# ENABLING TOOLS AND TECHNIQUES FOR ORGANIC SYNTHESIS

A PRACTICAL GUIDE TO EXPERIMENTATION, AUTOMATION, AND COMPUTATION

**EDITED BY STEPHEN G. NEWMAN** 



**Enabling Tools and Techniques for Organic Synthesis** 

## **Enabling Tools and Techniques for Organic Synthesis**

A Practical Guide to Experimentation, Automation, and Computation

Edited by

Stephen G. Newman Department of Chemistry & Biomolecular Sciences University of Ottawa Ottawa, Canada



Copyright © 2023 by John Wiley & Sons, Inc. All rights reserved.

Published by John Wiley & Sons, Inc., Hoboken, New Jersey. Published simultaneously in Canada.

No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording, scanning, or otherwise, except as permitted under Section 107 or 108 of the 1976 United States Copyright Act, without either the prior written permission of the Publisher, or authorization through payment of the appropriate per-copy fee to the Copyright Clearance Center, Inc., 222 Rosewood Drive, Danvers, MA 01923, (978) 750-8400, fax (978) 750-4470, or on the web at www.copyright. com. Requests to the Publisher for permission should be addressed to the Permissions Department, John Wiley & Sons, Inc., 111 River Street, Hoboken, NJ 07030, (201) 748-6011, fax (201) 748-6008, or online at http://www.wiley.com/go/permission.

Trademarks: Wiley and the Wiley logo are trademarks or registered trademarks of John Wiley & Sons, Inc. and/or its affiliates in the United States and other countries and may not be used without written permission. All other trademarks are the property of their respective owners. John Wiley & Sons, Inc. is not associated with any product or vendor mentioned in this book.

Limit of Liability/Disclaimer of Warranty: While the publisher and author have used their best efforts in preparing this book, they make no representations or warranties with respect to the accuracy or completeness of the contents of this book and specifically disclaim any implied warranties of merchantability or fitness for a particular purpose. No warranty may be created or extended by sales representatives or written sales materials. The advice and strategies contained herein may not be suitable for your situation. You should consult with a professional where appropriate. Further, readers should be aware that websites listed in this work may have changed or disappeared between when this work was written and when it is read. Neither the publisher nor authors shall be liable for any loss of profit or any other commercial damages, including but not limited to special, incidental, consequential, or other damages.

For general information on our other products and services or for technical support, please contact our Customer Care Department within the United States at (800) 762-2974, outside the United States at (317) 572-3993 or fax (317) 572-4002.

Wiley also publishes its books in a variety of electronic formats. Some content that appears in print may not be available in electronic formats. For more information about Wiley products, visit our web site at www.wiley.com.

#### Library of Congress Cataloging-in-Publication Data Applied for

ISBN: 9781119855637, ePDF: 9781119855651, epub:9781119855644, oBook:9781119855668

Cover Design: Wiley

Cover Image: © Sergey Tarasov/Adobe Stock Photos

Set in 9.5/12.5pt STIXTwoText by Straive, Pondicherry, India

#### **Contents**

**List of Contributors** xv

	Preface xix
1	Biocatalysis 101 – A Chemist's Guide to Starting Biocatalysis 1 Pablo Díaz-Kruik, David Lim, and Francesca Paradisi Glossary 1
1.1	Introduction 1
1.1.1	Enzymes – the Green and Sustainable Way of the Future 1
1.1.2	Enzymatic and Organic Catalysis Are Not too Different from Each Other 3
1.1.3	Enzymes 101 4
1.2	When Should I Choose an Enzyme over a Chemical Catalyst? 4
1.3	Key Considerations for Running Biocatalytic Reactions 6
1.3.1	Dispelling Myths 6
1.3.1.1	Enzymes Are Not Safe to Use 7
1.3.1.2	Enzymes Are Not as Readily Available as Chemical Catalysts 7
1.3.1.3	Enzymes Are Seldom Useful Due to Their Limited Substrate Scope 7
1.3.1.4	The Cost of Enzyme Production Is Very High 8
1.3.1.5	Enzymes Are Functionally Unstable Under Organic Conditions 9
1.3.1.6	Sustainability 9
1.3.2	Challenges of Using Enzymes: the Need for Strict Reaction Conditions 9
1.3.2.1	Enzymes from Extremophiles 10
1.3.2.2	Solvents (and Co-solvents) 10
1.3.2.3	Concentration and Ionic Strength of the Buffer 10
1.3.2.4	pH Dependence 11
1.3.2.5	Concentration of Reactants 11
1.3.2.6	Enzyme Concentration 12
1.3.2.7	Enzyme Forms 12
1.3.2.8	Toxicity 13
1.3.3	What Do I Need to Start Biocatalytic Experiments in My Lab? 13
1 3 /	Additional Considerations 14

