C, light/dark cycles with 900 µmol photons m ⁻² s ⁻¹	Ramkrishnan et al. (2014)
Enriched CO ₂ supply	Spirulina sp. DUT001	2	1.0	Zarrouk medium, $T = 25$ °C, photoperiod = 12:12, 188.7 µmol photons m ⁻² s ⁻¹	Zhu et al. (2020)
	Chlorella vulgaris	15	1.0	BBM medium, $T = 28$ °C, membrane PBR, photoperiod = 12:12, 120 μ mol photons m ⁻² s ⁻¹	Senatore et al. (2021)
	Chlorella vulgaris, Synechocystis salina, Microcystis aeruginosa,	5	0.101	OECD test medium, $T = 24$ °C, continuous illumination at 120 µmol photons m ⁻² s ⁻¹	Gonçalves et al. (2014)
	Scenedesmus obliquus	12	22.8	Soil extract medium, $T = 26$ °C, outdoor airlift PBR, 220–240 µmol photons m ⁻² s ⁻¹	Li et al. (2011)
	Chlorella vulgaris	5-25	0.27–0.47	ESP-31 medium, $T = 28$ °C, continuous illumination at 50 µmol photons m ⁻² s ⁻¹	Chou et al. (2019)
	Microalgae consortia	5.5	0.09–0.12	BBM medium, $T = 30$ °C, photoperiod = 12:12 at 1650.3 µmol photon m ⁻² s ⁻¹	Aslam et al. (2018)
	Scenedesmus almeriensis, Neochloris oleoabundans	Flue gas	2.8–2.64	Natural water from the river Seine and the artificial Seine river water, $T = 25$ °C, continuous illumination at 390 µmol photons m ⁻² s ⁻¹	Sepulveda et al. (2019)
	Spirulina sp.	2	0.81	Modified Zarrouk medium, $T = 20$ °C, pH 9, continuous illumination at 188.7 µmol photons m ⁻² s ⁻¹	Zhu et al. (2020)
	Chlorella sp	15	1.785	BG-11 medium, pH 8.2–8.7, $T = 25$ °C, photoperiod = 12:12 at 171.91 µmol photon m ⁻² s ⁻¹	Park et al. (2021)

Scenedesmus obliquus, and Synechococcus nidulans by using a simulated gas from coal combustion, containing 12% (v/v) CO₂, 100 ppm NO_x, and 60 ppm SO_x. They reported that the growth of *C. vulgaris* and *S. obliquus* was not inhibited, but this was not the case for *S. nidulans*.

In short, stationary point sources are excellent for supplying the required CO₂ concentration for carbon capture and biomass production in microalgae and cyanobacteria. However, direct use of flue gas is not, in general, possible without any separation or treatment. Identifying robust microalgae and cyanobacteria strains capable of high CO₂ bio-capture and adapted to the high concentration of other gases present in flue gas should be further explored to maximize the CO₂ bio-capture potential of microalgae and cyanobacteria culture. Furthermore, despite the large energy requirements for supplying CO_2 to the cultivation medium, a significant amount of the CO₂ provided is released into the atmosphere, decreasing net capture, and incurring inefficient energy use. Thus, additional research efforts must be directed at improving CO₂ diffusion rates and integrating different CO₂ capture techniques with microalgae and cyanobacteria cultures.

2.3.2 Biological Direct Air Capture with Microalgae and Cyanobacteria

Although capture from concentrated large-point sources of CO_2 is the most desirable and efficient option, about half of CO_2 emissions are from diffuse sources (Moreira and Pires 2016). Moreover, the CO_2 already accumulated in the atmosphere will continue to negatively contribute to climate change (Keith 2009). Thus, capture of atmospheric CO_2 using negative emissions technologies is needed to address emissions from diffuse sources and to restore the carbon balance in the atmosphere. Direct air capture (DAC) refers to technologies that directly remove CO_2 from the atmosphere. These technologies also offer the advantage of deployment in any location, independent of a specific source and without added costs for CO_2 transportation.

In the case of microalgal and cyanobacterial cultures, to capture 1 million ton of CO_2 per year, between 70 and 86 km² of the culture is needed, assuming an average productivity of 20 g m⁻² day⁻¹ of dry weight biomass and considering that about 1.6 to 2 grams of CO_2 are captured for every gram of biomass (Sayre 2010). Given the large land requirements for cultivation, it is more likely to find suitable land for deployment of large-scale cultures far away from industrial areas or population centers, where land may be scarce or expensive.

The low concentration of CO_2 in the atmosphere, however, is a major drawback as it limits CO_2 solubility and mass transfer rate into the cultivation media (Kumar et al. 2010). Carbon utilization has been shown to be more efficient when the supply rate of CO_2 matches closely with the demand of the growing biomass (Sobczuk et al. 2000; Vale et al. 2020). For DAC, active bubbling is not desirable as it requires a high energy input and will increase water evaporation. For a cost and energy-effective carbon capture process, the CO_2 supply to the cultivation needs to be improved by passive means.

To compensate for the low solubility of CO₂ in natural waters, several microalgae and cyanobacteria strains rely on the CCM to increase the intracellular concentration of bicarbonate ions and use CA to convert the HCO_3^- back to CO_2 to be used in photosynthesis. The ability of some microalgae and cyanobacteria to utilize HCO3⁻ have prompted several researchers to explore the use of alkaline culture conditions to enhance CO₂ mass transfer rate and total inorganic carbon concentration in the culture media (Chi et al. 2013; Canon-Rubio et al. 2016). Alkalinity is defined as the sum of the concentration of hydroxyl ions, bicarbonate ions, and carbonates ions, times the corresponding ion charge. As alkalinity increases, so does the concentration of dissolved inorganic carbon. High alkalinity also improves CO₂ mass transfer rate from the gas phase to the cultivation medium, as there is an increased driving force (Vadlamani et al. 2019). In addition, it provides a higher buffering capacity enabling the uncoupling of CO₂ absorption from biomass growth (Chi et al. 2013; Santos et al. 2013), as illustrated in Fig. 2.3.

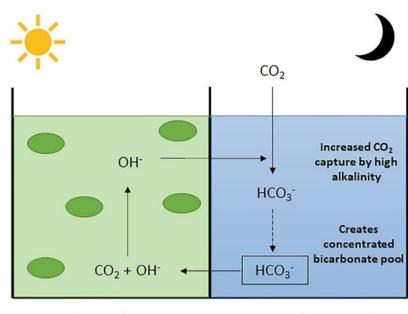
Because alkalinity can inhibit cell growth, it is necessary to operate at relatively low alkalinity or use alkali-tolerant or alkaliphilic microalgae or cyanobacteria strains. Extreme alkaline conditions together with alkaliphilic microalgae and cyanobacteria have been suggested for large-scale cultivation (Piiparinen et al. 2018; Song et al. 2019; Zhu et al. 2020). In soda lakes, at pH > 10, high concentrations of bicarbonate are present supporting a high growth rate of CO₂ fixation by photosynthetic microbes, while the consumed CO₂ is spontaneously replenished by passive diffusion from the air above the lakes (Sharp et al. 2017).