Transformations Catalyzed by Enzymes

EC – The Enzyme Commission Number 15

1.4

1.4.1

1.4.1.1	EC 1 – Oxoreductases 15
1.4.1.2	EC 2 – Transferases 16
1.4.1.3	EC 3 – Hydrolases 16
1.4.1.4	EC 4 – Lyases 17
1.4.1.5	EC 5 – Isomerases 18
1.4.1.6	EC 6 – Ligases 18
1.4.1.7	EC 7 – Translocases 18
1.4.2	Some Applications of Selected Commercially Available Enzymes 19
1.4.2.1	Horseradish Peroxidase 19
1.4.2.2	Lysozyme 19
1.4.2.3	Trypsin 20
1.4.2.4	Candida Lipase B 20
1.4.2.5	Amino Acid Dehydrogenase 20
1.4.2.6	Glycosidases 21
1.4.3	Engineered (Unnatural) Reactions 21
1.5	New Trends and Technologies in Biocatalysis 21
1.5.1	Flow Biocatalysis and New Technologies 21
1.5.1.1	What Is Flow Biocatalysis? 21
1.5.1.2	How Does Flow Biocatalysis Work? 21
1.5.1.3	When Is a Flow Process More Beneficial for a Specific Transformation? 23
1.5.1.4	Should One Implement Every Enzymatic Reaction in Flow? 23
1.5.2	Enzyme Engineering 24
1.5.3	Photobiocatalysis 25
1.6	Flow Chart to Biocatalysis 25
1.7	Case Study: Setting up a Biotransformation 27
1.8	Concluding Remarks 31
	Additional Resources 31
	References 31
2	Introduction to Photochemistry for the Synthetic Chemist 37
	Stefano Protti, Davide Ravelli, and Maurizio Fagnoni
	Glossary 37
2.1	Introduction 38
2.1.1	Light to Make Your Synthesis Greener 38
2.1.2	A Way to Overcome HOMO/LUMO Interactions 39
2.2	How to Plan a Photochemical Synthesis 45
2.2.1	The Choice of the Solvent 45
2.2.2	Concentration of the Absorbing Species 47
2.2.3	The Reaction Vessel 48
2.2.4	Light Sources 48
2.2.4.1	Low-Pressure Mercury Arcs 49
2.2.4.2	Medium- and High-Pressure Mercury Arcs 50
2.2.4.3	Other Light Sources 50
2.2.5	From Batch to Flow Conditions 52

2.2.6	Preparation of the Sample 54
2.2.7	Safety Equipment 54
2.3	Selected Applications of Photochemical/Photocatalyzed Reactions 55
2.3.1	Reactions Involving the C=C Double Bond 55
2.3.2	Reactions Involving the C=O Double Bond 58
2.3.3	Reactions Involving a Photoinduced Homolysis 60
2.3.4	Reactions Involving Singlet Oxygen 62
2.3.5	Reactions Involving a Photocatalytic Step 62
2.4	Conclusions 67
	Acknowledgment 67
	References 67
3	How to Confidently Become an Electrosynthetic Practitioner 73
	Sylvain Charvet, Taline Kerackian, Camille Z. Rubel, and Julien C. Vantourout
	Glossary 73
	Abbreviations 76
3.1	Introduction 77
3.2	General Definition of Organic Electrosynthesis 78
3.3	Why is Organic Electrosynthesis Used? 78
3.4	How is Organic Electrosynthesis Performed? 78
3.5	Where to Start with Electrosynthesis? 79
	Selected General Reviews 79
	Selected General Guides 79
3.6	Electrasyn 2.0 80
3.6.1	Machine and Consumables 80
3.6.1.1	Opening the IKA ElectraSyn 2.0 Box 80
3.6.1.2	Cell (Vial and Cap) 81
3.6.1.3	Electrodes 82
3.6.2	Interface 83
3.6.2.1	Hardware 83
3.6.2.2	Menus 84
3.6.3	How to Set Up the Cell 84
3.6.4	How to Start an Experiment 85
3.6.5	During the Reaction 88
3.6.6	After the Reaction 89
3.7	Case Study 90
3.7.1	Project Overview 90
3.7.2	Optimization of Parameters 92
3.7.2.1	Designing an Electrochemical Experiment 92
3.7.3	Proof of Concept 94
3.7.3.1	Optimization 94
3.7.3.2	Substrate Scope 102
3.8	Conclusion 103

References 103

4	Flow Chemistry 107
	Yosuke Ashikari and Aiichiro Nagaki
	Glossary 107
4.1	Introduction 109
4.1.1	What is Flow Microchemistry 109
4.1.1.1	Reaction Time Controllability 110
4.1.1.2	Fast Mixing 111
4.1.1.3	Temperature Controllability 112
4.1.2	Reactions Enabled by Flow Microreactors 112
4.1.2.1	Competitive Sequential Reactions 112
4.1.2.2	Reactions Mediated by Unstable Intermediates 114
4.1.2.3	Reactions Occurring at the Surface: Two-Phase Reactions,
	Electrochemical Reactions, and Photoreactions 117
4.1.3	Further Applicability of Flow Microsynthesis 118
4.1.3.1	Scalability 118
4.1.3.2	Safety Operation 118
4.2	General Information for Flow Microreactors 118
4.2.1	Tools and Equipment for Flow Chemistry 119
4.2.1.1	Micromixer 119
4.2.1.2	Tube Reactor 120
4.2.1.3	Pump 120
4.2.1.4	Pre-Cooling Tubes 121
4.2.1.5	PTFE Tubes 121
4.2.2	How to Perform Experiments 122
4.2.2.1	Selection of Reaction Conditions 122
4.2.2.2	Preparation of Reagent Solution 125
4.2.2.3	Preparation for Reactions 126
4.2.2.4	Preparation for Reaction Evaluation 128
4.2.2.5	Cleaning Up 129
4.3	Case Studies 129
4.3.1	Competitive Sequential Reaction (General Procedure) 129
4.3.1.1	Preparation 130
4.3.1.2	Experiment 132
4.3.1.3	Screening of Reaction Conditions 133
4.3.1.4	Analysis 134
4.3.1.5	Clean Up 136
4.3.2	Reactions Mediated by Short-Lived Intermediates 136
4.3.3	Reaction Integration 139
4.4	Further Expertise 142
4.4.1	Reaction Integration 142
4.4.2	Chemoselective Reactions 143
4.4.3	Heterogeneous Catalytic Reactions 143
4.5	Summary and Outlook 144
	References 144