Vadlamani et al. (2017) demonstrated high biomass productivity by cultivating *C. sorokiniana* (>16 g m⁻²·d⁻¹) in a 4.2 m² raceway pond using an alkaline cultivation medium and atmospheric CO₂ alone. In another study, Zhu et al. (2020) used extreme alkaline conditions with pH ranging between 10.0 and 12.5 for DAC using *Spirulina* sp. DUT001. Effective CO₂ bio-capture was reported with maximum biomass productivity about 1.00 g L⁻¹d⁻¹ and carbon-capture rate of 0.81 g L⁻¹ d⁻¹.

2.4 Integrated Biorefinery for a Carbon-Neutral Circular Bioeconomy

To foster the development of an integrated, sustainable, and robust biological CO_2 capture process, circular economy principles must be applied to ensure the efficient processing

Fig. 2.3 Mechanism of the bicarbonate pool's role in the efficient capture of CO_2 from the air and rapid carbon supply for photosynthesis (Adapted from Zhu et al. 2020)



Photosynthesis

CO₂ capture from atmosphere

and conversion of the generated biomass, while designing out or minimizing waste, maximizing the reutilization of resources, and regenerating natural systems. The microalgal or cyanobacterial biomass produced from the CO_2 capture process consists of several biochemical compounds, including lipids, proteins, polysaccharides, and pigments. These compounds can be extracted and converted into biobased products which, in turn, displace alternative products obtained from non-sustainable sources or that have a high carbon footprint (Daneshvar et al. 2022).

An integrated biorefinery can be conceived where bio-capture of CO_2 occurs simultaneously with the production of valuable products, thus converting waste CO_2 emissions into carbon-neutral products. As microalgae or cyanobacteria require several nutrients for growth, the biorefinery concept starts with cultivation using a nutrientrich waste stream, such as wastewater, as the primary source of nitrogen, phosphorus, sulfur, and trace metals; thus, allowing the recycling and reclamation of these materials and reducing nutrient supply costs (Razzak et al. 2013; Whitton et al. 2015; Yen et al. 2015; Singh et al. 2016).

Conventional downstream processing involves harvesting and separating biomass from the cultivation media, followed by biomass pretreatment by homogenization, beading, and chemical hydrolysis to extract products of interest, and finish with the upgrading to the final products (Khoo et al. 2020). This traditional downstream processing approach is wasteful and expensive. Thus, in the biorefinery approach, the goal is to utilize the biomass to generate multiple products within a single process.

The approach in microalgal biorefineries is the cascade system (Francavilla et al. 2015; Hemalatha et al. 2019),

which allows for different biomass fractions to be extracted either simultaneously or separately by different methods (Monlau et al. 2021). Selecting the most suitable downstream processing strategy depends on the nature of the bioproducts, the required energy, and technology availability (Bastiaens et al. 2017). The use of mild separation technologies, which require low pressure, less energy, and less chemicals, is preferred for downstream to increase energy efficiency and avoid damaging the most sensitive products, which are often the most valuable. Figure 2.4 presents a possible pathway to produce multiple products from microalgal and cyanobacterial biomass.

Carbajal Tejada et al. (2020) studied five different biorefinery scenarios using *Scenedesmus dimorphus* biomass as feedstock to produce biodiesel, dihydroxyacetone, fishmeal, glycerol, and vegetable oil. Their results showed that integrated reactive distillation with the biological oxidation of glycerol to produce dihydroxyacetone was the most efficient biorefinery scenario. In another study, Moncada et al. (2014) simulated two different integrated biorefineries using *Chlorella* sp. grown with a CO₂-rich stream and sugar cane to determine the most promising scenario. The use of *Chlorella* sp. biomass to produce biodiesel, glycerol, ethanol, sugar, and electricity was found more environmentally and economically viable option than just using sugarcane alone.

Microalgal and cyanobacterial biomass is mainly composed of lipids (7-60%), proteins (6-71%), and carbohydrates (5-60%), depending upon the species and culture conditions (García-Garibay et al. 2003; Chen et al. 2013; Aziz et al. 2020). These macromolecules can be converted into several different products. The most effective use of the produced biomass will be the one that displaces

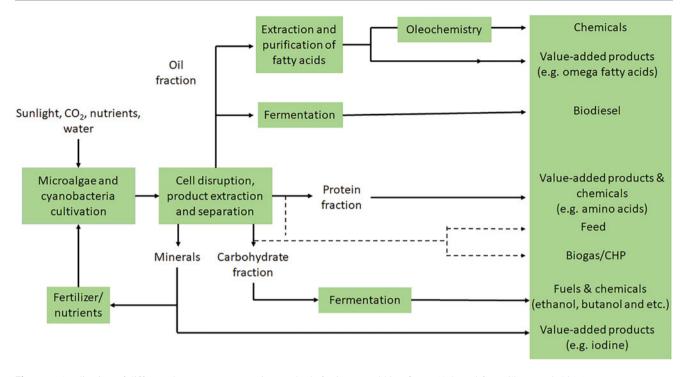


Fig. 2.4 Application of different downstream processing methods for integrated biorefinery (Adapted from Chew et al. 2017)

unsustainable feedstock or existing products with a high carbon footprint. In the following subsections, we discuss some of the products that can be more directly targeted as they either have a high carbon footprint or are high value, and therefore can help to improve the economics of the bio-capture process.

2.4.1 Biofuels

The high lipid content (15–60%) in many microalgae makes them very attractive for biodiesel production (Converti et al. 2009; Yeh and Chang 2011; Moazami et al. 2012; Polat and Altanbaş 2020). Lipid content can be further increased by manipulating several cultivation parameters (Huang and Su 2014); Polat and Altanbaş 2020). Although biodiesel production from microalgae is quite advantageous, it has not yet been commercially deployed due to the high energy-intensive downstream processing methods.

The carbohydrates in microalgal and cyanobacterial biomass are a suitable feedstock for hydrogen, bioethanol, and biogas production by fermentative pathways (Ho et al. 2013; Lakatos et al. 2019; Nagappan et al. 2019). Microalgal and cyanobacterial biomasses do not contain rigid cell wall components such as lignin, which makes them easier to process. Biodiesel production can be integrated with fermentation and anaerobic digestion to simultaneously produce a variety of energy products (Harun et al. 2011; González-González et al. 2018). After extraction of lipids for biodiesel production, the rest of the biomass, including carbohydrates, can be used as a feedstock for hydrothermal liquefaction, fermentation, or anaerobic digestion to produce biodiesel, bioethanol and/or biogas respectively. A recent technoeconomic analysis has shown that when microalgal biofuel production was integrated with other processes to obtain multiple valuable products (polyhydroxy butyrate and astaxanthin), the biofuel price was reduced to a competitive value of \$0.54/L (Rafa et al. 2021).

2.4.2 Bioactive Compounds

The main bioactive compounds produced by microalgae include polyunsaturated fatty acids (PUFAs), carotenoids, chlorophylls, phycobiliproteins, polysaccharides, and proteins.

Several microalgae produce PUFAs with known bioactive properties, such as eicosapentaenoic acid (EPA, C20:5 ω -3), docosahexaenoic acid (DHA, C22:6 ω -3), arachidonic acid (ARA, 20:6 ω -6), and γ -linolenic acid (GLA, 18:3 ω -6) (López et al. 2019). Many of these fatty acids have been studied for their anti-inflammatory activity and have been shown to prevent many diseases such as asthma, diabetes, and cardiovascular diseases (Cheng et al. 2018; Hess et al. 2018).

Some microalgal polysaccharides are considered biologically active molecules with promising applications in food, cosmetic additives, and pharmaceutical products (De Jesus Raposo et al. 2013; Barkia et al. 2019; Gouda et al. 2022). Sulfated polysaccharides are prominent for having antioxidant, anti-inflammatory, antitumoral, antiviral, antibacterial, and immunomodulatory activities (De Jesus Raposo et al. 2013). Among these, sulfated polysaccharides extracted from Porphyridium sp. and Nannochloropsis oculata have been shown to have antiviral, antitumoral, and immunostimulatory properties in pharmaceutical and therapeutic applications (Custódio et al. 2015; Casas-Arrojo et al. 2021). Apart from these benefits, polysaccharides obtained from microalgae are used as stabilizers, thickening agents, emulsifiers, and lubricants in foods, cosmetics, and textiles (Costa et al. 2021). Because some of these polysaccharides are released into the growth media during cultivation, their recovery and purification is much simpler than in the case of intracellular compounds.

Microalgae and cyanobacteria also produce essential amino acids, peptides, and proteins (Amorim et al. 2020). These valuable compounds have found multiple applications in the pharmaceutical, cosmetic, and food industries (Costa et al. 2021). *Spirulina platensis* and *C. vulgaris* are known for their high protein contents of 46–71% (Lupatini et al. 2017; Tokuşoglu and Ünal 2003). They have been used as food and feed supplements for decades, as they contain several essential amino acids, such as threonine, methionine, isoleucine, valine, leucine, lysine, and histidine (Wang et al. 2021). Montalvo et al. (2019) reported antioxidant, chelating, antimicrobial, anti-inflammatory, and anti-collagenase activity of three biopeptide fractions from *Arthrospira maxima* OF15 for potential applications in the pharmaceutical, cosmetic, and food industries.

Although all these metabolites have various applications, there are several limitations to be addressed. The main problem is the high cost of cultivation and extraction. Applying the biorefinery concept to obtain more than one product is essential to reach a cost-effective production system (Balasubramaniam et al. 2021).

2.4.3 Pigments

The three main pigment classes obtained from microalgal and cyanobacterial biomass include chlorophylls, carotenoids, and phycobilins (Koyande et al. 2019). These pigments are mainly utilized as food and feed supplements, food coloring, and as pharmaceutical and cosmetic additives because of their high antioxidant action (Begum et al. 2016; Morocho-Jácome et al. 2020). β -carotene produced by microalgae can be used as a food colorant, food and feed additive, or as precursor for vitamin A, and antioxidants (Wolf et al. 2021). Lutein and zeaxanthin are other carotenoids that are produced by microalgae and cyanobacteria and have been used as food additives due to their antioxidant activities

(Granado-Lorencio et al. 2009). Phycocyanin is a bluecolored pigment-protein complex that is extracted from cyanobacteria species such as *Arthrospira platensis* (Zeng et al. 2012). It has antioxidant, anticancer, and antiinflammatory properties and helps to improve immune function and inhibit cancer cell growth (Zeng et al. 2012). It has been used as a food ingredient and as an additional supplement to fight or prevent cancer. Furthermore, due to its naturally blue color, it has been used as a colorant for the textile and food industries to replace synthetic colorants (Rahman et al. 2017).

Pigments are one the most valuable products that can be obtained from microalgae. Although market size and product price are higher than other products, pigment extraction methods are expensive and involve the use of toxic materials. Economic feasibility and sustainability need to be improved by, e.g., investigating more environmentally friendly and non-toxic extraction methods (Rajesh et al. 2020).

2.4.4 Plastics

The global demand for plastics has increased exponentially since large-scale production of plastics started in the 1950s (Geyer et al. 2017). The carbon footprint of plastic production was estimated at 1.7 Gigaton (Gt) CO₂ equivalent in 2015 (Cabernard et al. 2021), representing 4.5% of global greenhouse gas (GHG) emissions. Because many plastics have long life spans, with some taking tens to hundreds of years to decompose, replacing fossil fuel–derived plastics with microalgal plastics is a feasible strategy for long-term carbon sequestration.

Some microalgae and cyanobacteria species produce metabolites that can be used directly for the fabrication of bioplastics, such as polyhydroxyalkanoates (PHAs) (Balaji et al. 2013), while other plastics can be obtained by chemical routes using the lipid, protein, and carbohydrate fraction of the microalgal and cyanobacterial biomass. Plastics obtained from microalgae and cyanobacteria can be designed to have properties comparable to those of fossil fuel–derived plastics (Rahman and Miller 2017).

Polyhydroxybutyrate (PHB), a type of PHA, is frequently found in cyanobacteria as an energy and carbon storage compound. Several cyanobacteria, such as *Synechocystis*, *Synochoccocus*, *Nostoc*, and *Spirulina*, are known PHB producers (Yashavanth et al. 2021), while *Synechocystis* PCC6803 has been used as a model organism to study the production of PHB (Singh et al. 2019; Koch et al. 2020). PHB is a biodegradable alternative to thermoplastics, such as polyethylene and polypropylene, and it is being commercially produced for applications in disposable food ware (McAdam et al. 2020). Recent studies have focused on the use of genetic engineering to increase PHB yield and integrate cultivation system with wastewater to reduce cultivation cost (Larkum et al. 2012; Katayama et al. 2018; López Rocha et al. 2020; Chong et al. 2021).

The triglycerides accumulated by many microalgae can be used as feedstock for the synthesis of different polyols, by chemically attaching hydroxyl groups to the unsaturated bonds in the fatty acid chains. These polyols can be converted into polyurethanes for a variety of applications by means of epoxidation and ring-opening by methanol, ethylene glycol, or lactic acid; hydroformylation; or urethane reaction with isocyanates (Hai et al. 2020; Peyrton et al. 2020). Another attractive material that can be derived from microalgae is acrylonitrile, which is a monomer widely used in the production of a variety of plastics, rubbers, resins, acrylic fibers, and polyacrylonitrile (PAN) carbon fibers (Karp et al. 2017). Glycerol obtained from the transesterification of algal oils can be converted into acrylonitrile by direct ammoxidation in the gas phase (Guerrero-Pérez and Bañares 2008).