5	<b>Reaction Optimization Using Design of Experiments</b> 149 Laura Forfar and Paul Murray
	Glossary 149
<i>5</i> 1	•
5.1	Introduction 151
5.1.1	How Do We Experiment and DoE Terminology 151
	OVAT vs. DoE 153
	A Simple Chemical Example 153
5.1.3	A Note on Error, Accuracy, and Precision 156
5.2	When and How Can DoE Be Used? 157
5.3	What Information Can I Get from a DoE and How Is It Obtained? 158
5.3.1	Which Factors Are Important? 159
5.3.2	How Are the Models Generated? 161
5.4	What Types of Design Are Available? 164
5.4.1	Screening Designs 164
5.4.1.1	Fractional Factorial Designs 165
5.4.1.2	Definitive Screening Designs 166
5.4.2	Designs for Optimizing Reactions 167
5.4.3	Response Surface Designs 167
5.5	The DoE Process 169
5.5.1	Aim and Objective 170
5.5.2	Selecting Factors and Ranges 171
5.5.2.1	Factors 171
5.5.2.2	
5.5.3	Selecting Responses 175
5.5.4	• •
5.5.5	
5.5.6	
	Model Data 179
	General Steps for Developing a Model 180
	Wittig Reaction 181
5.5.7.3	
5.5.8	Validate Predictions 189
5.6	Combining DoE with Other Screening and Optimization
5.0	Techniques 191
<i>-</i> 7	•
5.7	Software 192 "I Tried Formaria and Decim But It Pid Not World." 103
5.8	"I Tried Experimental Design But It Did Not Work" 193
5.9	Conclusion 194
	References 195
6	Introduction to High-Throughput Experimentation (HTE) for the
-	Synthetic Chemist 197
	Stephanie Felten, Michael Shevlin, and Marion H. Emmert
	Glossary 197
6.1	What Is HTE? 199

6.2	Why HTE and What Can It Achieve? 199
6.2.1	Commonly Perceived Barriers to Employing HTE in Synthetic
0.2.1	Chemistry 200
6.2.1.1	Cost 200
6.2.1.2	Availability of Dedicated HTE Facilities 200
6.2.1.3	Access to Knowledge and Training 201
6.2.1.4	Perception of HTE as Antithesis of Hypothesis-driven Research 201
6.2.2	
6.2.2.1	Advantages of HTE Workflows vs. Traditional Reaction Setup 203 Setup Time per Reaction 203
6.2.2.2	Miniaturization and Efficient Reagent Use 203
6.2.2.3	Multivariable vs. Sequential Optimization 203
6.2.2.4	Visualizing Reactivity Patterns 204
6.2.2.5	Serendipity in Reaction Discovery 204
	Avoiding Cross-contamination 206
6.2.2.6	
6.3 6.3.1	Practical Considerations and Tools for HTE 206 Outline of a Typical HTE Workflow 207
	• •
6.3.2	Types of HTE Designs 209
6.3.2.1	HTE for Reaction Discovery 209
6.3.2.2	HTE for Reaction Optimization 210
6.3.3	HTE Design Software: Tools for Building Arrays 211
6.3.4	HTE Reactors and Consumables 214
6.3.4.1	Reaction Blocks 214
	HTE Vials 214
6.3.4.3	Reaction Blocks with Sealing Top Plate 215
6.3.4.4	Special Reactors for Photochemistry, Electrochemistry, and
6245	High-Pressure Reactions 215
6.3.4.5	Reaction Stirring and Temperature Control 217
6.3.4.6	Consumables 219
6.3.5	Considerations for Experimental Setup 220
6.3.5.1	Reaction Atmosphere 220
6.3.5.2	Reagent Preparation and Dispensing 221
6.3.5.3	Storage of Preplated Reagents 223
6.3.5.4	Pipetting 224
6.3.5.5	Solvent Evaporation 225
6.3.6	Analysis of HTE Screens 226
6.3.6.1	
6.3.6.2	Autosampler Configurations 226
6.3.6.3	Analytical Methods 227
6.3.6.4	Internal Standards and Assay Yields 227
6.3.6.5	Data Visualization and Analysis 228 The Role of Automation and Robotics in HTE 229
6.3.7	
6.4	Section Summary and Outlook 232 Case Study 1: Development of an HTE Platform for Nickel-Catalyzed
6.5	- Case Singy i: Development of an Hite Planorm for Nickel-Cafalyzed