2.4.5 Oleochemicals

Although the lipids in microalgae are thought of mainly as precursors for biofuel production, they can also be used as feedstock to produce many oleochemicals. Oleochemicals are products derived from triglycerides, including fatty acids, fatty alcohols, methyl esters, and glycerin with a wide range of applications, from food and cosmetic additives to drilling fluids and lubricants. Most oleochemicals are derived from palm, soya, canola, coconut, and palm kernel oils (Parsons et al. 2020). The sustainability of the oleochemical industry, especially palm oil, has been the source of growing public concern due to its many negative environmental impacts (Rival and Levang 2014). Although biobased, the oleochemical industry has an elevated carbon footprint and its expansion through land-use conversion has resulted in permanent damage to the biodiversity of sensitive ecosystems along with the release of massive amounts of GHG (Parsons et al. 2020). Palm-driven land-use change in Southeast Asia emits nearly 0.5 Gt of CO₂ equivalent each year, roughly 1.4% of global net GHG emissions, and is responsible for extensive ecosystem degradation. The use of algal-derived lipids to produce oleochemicals will allow phasing out unsustainable feedstock.

2.5 Challenges and Recent Progress

The main challenge for scaling up CO_2 bio-capture using microalgal and cyanobacterial cultures is the low CO_2 diffusion rates from the gas phase into the liquid culture medium, which translates into reduced CO_2 capture efficiency (Lam et al. 2012; Yen et al. 2015). To enhance the CO_2 absorption and mass transfer some approaches have been suggested, such as improving the existing photobioreactors (PBR), designing new PBR systems, and evaluating the influence of several parameters (temperature, pH, mixing, culture type, culture density, and CO_2 concentration) in the CO_2 diffusion (Morales et al. 2018).

2.5.1 PBR Design

Microalgal and cyanobacteria cultivation can be done in open ponds or closed PBRs. Open ponds are low in capital and operating costs, which is beneficial for scaling up the production; however, biomass productivity is lower than in closed systems due to high CO_2 losses, evaporation, uncontrolled climate conditions, and contamination risk (Acién et al. 2017). On the other hand, closed systems provide a bettercontrolled environment and prevent CO_2 and evaporation losses, allowing to reach higher biomass productivity (Acién et al. 2017). The existing PBRs designs for CO_2 bio-capture and biomass cultivation are vertical column reactors (bubble columns or airlift), tubular reactors, flatplate reactors, and stirrer tank reactors.

A key limiting factor in CO_2 bio-capture is the low photosynthetic conversion efficiency. Although PBRs are designed to provide a better light path than what is achieved in open ponds, light conversion efficiency is much lower than what can be theoretically achieved. Implementing new strategies to improve light penetration and delivery directly to cells minimizes energy losses and maximizes productivity. Light penetration can be increased by changing PBR orientation (horizontal, vertical, or tilted), using solar tracking devices to change the direction of the light coming to the PBR surface (Castrillo et al. 2018), optimizing light intensity and spectral distribution to prevent photoinhibition from excess light (Ooms et al. 2016), and maintaining heterogeneous light distribution to eliminate dark zones (Nwoba et al. 2019; De la Hoz Siegler 2022).

Several studies optimizing microalgae and cyanobacteria cultivation have focused on improving CO_2 gas–liquid mass transfer by improving reactor configurations to increase the contact between the gas and liquid phases. The initial bubble size is known to affect CO_2 mass transfer rate, with a smaller initial bubble size (R = 0.98 mm) resulting in increased CO_2 fixation. Hence, a small bubble is more suitable to be supplied in PBRs for the purpose of high CO_2 fixation (Barahoei et al. 2020). However, producing micro- or nanobubbles is a high energy-consuming process in which high-pressure devices are needed. Besides, high shear stress that is generated because of the bursting of small bubbles is damaging to algal cells.

Xu et al. (2020) developed a spiral-ascending CO_2 dissolver to enhance the CO_2 dissolution rate and prolong gasliquid contact time to improve microalgal growth in a horizontal tubular PBR. This cost-efficient and effective CO₂ dissolver reduced the bubble generation diameter by 23.4% and increased the CO₂ mass transfer rate by 69.2%. In another study, Gonçalves et al. (2021) designed an Oscillatory Flow Reactor with Smooth Periodic Constrictions (OFR-SPC) to improve CO₂ mass transfer without compromising fluid turbulence, which can negatively impact the most sensitive cells. This system promoted high gas–liquid mass transfer rates with low power consumption and controlled fluid turbulence. Therefore, it can be a promising technology to be used in microalgal cultivation, replacing the commonly used bubble-column and airlift PBRs.

2.5.2 Cultivation Parameters

The efficiency of the biological CO_2 capture process is also affected by CO₂ concentration, temperature, and pH. Temperature affects the solubility of CO₂ as well as the specific growth rates. CO₂ solubility decreases as temperature increases. Thus, to improve the solubility of CO_2 , the culture medium must be maintained at cooler temperatures. Each microorganism has its own optimal growth temperature, with thermophilic strains typically having higher specific growth rate than mesophilic or psychrophilic organisms. For CO₂ bio-capture from stationary point sources, thermophilic microalgal and cyanobacterial strains can be used to tolerate the high temperature of flue gas without causing a decrease in cell growth. Varshney et al. (2018) reported isolation of two novel green algal strains, Asterarcys quadricellulare and C. sorokiniana, from water bodies that were near a steel plant in India. These strains had high specific growth rates of up to 0.06 h^{-1} and 0.1 h^{-1} , respectively. Furthermore, they were able to tolerate high temperatures up to 43 °C and high concentration of CO₂ and NO_x. In addition, they reported that when they eliminated NO coming from the flue gas, strains were able to accumulate lipids up to 44% to 46% of dry biomass. The tolerance for high temperature and CO₂ concentration and their ability to accumulate lipid make these strains very attractive for CO₂ bio-capture applications.

The pH of the culture media also has a direct impact on the dissolution of CO_2 and other inorganic molecules, as it affects the chemical equilibrium between HCO_3^- and CO_3^{2-} , precipitation of phosphates, volatilization of ammonia, and the solubility of trace elements. Moreover, it directly affects cell growth due to its effect on the activity of different enzymes. For many microalgae and cyanobacteria strains, the optimum pH value is between 7 and 9. However, some organisms are known to thrive in extreme conditions. For instance, *Spirulina* grows optimally in highly alkaline conditions (pH 9–11). Alkaline conditions, at pH above

Mixing is another key cultivation parameter that affects the gas–liquid mass transfer in PBRs. Poor mixing can cause dead areas that lack nutrients and CO_2 , thus reducing overall reactor productivity. Increasing mixing rates through mechanical agitation or aeration can improve the CO_2 mass transfer, but it results in higher power consumption and introduce excessive shear stress and cellular damage.

2.5.3 Future Direction and Opportunities

Carbon capture technologies based on microalgae and cyanobacteria cultures are promising for a carbon-neutral future. However, for successful implementation of this technology, innovation in culture strategies, integration with other processes, and process optimization are needed.

Integration of other CO_2 capture processes with microalgae and cyanobacteria cultures can increase overall CO_2 capture and recovery. Chemical absorption technologies are based on the ability of different solvents, such as ionic liquids and alkaline solutions, to react with CO_2 (Vega et al. 2020). The successful integration of chemical absorption of CO_2 with *Spirulina* cultivation was demonstrated by De Rosa et al. (2015). Membrane separation uses a CO_2 permeable membrane to allow the CO_2 to pass through, while preventing other flue gasses from reaching the culture media. In this way, inhibition of the cell by toxic gasses is avoided (Cheng et al. 2021).

Recent studies have focused on the identification of highly efficient microalgae and cyanobacteria strains and on enhancing the efficiency of CO₂-fixing enzymes through genetic engineering (Barati et al. 2021). Increased CO₂ assimilation and biomass growth were reported by construction of new NADPH consumption pathways (Zhou et al. 2016), while the photosynthetic efficiency of *Nannochloropsis* sp. was improved by overexpression of RuBisCO activase (Wei et al. 2017). These achievements, however, need to be demonstrated at larger scale to ensure that the genetic constructs are stable over long cultivation periods.

2.6 Conclusion

Biological capture of CO_2 is a promising approach to mitigate CO_2 emissions and remove excess carbon from the atmosphere. Large-scale cultivation of microalgae and cyanobacteria provides environmental benefits with the possibility of producing a wide portfolio of valuable products for further a more sustainable and circular bioeconomy. Although significant progress in bio-capture of CO_2 by microalgae and cyanobacteria has been achieved, additional research efforts are needed to improve CO_2 capture efficiency.

References

- Acién FG, Molina E, Reis A, Torzillo G, Zittelli GC, Sepúlveda C, Masojídek J (2017) Photobioreactors for the production of microalgae. In: Gonzalez-Fernandez C, Muñoz R (eds) Microalgae-based biofuels and bioproducts from feedstock cultivation to end-products. Elsevier, Duxford, pp 1–44
- Alesi WR, Kitchin JR (2012) Evaluation of a primary aminefunctionalized ion-exchange resin for CO₂ capture. Ind Eng Chem Res 51(19):6907
- Amorim ML, Soares J, Sélia J, Coimbra R, De M, Leite O, Fernando L, Albino T, Martins MA, Lopes Amorim M, Elia JS, De Oliveira LM, Ar M, Martins E (2020) Microalgae proteins: production, separation, isolation, quantification, and application in food and feed. Crit Rev Food Sci Nutr 61(12):1976–2002
- Aslam A, Thomas-Hall SR, Mughal TA, Schenk PM (2017) Selection and adaptation of microalgae to growth in 100% unfiltered coal-fired flue gas. Bioresour Technol 233:271–283
- Aslam A, Thomas-Hall SR, Manzoor M, Jabeen F, Iqbal M, uz Zaman Q, Schenk PM, Asif Tahir M (2018) Mixed microalgae consortia growth under higher concentration of CO₂ from unfiltered coal fired flue gas: Fatty acid profiling and biodiesel production. J Photochem Photobiol B Biol 179:126–133
- Aziz MMA, Kassim KA, Shokravi Z, Jakarni FM, Lieu HY, Zaini N, Tan LS, Islam S, Shokravi H (2020) Two-stage cultivation strategy for simultaneous increases in growth rate and lipid content of microalgae: a review. Renew Sust Energ Rev 119:109621
- Balaji S, Gopi K, Muthuvelan B (2013) A review on production of poly β hydroxybutyrates from cyanobacteria for the production of bio plastics. Algal Res 2(3):278–285
- Balasubramaniam V, Gunasegavan RDN, Mustar S, Lee JC, Noh MFM (2021) Isolation of industrial important bioactive compounds from microalgae. Molecules 26(4):943
- Barahoei M, Hatamipour MS, Afsharzadeh S (2020) CO₂ capturing by chlorella vulgaris in a bubble column photo-bioreactor; Effect of bubble size on CO₂ removal and growth rate. J CO2 Util 37:9–19
- Barati B, Zeng K, Baeyens J, Wang S, Addy M, Gan SY, El-Fatah Abomohra A (2021) Recent progress in genetically modified microalgae for enhanced carbon dioxide sequestration. Biomass Bioenergy 145:105927
- Barkia I, Saari N, Manning SR (2019) Microalgae for high-value products towards human health and nutrition. Mar Drugs 17(5):304
- Barsanti L, Gualtieri P (2005) Algae: anatomy, biochemistry, and biotechnology, 2nd edn. CRC Press, Boca Raton, p 145
- Bastiaens L, Van Roy S, Thomassen G, Elst K (2017) Biorefinery of algae: technical and economic considerations. In: Gonzalez-Fernandez C, Muñoz R (eds) Microalgae-based biofuels bioprod. From Feed. Cultiv. to End-Products. Elsevier, Duxford, pp 327–345
- Beer C, Reichstein M, Tomelleri E, Ciais P, Jung M, Carvalhais N, Rödenbeck C, Arain MA, Baldocchi D, Bonan GB, Bondeau A, Cescatti A, Lasslop G, Lindroth A, Lomas M, Luyssaert S, Margolis H, Oleson KW, Roupsard O, Veenendall E, Viovy N, Williams C, Woodward FI, Papale D (2010) Terrestrial gross carbon dioxide uptake: global distribution and covariation with climate. Science 329(5993):834–838
- Begum H, Yusoff FMD, Banerjee S, Khatoon H, Shariff M (2016) Availability and utilization of pigments from microalgae. Crit Rev Food Sci Nutr 56(13):2209–2222