Suzuki–Miyaura Reactions 233

6.5.1	Motivation 233
6.5.2	Design of Test Reaction and Initial Ligand Screen 233
6.5.3	Second Round of Ligand/Base/Solvent Screens 235
6.5.4	Final Platform Design 237
6.5.5	Validation of Platform Design 237
6.6	Case Study 2: HTE Enabled Reaction Discovery and Optimization of
	Silyl-Triflate-Mediated C-H Aminoalkylation of Azoles 240
6.6.1	Motivation 240
6.6.2	Reaction Discovery Plate Design 240
6.6.3	Ligand Screen 243
6.6.4	Parallel Optimization of Three Reagents 244
6.6.5	Base Screen 244
6.7	Current Challenges and the Future of HTE 247
6.7.1	Summary and Conclusions 247
6.7.2	Remaining Challenges: The Next Frontiers 248
6.7.2.1	Biphasic Reaction Mixtures 248
6.7.2.2	Flow Chemistry and HTE 248
6.7.2.3	Reaction Profiling 249
6.7.2.4	Building Machine Learning Models to Predict Reactivity 249
6.7.2.5	Addressing Future Challenges 250
	Acknowledgments 250
	Further Recommended Reading 250
	References 250
7	Concepts and Practical Aspects of Computational Chemistry 259
	Martin Breugst
	Glossary 259
7.1	Introduction 261
7.2	Hardware and Software Requirements for Computational
	Investigations 264
7.3	Typical Methods in Computational Organic Chemistry 265
7.3.1	General Aspects 265
7.3.2	Molecular Mechanics and Force Fields 266
7.3.3	Wave-Function Methods I – Hartree–Fock Theory 267
7.3.4	Wave-Function Methods II – Post-Hartree–Fock Theory 267
7.3.5	Semiempirical Methods 269
7.3.6	Density Functional Theory 269
7.3.7	Dispersion-Corrected Density Functional Theory 271
7.3.8	Typical Computational Times 272
7.4	Basis Sets Used in Computational Organic Chemistry 273
7.4.1	General Aspects of Basis Sets 273
7.4.2	Introduction to the Mathematical Formalism in Basis Sets 274
7.4.3	Polarization and Diffuse Functions 275
7.4.4	Basis Set Families 276

7.4.5	Effective Core Potentials (Pseudopotentials) 278
7.4.6	The Basis Set Superposition Error (BSSE) 279
7.5	Typical Computational Tasks in Organic Chemistry 279
7.5.1	Preliminary Remarks 279
7.5.2	Single-Point Calculations 281
7.5.3	Geometry Optimizations 281
7.5.4	Frequency Calculations 282
7.5.5	Intrinsic Reaction Coordinate (IRC) Calculations 284
7.5.6	Conformational Analysis 285
7.6	Notation of the Model Chemistry 286
7.7	The Diels–Alder Reaction as a Tutorial Case Study 286
7.7.1	General Aspects and Requirements 286
7.7.2	Preparing Input Files 288
7.7.3	Conformational Sampling – Generation of Initial Geometries 290
7.7.4	Geometry Optimizations of Starting Materials and Products 291
7.7.5	Locating the Transition States 294
7.7.6	Verifying the Nature of the Transition State 298
7.8	More Advanced Aspects 300
7.8.1	General Comments 300
7.8.2	Influence of Solvation 300
7.8.3	Integration Grid 302
7.8.4	Standard States 302
7.8.5	Treating Unpaired Electrons 303
7.9	Important and Frequently Used Keywords 304
7.10	Practical Considerations 304
7.11	Conclusions 306
	References 306
8	NMR Prediction with Computational Chemistry 313
	Amy T. Merrill, Wentao Guo, and Dean J. Tantillo
	Glossary 313
8.1	Introduction 314
8.2	Quantum-Chemistry-Based Computational NMR 315
8.2.1	Methods 315
8.2.1.1	Time/Resources for Calculations 316
8.2.1.2	Structural Considerations in Modeling 317
8.2.1.3	Geometry Optimizations 323
8.2.1.4	Calculating Isotropic Shielding Constants 324
8.2.1.5	Common Pitfalls and How to Address Them 328
8.2.1.6	Converting to Chemical Shifts 329
8.2.1.7	Calculating Coupling Constants 330
8.2.2	Confidence Analysis 330
8.2.3	Computer-Aided Automated Approaches 332

8.2.3.1	CASE 332
8.2.4	A Case Study 336
8.2.5	Practicing <sup>1</sup> H and <sup>13</sup> C Chemical Shift Prediction 338
8.3	Summary and Outlook 339
	Key References 339
	References 340
9	Introduction to Programming for the Organic Chemist 347
	lason M. Stevens
9.1	Introduction 347
9.2	Better Visualizations: Communicating Structure–Data
	Relationships 351
9.3	Text Extraction: Automating Density Functional Theory
	Calculations 354
9.4	Statistical Analysis: Deriving Insight from Historical Data 357
9.5	Machine Learning: A Predictive Model for Deoxyfluorination 359
9.6	Working with Public Datasets: Identifying Reactivity Cliffs 364
9.7	Running Simulations: Process Greenness 367
9.8	Application Development: Process Mass Intensity Predictor 371
9.9	Machine Learning for Reaction Optimization 374
9.10	Executing Robotic Tasks 378
9.11	Autonomous Reaction Optimization 381
9.12	Conclusion 384
	References 385
10	Machine Learning for the Optimization of Chemical Reaction
10	Conditions 393
	A. Filipa de Almeida and Tiago Rodrigues
	Glossary 393
10.1	Introduction 394
10.1	Prior Art and Alternative Methods for Rational Reaction
10.2	Optimization 396
10.3	Reaction Optimization Using LabMate.ML 400
10.3.1	Step One: Accessing the LabMate.ML Code and Installation 401
10.3.2	Step Two: Initializing the Optimization Routine in LabMate.ML 402
10.3.3	Step Three: Iterative Optimization Routine 404
10.3.4	Examples 406
10.4	Primer on Evaluation Guidelines 408
10.4.1	Code and Dataset Availability 408
10.4.2	Retrospective Evaluation 409
10.4.3	Baselines and Comparing Tools 410
10.4.4	Prospective Evaluation 412