- Brennan L, Owende P (2010) Biofuels from microalgae—a review of technologies for production, processing, and extractions of biofuels and co-products. Renew Sust Energ Rev 14(2):557–577
- Cabernard L, Pfister S, Oberschelp C, Hellweg S (2021) Growing environmental footprint of plastics driven by coal combustion. Nat Sustain 5(2):139–148
- Caia M, Bernard O, Béchet Q (2018) Optimizing CO₂ transfer in algal open ponds. Algal Res 35:530–538
- Camacho-Rubio F, Chamago FG, Fernández Sevilla JM, Chisti Y, Molina Grima E (2003) A mechanistic model of photosynthesis in microalgae. Biotechnol Bioeng 81:4
- Canon-Rubio KA, Sharp CE, Bergerson J, Strous M, De la Hoz Siegler H (2016) Use of highly alkaline conditions to improve cost-effectiveness of algal biotechnology. Appl Microbiol Biotechnol 100:1611–1622
- Carbajal Tejada EM, Martínez Hernández E, Fernández Linares L, Novelo Maldonado E, Limas Ballesteros R (2020) Technoeconomic analysis of *Scenedesmus dimorphus* microalgae biorefinery scenarios for biodiesel production and glycerol valorization. Bioresour Technol Reports 12:100605
- Casas-Arrojo V, Decara J, Arrojo-Agudo MÁ, Pérez-Manríquez C, Abdala-Díaz RT (2021) Immunomodulatory, antioxidant activity and cytotoxic effect of sulfated polysaccharides from *Porphyridium cruentum*. (S.F.Gray) Nägeli. Biomolecules 11(4)
- Castrillo M, Díez-Montero R, Tejero I (2018) Model-based feasibility assessment of a deep solar photobioreactor for microalgae culturing. Algal Res 29:304–318
- Cheah WY, Show PL, Chang JS, Ling TC, Juan JC (2015) Biosequestration of atmospheric CO₂ and flue gas-containing CO₂ by microalgae. Bioresour Technol 184:190–201
- Chen CY, Zhao XQ, Yen HW, Ho SH, Cheng CL, Lee DJ, Bai FW, Chang JS (2013) Microalgae-based carbohydrates for biofuel production. Biochem Eng J 78:1–10
- Cheng F, Cui Z, Mallick K, Nirmalakhandan N, Brewer CE (2018) Hydrothermal liquefaction of high- and low-lipid algae: mass and energy balances. Bioresour Technol 258:158–167
- Cheng YW, Lim JSM, Chong CC, Lam MK, Lim JW, Tan IS, Foo HCY, Show PL, Lim S (2021) Unravelling CO₂ capture performance of microalgae cultivation and other technologies via comparative carbon balance analysis. J Environ Chem Eng 9(6):106519
- Chew KW, Yap JY, Show PL, Suan NH, Juan JC, Ling TC, Lee DJ, Chang JS (2017) Microalgae biorefinery: high value products perspectives. Bioresour Technol 229:53–62
- Chi Z, Xie Y, Elloy F, Zheng Y, Hu Y, Shulin C (2013) Bicarbonatebased integrated carbon capture and algae production system with alkalihalophilic cyanobacterium. Bioresour Technol 133:513–521
- Chong JWR, Yew GY, Khoo KS, Ho SH, Show PL (2021) Recent advances on food waste pretreatment technology via microalgae for source of polyhydroxyalkanoates. J Environ Manag 293:112782
- Chou HH, Su HY, Song XD, Chow TJ, Chen CY, Chang JS, Lee TM (2019) Isolation and characterization of *Chlorella* sp. mutants with enhanced thermo- and CO₂ tolerances for CO₂ sequestration and utilization of flue gases. Biotechnol Biofuels 12(1):1–14
- Converti A, Casazza AA, Ortiz EY, Perego P, Del Borghi M (2009) Effect of temperature and nitrogen concentration on the growth and lipid content of *Nannochloropsis oculata* and *Chlorella vulgaris* for biodiesel production. Chem Eng Process Process Intensif 48(6): 1146–1151
- Costa JAV, Lucas BF, Alvarenga AGP, Moreira JB, de Morais MG (2021) Microalgae polysaccharides: an overview of production, characterization, and potential applications. Polysaccharides 2(4): 759–772
- Custódio L, Soares F, Pereira H, Rodrigues MJ, Barreira L, Rauter AP, Alberício F, Varela J (2015) *Botryococcus braunii* and *Nannochloropsis oculata* extracts inhibit cholinesterases and protect

human dopaminergic SH-SY5Y cells from H₂O₂-induced cytotoxicity. J Appl Phycol 27(2):839–848

- Daneshvar E, Wicker RJ, Show PL, Bhatnagar A (2022) Biologicallymediated carbon capture and utilization by microalgae towards sustainable CO₂ biofixation and biomass valorization – a review. Chem Eng J 427:130884
- Darnoko D, Cheryan M (2000) Kinetics of palm oil transesterification in a batch reactor. J Am Oil Chem Soc 77(12):1263–1267
- De Jesus Raposo MF, De Morais RMSC, De Morais AMMB (2013) Bioactivity and applications of sulphated polysaccharides from marine microalgae. Mar Drugs 11(1):233
- De la Hoz Siegler H (2022) Process intensification for sustainable algal fuels production. In: El-Sheekh M, Abomohra A (eds) Handbook of algal biofuels: aspects of cultivation, conversion, and biorefinery. Elsevier, pp 503–521
- De Rosa E, Checchetto V, Franchin C, Bergantino E, Berto P, Szabò I, Giacometti GM, Arrigoni G, Costantini P (2015) [NiFe]hydrogenase is essential for cyanobacterium *Synechocystis* sp. PCC 6803 aerobic growth in the dark. Sci Reports 5(1):1–12
- DiMario RJ, Machingura MC, Waldrop GL, Moroney JV (2018) The many types of carbonic anhydrases in photosynthetic organisms. Plant Sci 268:11–17
- Duarte JH, Fanka LS, Costa JAV (2016) Utilization of simulated flue gas containing CO₂, SO₂, NO and ash for *Chlorella fusca* cultivation. Bioresour Technol 214:159–165
- Fal S, Benhima R, El Mernissi N, Kasmi Y, Smouni A, El Arroussi H (2021) Microalgae as promising source for integrated wastewater treatment and biodiesel production. Int J Phytoremediation 24(1): 34–46
- Fan LH, Zhang YT, Zhang L, Chen HL (2008) Evaluation of a membrane-sparged helical tubular photobioreactor for carbon dioxide biofixation by *Chlorella vulgaris*. J Memb Sci 325(1):336–345
- Feron P, Cousins A, Jiang K, Zhai R, Garcia M (2020) An update of the benchmark post-combustion CO₂-capture technology. Fuel 273(1): 117776
- Francavilla M, Kamaterou P, Intini S, Monteleone M, Zabaniotou A (2015) Cascading microalgae biorefinery: fast pyrolysis of *Dunaliella tertiolecta* lipid extracted-residue. Algal Res 11:184–193
- García-Garibay M, Gómez-Ruiz L, Cruz-Guerrero AE, Bárzana E (2003) Single-cell protein | algae. In: Batt C, Tortorello ML (eds) Encyclopedia of food microbiology. Elsevier, London, pp 425–430
- Geyer R, Jambeck J, Law K (2017) Production, use, and fate of all plastics ever made. Sci Adv 3(7):e1700782
- Gonçalves AL, Simões M, Pires JCM (2014) The effect of light supply on microalgal growth, CO₂ uptake and nutrient removal from wastewater. Energy Convers Manag 85:530–536
- Gonçalves AL, Almeida F, Rocha FA, Ferreira A (2021) Improving CO₂ mass transfer in microalgal cultures using an oscillatory flow reactor with smooth periodic constrictions. J Environ Chem Eng 9(6): 106505
- González-González LM, Correa DF, Ryan S, Jensen PD, Pratt S, Schenk PM (2018) Integrated biodiesel and biogas production from microalgae: towards a sustainable closed loop through nutrient recycling. Renew Sustain Energy Rev 82:1137–1148
- Gouda M, Tadda MA, Zhao Y, Farmanullah F, Chu B, Li X, He Y (2022) Microalgae bioactive carbohydrates as a novel sustainable and eco-friendly source of prebiotics: emerging health functionality and recent technologies for extraction and detection. Front Nutr 0: 391
- Granado-Lorencio F, Herrero-Barbudo C, Acién-Fernández G, Molina-Grima E, Fernández-Sevilla JM, Pérez-Sacristán B, Blanco-Navarro I (2009) In vitro bioaccesibility of lutein and zeaxanthin from the microalgae Scenedesmus almeriensis. Food Chem 114(2):747–752
- Gruber AV, Feiz L (2018) Rubisco assembly in the chloroplast. Front Mol Biosci 5:24