xiv	Contents
xiv	Contents

10.5	Outlook 414			
	References 416			
11	Computer-Assisted Synthesis Planning 423			
	Zhengkai Tu, Itai Levin, and Connor W. Coley			
	Glossary 423			
11.1	Introduction to Computer-Aided Synthesis Planning 424			
11.1.1	Defining the Tasks and Use Cases 424			
11.1.2	Historical Approaches to Computer-Aided Synthesis Planning 425			
11.1.3	The Inflection Point of CASP Methods 425			
11.1.4	Preliminaries on Molecular Representation and Cheminformatics 426			
11.1.5	Outline of the Rest of the Chapter 428			
11.2	Approaches and Algorithms for Retrosynthesis 428			
11.2.1	Data-driven v. Expert-Driven Programs 428			
11.2.2	Template-Based Approaches 429			
11.2.3	Template-free Approaches with Graphs and Sequences 431			
11.2.4	Multistep Planning Algorithms 433			
11.3	Approaches and Algorithms for Condition Recommendation			
	and Forward Synthesis 436			
11.3.1	Condition Recommendation Approaches 436			
11.3.2	Forward Synthesis Approaches 437			
11.4	Select Examples of Software Tools for CASP 439			
11.4.1	Open-Source Tools 439			
11.4.1.1	ASKCOS 439			
11.4.1.2	AiZynthFinder 440			
11.4.1.3	Retro* 442			
11.4.2	Closed-Source Tools 443			
11.4.3	CASP Tools for Enzymatic Catalysis 446			
11.4.4	Practical Considerations for CASP Programs 446			
11.4.4.1	Traceability to Literature Precedent 447			
11.4.4.2	How to Use CASP: Command Line Versus Graphical User Interface 447			
11.4.4.3	Data Privacy 448			
11.4.4.4	Customization Ability 448			
11.5	Case Studies 448			
11.5.1	Segler et al.'s Data-driven Program and A/B Testing Success 449			
11.5.2	MIT's ASKCOS Program and Robotic Synthesis			
	Demonstration 449			
11.5.3	Grzybowski's Chematica/Synthia Program's Experimental Validations			
11.6	and Acquisition 450			
11.6	Conclusion 451			
	Key References 453			
	References 453			

#### **List of Contributors**

#### Yosuke Ashikari

Department of Chemistry Faculty of Science Hokkaido University Sapporo, Japan

#### Martin Breugst

Institut für Chemie Technische Universität Chemnitz Chemnitz, Germany

#### Sylvain Charvet

Institut de Chimie et Biochimie Moléculaires et Supramoléculaires (ICMBS, UMR 5246 du CNRS) Université Lyon Villeurbanne, France

#### Connor W. Coley

Department of Chemical Engineering Massachusetts Institute of Technology Cambridge, MA, USA

and

Department of Electrical Engineering and Computer Science Massachusetts Institute of Technology Cambridge, MA, USA

#### Pablo Díaz-Kruik

Department of Chemistry Biochemistry and Pharmaceutical Sciences Bern, Switzerland

#### Marion H. Emmert

Process Research & Development Merck & Co., Inc. Rahway, NJ, USA

#### Maurizio Fagnoni

PhotoGreen Laboratory Department of Chemistry University of Pavia Pavia, Italy

#### Stephanie Felten

Process Research & Development Merck & Co., Inc. Rahway, NJ, USA

#### A. Filipa de Almeida

Instituto de Investigação do Medicamento (iMed) Faculdade de Farmácia Universidade de Lisboa Lisbon, Portugal

#### Laura Forfar

Paul Murray Catalysis Consulting Ltd Yate, Bristol, UK

#### Wentao Guo

Department of Chemistry University of California Davis, CA, USA

#### Taline Kerackian

Institut de Chimie et Biochimie Moléculaires et Supramoléculaires (ICMBS, UMR 5246 du CNRS) Université Lyon Villeurbanne, France

#### Itai Levin

Synthetic Biology Center Department of Biological Engineering Massachusetts Institute of Technology Cambridge, MA, USA

and

Department of Chemical Engineering Massachusetts Institute of Technology Cambridge, MA, USA

#### David Lim

Department of Chemistry Biochemistry and Pharmaceutical Sciences Bern, Switzerland

#### Amy T. Merrill

Department of Chemistry University of California Davis, CA, USA

#### **Paul Murray**

Paul Murray Catalysis Consulting Ltd Yate, Bristol, UK

#### Aiichiro Nagaki

Department of Chemistry Faculty of Science Hokkaido University Sapporo, Japan

#### Francesca Paradisi

Department of Chemistry Biochemistry and Pharmaceutical Sciences Bern, Switzerland