- Guerrero-Pérez MO, Bañares MA (2008) New reaction: conversion of glycerol into acrylonitrile. ChemSusChem 1(6):511–513
- Guo P, Zhang Y, Zhao Y (2018) Biocapture of CO2 by different microalgal-based technologies for biogas upgrading and simultaneous biogas slurry purification under various light intensities and photoperiods. Int J Environ Res Public Health 15(3)
- Hai TAP, Neelakantan N, Tessman M, Sherman SD, Griffin G, Pomeroy R, Mayfield SP, Burkart MD (2020) Flexible polyurethanes, renewable fuels, and flavorings from a microalgae oil waste stream. Green Chem 22:3088–3094
- Harun R, Davidson M, Doyle M, Gopiraj R, Danquah M, Forde G (2011) Technoeconomic analysis of an integrated microalgae photobioreactor, biodiesel and biogas production facility. Biomass Bioenergy 35(1):741–747
- Hemalatha M, Sravan JS, Min B, Venkata Mohan S (2019) Microalgaebiorefinery with cascading resource recovery design associated to dairy wastewater treatment. Bioresour Technol 284:424–429
- Hess SK, Lepetit B, Kroth PG, Mecking S (2018) Production of chemicals from microalgae lipids – status and perspectives. Eur J Lipid Sci Technol 120(1):1700152
- Ho SH, Huang SW, Chen CY, Hasunuma T, Kondo A, Chang JS (2013) Bioethanol production using carbohydrate-rich microalgae biomass as feedstock. Bioresour Technol 135:191–198
- Huang YT, Su CP (2014) High lipid content and productivity of microalgae cultivating under elevated carbon dioxide. Int J Environ Sci Technol 11(3):703–710
- Hulatt CJ, Thomas DN (2011) Productivity, carbon dioxide uptake and net energy return of microalgal bubble column photobioreactors. Bioresour Technol 102(10):5775–5787
- IEA (2021) Global Energy Review 2021, International Energy Agency, Paris, France. https://www.iea.org/reports/global-energyreview-2021
- Issariyakul T, Dalai AK (2012) Comparative kinetics of transesterification for biodiesel production from palm oil and mustard oil. Can J Chem Eng 90:342–350
- Jensen E, Clément R, Maberly SC, Gontero B (2017) Regulation of the Calvin–Benson–Bassham cycle in the enigmatic diatoms: biochemical and evolutionary variations on an original theme. Philos Trans R Soc B Biol Sci 372(1728)
- Karp EM, Eaton T, Sanchez i Nogué V, Vorotnikov V, Biddy MJ, Tan ECD, Brandner DG, Cywar RM, Liu R, Manker LP, Michener WE, Gilhespy M, Skoufa Z, Watson MJ, Fruchey OS, Vardon DR, Gill RT, Bratis AD, Beckham GT (2017) Renewable acrylonitrile production. Science 358:1307–1310
- Katayama N, Iijima H, Osanai T (2018) Production of bioplastic compounds by genetically manipulated and metabolic engineered cyanobacteria. Adv Exp Med Biol 1080:155–169
- Kaufman AJ, Xiao S (2003) High CO₂ levels in the Proterozoic atmosphere estimated from analyses of individual microfossils. Nature 425(6955):279–282
- Keith DW (2009) Why capture CO_2 from the atmosphere? Science 325(5948):1654–1655
- Khoo KS, Chew KW, Yew GY, Leong WH, Chai YH, Show PL, Chen WH (2020) Recent advances in downstream processing of microalgae lipid recovery for biofuel production. Bioresour Technol 304:122996
- Koch M, Berendzen KW, Forchhammer K (2020) On the role and production of Polyhydroxybutyrate (PHB) in the Cyanobacterium *Synechocystis* sp. PCC 6803. Life (Basel) 10(4):47
- Koyande AK, Chew KW, Rambabu K, Tao Y, Chu DT, Show PL (2019) Microalgae: a potential alternative to health supplementation for humans. Food Sci Human Wellness 8(1):16–24
- Kumar A, Ergas S, Yuan X, Sahu A, Zhang Q, Dewulf J, Malcata FX, van Langenhove H (2010) Enhanced CO₂ fixation and biofuel production via microalgae: recent developments and future directions. Trends Biotechnol 28(7):371–380