#### Stefano Protti

PhotoGreen Laboratory Department of Chemistry University of Pavia Pavia, Italy

#### Davide Ravelli

PhotoGreen Laboratory Department of Chemistry University of Pavia Pavia, Italy

#### Tiago Rodrigues

Instituto de Investigação do Medicamento (iMed) Faculdade de Farmácia Universidade de Lisboa Lisbon, Portugal

#### Camille Z. Rubel

Institut de Chimie et Biochimie Moléculaires et Supramoléculaires (ICMBS, UMR 5246 du CNRS) Université Lyon Villeurbanne, France

and

Department of Chemistry The Scripps Research Institute La Jolla, CA, USA

#### Michael Shevlin

Process Research & Development Merck & Co., Inc. Rahway, NJ, USA

#### Jason M. Stevens

Bristol Myers Squibb Summit, NJ, USA

#### Dean J. Tantillo

Department of Chemistry University of California Davis, CA, USA

#### Zhengkai Tu

Computational Science and Engineering Massachusetts Institute of Technology Cambridge, MA, USA

#### Julien C. Vantourout

Institut de Chimie et Biochimie Moléculaires et Supramoléculaires (ICMBS, UMR 5246 du CNRS) Université Lyon Villeurbanne, France

#### **Preface**

At the undergraduate level, the organic chemistry curriculum at most universities is similar. Professors emphasize the fundamental concepts necessary to understand how, when, and why organic molecules interact, while lab instructors familiarize students with important hands-on aspects of carrying out experiments. Bachelor's students can expect to finish their studies with an idea of how molecules behave, how they are made, and how technologies such as NMR and IR can be used for their characterization. Those that enter graduate school are often surprised at the breadth of powerful technologies that make advanced organic chemistry the discipline it is today. Instead of a typical stirred round-bottomed flask, many reactions are better done using photochemical, electrochemical, or flow reactors. Computational chemistry, once reserved for dedicated experts, can now be used by organic chemists to help predict outcomes, understand selectivity, and decipher reaction mechanisms. Automation technology can be used to generate large amounts of data with limited amounts of material, and data processing software can be used to extract subtle trends.

Due to their prominence in the recent literature, trainees and established chemists alike would benefit from gaining expertise with these technologies to be best prepared for solving the diverse synthetic challenges that come their way. However, the barrier to learning techniques without formal instruction can be high. Even if one is fortunate enough to have access to advanced training, the expert instructor may not necessarily curate the course to the needs and the background of a synthetic chemist. The primary literature and recent textbooks have similar limitations – while there is no shortage of resources, experts generally write to other experts, and the interested organic chemist may have critical gaps in their understanding and struggle with subdiscipline-specific jargon.

The goal of this text is to help fill this gap by providing synthetic chemists with a user-friendly starting point to initiate their journey in developing new skills and knowledge. In each of the 11 chapters, experts communicate basic information about an impactful technology in a manner accessible to a classically trained

synthetic chemist. Chapters also includes a glossary of common terminology, a general introduction to the technology of interest, case-study examples of how it may useful to synthetic chemists, a practical discussion about steps one may take to put knowledge into practice, and references to recommended further reading. The book seeks to be a go-to resource for organic chemists at or above the graduate level that wish to expand the breadth of tools they can use to perform, analyze, and interpret chemistry experiments. After completion, the reader will be armed with the practical knowledge needed to comprehend the literature, to assess the strengths and limitations of each technique, and to begin applying modern tools to solve synthetic challenges. This will make it useful as a general resource for graduate students looking to expand their expertise, for instructors of graduatelevel courses on advanced techniques for organic synthesis, and for industrial scientists seeking a beginner-friendly way to expand their knowledge.

The book is organized into four subsections. Chapters 1-4 describe different enabling technologies for performing chemical experiments - biocatalysis, photochemistry, electrochemistry, and flow chemistry. While none of these topics are fundamentally new, their power as a tool for organic synthesis is becoming increasingly evident. These chapters will help the reader overcome the technical barrier hindering them from comfortably replicating experiments and designing their own. Chapters 5 and 6 focus on improved approaches to select, carry out, and analyze experiments. Specifically, Chapter 5 describes a statistical approach to experimentation that can be used to understand and optimize chemical reactions. This Design of Experiments (DoE) technique is commonly employed by practicing scientists in many fields but is seldom taught to chemists. Chapter 6 describes techniques that researchers can use to get more data using less time and fewer resources. This high-throughput experimentation (HTE) approach shows the reader how to carry out reactions in parallel and how the collected data can be interpreted to gain insights that might otherwise be missed. Chapters 7 and 8 introduce the reader to computational chemistry tools that enable molecules and reactions to be modeled in silico, providing predictions and mechanistic insight to supplement experimentation. Chapter 7 provides a general overview of the most common computational tasks that an organic chemist may want to carry out and walks the reader through a beginner-friendly case study wherein the reactants, transition states, and products of a Diels-Alder reaction are calculated. Chapter 8 builds upon the general knowledge given in the previous chapter and describes how computational chemistry can be used to predict the NMR spectrum of organic molecules. The goal of this chapter is to put this powerful technique into the hands of experimental chemists, which should be achievable after familiarizing themselves with the simplified approach detailed throughout. Chapters 9-11 provide the reader with an introduction to programming and machine learning. Computers already play a critical role in the daily life of a synthetic chemist, and

a little bit of familiarity with modern techniques can go a long way. Chapter 9 provides a blueprint for understanding how and why a chemist may go about familiarizing themselves with programming. Chapter 10 describes a deep dive case study for using machine learning to facilitate reaction optimization, providing a step-by-step guide that a beginner may follow to use the tool and to gain confidence in harnessing other published algorithms. Chapter 11 explains how computers can facilitate the planning of multistep synthesis by suggesting synthetic routes and reaction conditions. Helpful discussions on the current tools available, how they work, and their associated strengths and weaknesses are also described.

This project was only possible due to an immense amount of work by the authors who generously agreed to share their knowledge and meet the formidable task of communicating with a general audience. I am also indebted to the many students and postdoctoral fellows at the University of Ottawa that served as reviewers to help ensure that the content serves as a welcoming and beginnerfriendly introduction to these topics that are becoming increasingly important to the modern synthetic chemists. I hope the readers agree that this goal has been met and that this marks the beginning of their journey to being a more wellrounded scientist capable of tackling diverse problems that come their way.

> Stephen G. Newman Ottawa

> > July 2023

#### 1

## Biocatalysis 101 – A Chemist's Guide to Starting Biocatalysis

Pablo Díaz-Kruik, David Lim, and Francesca Paradisi

Department of Chemistry, Biochemistry and Pharmaceutical Sciences, Bern, Switzerland

#### **Glossary**

API Active pharmaceutical ingredient

**BRENDA** A comprehensive enzyme information system

CALB Lipase B from Candida antarctica

**Cofactor** A non-protein chemical compound or metallic ion that is required for an enzyme's role as a catalyst

**DKR** Dynamic kinetic resolution

**IRED** Imine reductase

NAD+/NADH Nicotinamide adenine dinucleotide

**Protein expression** Biological process where the protein is synthesized inside a cell **Recombinant DNA** DNA scaffold that contains the protein sequence of interest **TRIS** Tris(hydroxymethyl)aminomethane

#### 1.1 Introduction

#### 1.1.1 Enzymes – the Green and Sustainable Way of the Future

Recent efforts by chemists to actively reduce toxic waste production and minimize costs have led to the discovery of many green and sustainable technologies. Not surprisingly, the use of enzymes, Nature's catalysts, has seen a major resurgence in academic and industrial interest over the past decade – not only for their sustainability and natural activities but for engineering them to perform novel transformations beyond capabilities observed in a synthetic organic lab [1, 2].

Enabling Tools and Techniques for Organic Synthesis: A Practical Guide to Experimentation, Automation, and Computation, First Edition. Edited by Stephen G. Newman.

© 2023 John Wiley & Sons, Inc. Published 2023 by John Wiley & Sons, Inc.

The attractiveness of using enzymes for transformations stems from their exquisite regio- and stereoselectivities – something that traditional chemists still struggle to achieve in the lab – that enzymes often execute effortlessly. Moreover, we have seen the emergence of multienzyme cascades for the synthesis of active pharmaceutical ingredients (APIs). A recent landmark example involves the synthesis of molnupiravir (MK-4482), an orally dosed ribonucleoside analogue and inhibitor of influenza viruses, which has demonstrated activity against COVID-19 when administered in animal models [3, 4]. In this work, McIntosh et al. developed a scalable three-step route toward MK-4482 [5]. Using a cascade of five enzymes, MK-4482 could be accessed from 5-isobutyrylribose (Figure 1.1).

To the uninitiated, entering the world of enzyme-catalyzed chemical transformations can be incredibly daunting, especially when one is not equipped with a foundational understanding of what an enzyme is and how these macromolecules work. However, you may be surprised to hear that enzymology and chemistry are not too different from each other at all! With an undergraduate chemistry background, a chemist can easily harness the power of enzymes to perform desired transformations – a *fact* that we aim to convince you of over the next few pages.

However, while this chapter aims to illustrate the power of enzymes for novel and sustainable transformations, we do not want to inadvertently imply the use of these macromolecules is the be-all-end-all solution – sometimes the use of traditional organic synthesis to access target molecules is the more logical solution. Therefore, when an enzyme *might* be used is a weighted question often involving the combination of various intricate factors, including efficiency and cost.

Over the following sections, we will do our best to educate you on these factors so that you can begin making an informed decision on this matter. We also aim to

**Figure 1.1** A combined enzymatic cascade/hydroxylamination for the synthesis of molnupiravir (MK-4482).

convince the reader that the use of enzymes is not limited to biologists and biochemists but also readily available for use by synthetic chemists. With the following breakdown of important considerations to make when using an enzyme, we hope to instill confidence in the reader that a biological catalyst is not too dissimilar to a chemical catalyst and can be readily obtainable from common suppliers.

We will also dispel common misconceptions and myths surrounding the use of enzymes and then give an overview of several classes of reactions that can be performed with enzymes, including recent developments into more exotic transformations such as photobiocatalysis.