- Lakatos GE, Ranglová K, Manoel JC, Grivalský T, Kopecký J, Masojídek J (2019) Bioethanol production from microalgae polysaccharides. Folia Microbiol (Praha) 64(5):627–644
- Lam MK, Lee KT, Mohamed AR (2012) Current status and challenges on microalgae-based carbon capture. Int J Greenh Gas Control 10: 456–469
- Larkum AWD, Ross IL, Kruse O, Hankamer B (2012) Selection, breeding and engineering of microalgae for bioenergy and biofuel production. Trends Biotechnol 30(4):198–205
- Li FF, Yang ZH, Zeng R, Yang G, Chang X, Yan JB, Hou YL (2011) Microalgae capture of CO₂ from actual flue gas discharged from a combustion chamber. Ind Eng Chem Res 50:6496–6502
- López Rocha CJ, Álvarez-Castillo E, Estrada Yáñez MR, Bengoechea C, Guerrero A, Orta Ledesma MT (2020) Development of bioplastics from a microalgae consortium from wastewater. J Environ Manag 263:110353
- López G, Yate C, Ramos FA, Cala MP, Restrepo S, Baena S (2019) Production of polyunsaturated fatty acids and lipids from autotrophic, mixotrophic and heterotrophic cultivation of *Galdieria* sp. strain USBA-GBX-832. Sci Reports 9(1):1–13
- Lupatini AL, Colla LM, Canan C, Colla E (2017) Potential application of microalga Spirulina platensis as a protein source. J Sci Food Agric 97(3):724–732
- Markou G, Vandamme D, Muylaert K (2014) Microalgal and cyanobacterial cultivation: the supply of nutrients. Water Res 65: 186–202
- McAdam B, Brennan Fournet M, McDonald P, Mojicevic M (2020) Production of Polyhydroxybutyrate (PHB) and factors impacting its chemical and mechanical characteristics. Polymers 12(12):2908
- Moazami N, Ashori A, Ranjbar R, Tangestani M, Eghtesadi R, Nejad AS (2012) Large-scale biodiesel production using microalgae biomass of *Nannochloropsis*. Biomass Bioenergy 39:449–453
- Moncada J, Tamayo JA, Cardona CA (2014) Integrating first, second, and third generation biorefineries: incorporating microalgae into the sugarcane biorefinery. Chem Eng Sci 118:126–140
- Monlau F, Suarez-Alvarez S, Lallement A, Vaca-Medina G, Giacinti G, Munarriz M, Urreta I, Raynaud C, Ferrer C, Castañón S (2021) A cascade biorefinery for the valorization of microalgal biomass: biodiesel, biogas, fertilizers and high valuable compounds. Algal Res 59:102433
- Montalvo GEB, Thomaz-Soccol V, Vandenberghe LPS, Carvalho JC, Faulds CB, Bertrand E, Prado MRM, Bonatto SJR, Soccol CR (2019) Arthrospira maxima OF15 biomass cultivation at laboratory and pilot scale from sugarcane vinasse for potential biological new peptides production. Bioresour Technol 273:103–113
- Morales M, Sánchez L, Revah S (2018) The impact of environmental factors on carbon dioxide fixation by microalgae. FEMS Microbiol Lett 365(3):262
- Moreira D, Pires JCM (2016) Atmospheric CO_2 capture by algae: negative carbon dioxide emission path. Bioresour Technol 215: 371-379
- Morocho-Jácome AL, Ruscinc N, Martinez RM, de Carvalho JCM, Santos de Almeida T, Rosado C, Costa JG, Velasco MVR, Baby AR (2020) (Bio)Technological aspects of microalgae pigments for cosmetics. Appl Microbiol Biotechnol 104(22):9513–9522
- Moroney JV, Ynalvez RA (2007) Proposed carbon dioxide concentrating mechanism in *Chlamydomonas reinhardtii*. Eukaryot Cell 6(8):1251–1259
- Nagappan S, Devendran S, Tsai PC, Dahms HU, Ponnusamy VK (2019) Potential of two-stage cultivation in microalgae biofuel production. Fuel 252:339–349
- Napan K, Teng L, Quinn JC, Wood BD (2015) Impact of heavy metals from flue gas integration with microalgae production. Algal Res 8: 83–88

- Nwoba EG, Parlevliet DA, Laird DW, Alameh K, Moheimani NR (2019) Light management technologies for increasing algal photobioreactor efficiency. Algal Res 39:101433
- Ooms MD, Dinh CT, Sargent EH, Sinton D (2016) Photon management for augmented photosynthesis. Nat Commun 7(1):1–13
- Park J, Kumar G, Bakonyi P, Peter J, Nemestóthy N, Koter S, Kujawski W, Bélafi-Bakó K, Pientka Z, Muñoz R, Kim SH (2021) Comparative evaluation of CO₂ fixation of microalgae strains at various CO₂ aeration conditions. Waste Biomass Valorization 12(6):2999–3007
- Parsons S, Raikova S, Chuck CJ (2020) The viability and desirability of replacing palm oil. Nat Sustain 3(6):412–418
- Peyrton J, Chambaretaud C, Sarbu A, Avérous L (2020) Biobased polyurethane foams based on new polyol architectures from microalgae oil. ACS Sustain Chem Eng 8(32):12187–12196
- Piiparinen J, Barth D, Eriksen NT, Teir S, Spilling K, Wiebe MG (2018) Microalgal CO2 capture at extreme pH values. Algal Res 32:321– 328
- Pires JCM, Gonçalves AL, Martins FG, Alvim-Ferraz MCM, Simões M (2014) Effect of light supply on CO₂ capture from atmosphere by *Chlorella vulgaris* and *Pseudokirchneriella subcapitata*. Mitig Adapt Strateg Glob Chang 19(7):1109–1117
- Polat E, Altinbaş M (2020) Optimization of Auxenochlorella protothecoides lipid content using response surface methodology for biofuel production. Biomass Convers Biorefinery:1–15
- Radmann EM, Camerini FV, Santos TD, Costa JAV (2011) Isolation and application of SO_x and NO_x resistant microalgae in biofixation of CO₂ from thermoelectricity plants. Energy Convers Manag 52(10):3132–3136
- Rafa N, Ahmed SF, Badruddin IA, Mofijur M, Kamangar S (2021) Strategies to produce cost-effective third-generation biofuel from microalgae. Front Energy Res 9:517
- Rahaman MSA, Cheng LH, Xu XH, Zhang L, Chen HL (2011) A review of carbon dioxide capture and utilization by membrane integrated microalgal cultivation processes. Renew Sust Energ Rev 15(8):4002–4012
- Raheem A, Prinsen P, Vuppaladadiyam AK, Zhao M, Luque R (2018) A review on sustainable microalgae based biofuel and bioenergy production: Recent developments. J Clean Prod 181:42–59
- Rahman A, Miller CD (2017) Microalgae as a source of bioplastics. Algal Green Chem Recent Prog Biotechnol:121–138
- Rahman DY, Sarian FD, van Wijk A, Martinez-Garcia M, van der Maarel MJEC (2017) Thermostable phycocyanin from the red microalga Cyanidioschyzon merolae, a new natural blue food colorant. J Appl Phycol 29(3):1233–1239
- Rajesh BJ, Preethi KS, Gunasekaran M, Kumar G (2020) Microalgae based biorefinery promoting circular bioeconomy-techno economic and life-cycle analysis. Bioresour Technol 302:122822
- Ramkrishnan U, Bruno B, Swaminathan S (2014) Sequestration of CO₂ by halotolerant algae. J Environ Heal Sci Eng 12(1):1–7
- Razzak SA, Hossain MM, Lucky RA, Bassi AS, De Lasa H (2013) Integrated CO₂ capture, wastewater treatment and biofuel production by microalgae culturing—a review. Renew Sustain Energy Rev 27: 622–653
- Rival A, Levang P 2014. Palms of controversies oil palm and development challenges. Center for International Forestry Research. Bogor, Indonesia, 68p
- Sánchez-Baracaldo P, Cardona T (2020) On the origin of oxygenic photosynthesis and Cyanobacteria. New Phytol 225(4):1440–1446
- Santos A, Lamers P, Janssen M, Wijfels R (2013) Biomass and lipid productivity of Neochloris oleoabundans under alkaline–saline conditions. Algal Res 2:204–211
- Sayre R (2010) Microalgae: the potential for carbon capture. BioSci 60(9):722–727
- Senatore V, Buonerba A, Zarra T, Oliva G, Belgiorno V, Boguniewicz-Zablocka J, Naddeo V (2021) Innovative membrane photobioreactor