This chapter will then conclude with a snippet into recent trends and technologies that have harnessed the use of enzymes in novel ways. We hope that the information gained from reading this chapter will provide a strong foundation for the reader to develop confidence in the use of enzymes and begin their venture into the world of biocatalysis.

#### **Enzymatic and Organic Catalysis Are Not too Different** from Each Other

A seasoned chemist may be quite familiar with several stereoselective reactions whereby stereocontrol is dictated by the chiral environment of the reaction. For example, one model for the Corey-Bakshi-Shibata (CBS) reduction involves coordination of the respective carbonyl to the CBS catalyst in a specific spatial orientation, leading to stereoselective reduction of the carbonyl to the corresponding alcohol (Figure 1.2) [6]. Enzymes utilize a very similar concept to this reaction – the catalyst (enzyme) places a reactant (substrate) in a chiral environment (the active site), whereby stereoselectivity is dictated by the local reactive environment, leading to a selective reaction outcome. In the next subsection, we will look at how an enzyme achieves these feats.

Figure 1.2 The CBS reaction has been used in undergraduate texts as a classic example of where an achiral reactant is stereoselectively transformed in a chiral environment to the corresponding product in high enantiomeric excesses.

#### 1.1.3 Enzymes 101

Enzymes are known to accelerate reactions by more than 10<sup>17</sup>-fold [7]. How an enzyme achieves these colossal rate increases under aqueous conditions requires an understanding of the active site architecture in great molecular detail.

There are 20 essential amino acids found in nature. Enzymes are formed by cellular machinery, which stitch together combinations of these amino acids in a genetically pre-defined sequence, making one very long polymer. This polymer is folded to give a precise three-dimensional structure (Figure 1.3). The active site is defined as the region of the enzyme where substrates bind and undergo catalysis. The catalytic cycle begins with the binding of the substrate in the active site. This process precisely positions all molecules involved in the catalysis (metals, solvents, cofactors, etc.) in their respective orientations ready to achieve regio- and stereoselectivity. Subsequent activation of the substrate initiates the reaction, generating a transition state, which is stabilized by interactions with the active site residues of the enzyme. Following effective conversion of the substrate, the product is then released from the active site of the enzyme, completing one turnover and returning the catalyst back to its original state.

## 1.2 When Should I Choose an Enzyme over a Chemical Catalyst?

The choice of using a chemical catalyst over a biochemical solution needs to be assessed on a case-by-case basis, often involving a detailed cost–benefit analysis. For example, chemical asymmetric imine reduction often requires the use of expensive precious metals, such as Ir, Rh, Ru, and Pd (Figure 1.4) [8]. While recent methods have moved toward Earth-abundant solutions, such as employing iron or nickel, all these still require decoration with expensive chiral ligands that cannot be recycled [8], making the overall synthesis very environmentally and economically demanding.

In contrast, imine reductases (IREDs) can perform stereoselective reductions without the use of expensive metals and can be performed under aqueous conditions mitigating the need for organic solvents. Since the initial report of IREDs in 2010 [9], many advancements have been made to use these enzymes for novel synthetic transformations [9, 10]. In fact, Matzel et al. published an elegant procedure for performing biocatalytic dynamic kinetic resolutions (DKRs) of aldehydes using IREDs (Figure 1.5). This method exploits the stereo-preference of the enzyme for either the *R*- or *S*-chiral center [11].

The use of enzymes in this case showcases the re-opening of the chemical window, enabling unprecedented reaction conditions, merging asymmetric reduction

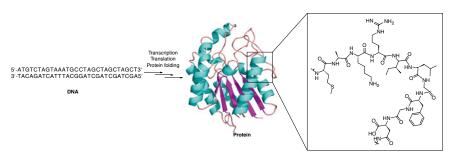


Figure 1.3 Proteins are formed by intracellular machinery that uses genetic information (DNA) to form a polymeric amino acid chain, which is then precisely folded to give a protein.

$$R_3$$
  $N$   $R_1$   $R_2$   $R_3$   $NH$   $R_1$   $R_2$ 

**Figure 1.4** Asymmetric reduction of imines to amines in the presence of a chiral catalyst.

**Figure 1.5** Biocatalytic dynamic kinetic resolutions of aldehydes using imine reductases. GDH = qlucose dehydrogenase.

and water media. This would be near impossible to achieve with classical reducing agents, such as  $NaBH_4$  or  $Na(CH_3COO)_3BH$ .

Next, we move the reader on to build an understanding of the variables associated with using a biocatalyst and here, they will gain foundational knowledge on how to gauge whether a chemical or biochemical solution is appropriate for solving a target problem.

### 1.3 Key Considerations for Running Biocatalytic Reactions

With a primary level of appreciation of the power of enzymes, we can now continue our journey by addressing the variables that define a biocatalytic reaction. We will also illustrate how these variables change depending on the system that is being applied for the reaction. Before we advance in that direction, we will first begin by dissipating common myths surrounding the use of enzymes.

#### 1.3.1 Dispelling Myths

The uptake of biocatalysis in academic and industrial applications has increased significantly in recent years [12, 13]. Despite the positive perception of the technology, the breadth of applications remains rather modest. A factor that contributes to this lack of progress may be associated with the perceived notion of the limitations of biocatalysts – their availability, cost, ease of use, substrate scope, and operational stability. We aim to address these factors in the following subsections.