Tariq Aftab Khalid Rehman Hakeem *Editors*

Antioxidant Defense in Plants

Molecular Basis of Regulation



Antioxidant Defense in Plants

Tariq Aftab • Khalid Rehman Hakeem Editors

Antioxidant Defense in Plants

Molecular Basis of Regulation



Editors
Tariq Aftab Department of Botany
Aligarh Muslim University
Aligarh, Uttar Pradesh, India

Khalid Rehman Hakeem Department of Biological Sciences King Abdulaziz University Jeddah, Saudi Arabia

ISBN 978-981-16-7980-3 ISBN 978-981-16-7981-0 (eBook) https://doi.org/10.1007/978-981-16-7981-0

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Singapore Pte Ltd. The registered company address is: 152 Beach Road, #21-01/04 Gateway East, Singapore 189721, Singapore

Preface

Environmental stresses, such as drought, salinity, or floods, induce the generation of reactive oxygen species (ROS) which causes severe damage to cell membrane integrity by accelerating lipid peroxidation. Growing evidence has suggested that ROS play a critical role as the signaling molecules throughout the entire cell death pathway. Though ROS act as a signaling molecule, they can cause oxidative burst if there is an imbalance between ROS generation and their scavenging. Oxidative stresses also decrease the efficiency of PSI and PSII by disrupting the electron transport chain and chloroplast integrity. Moreover, under severe stress conditions, the generation of ROS often exceeds beyond the antioxidant potential of the plants, resulting in oxidative damages. To counteract the detrimental effect of ROS, plants are inherited with an intricate and vibrant antioxidant defense system, composed of enzymatic (catalase, peroxidase, superoxide dismutase, glutathione reductase, glutathione S-transferase, guaiacol peroxidase, monodehydroascorbate reductase, dehydroascorbate reductase, etc.) and nonenzymatic (glutathione, ascorbate, αtocopherol, carotenoids, flavonoids, etc.) antioxidants, which scavenge and/or reduce excess ROS and improve plant tolerance to abiotic stresses. Stress tolerance in most crop plants is positively correlated with an efficient oxidative system. Therefore, studying the efficiency of antioxidant defense systems in plants is necessary for facilitating the plant's nature of adaptation against abiotic stresses.

Knowledge about the oxidative mechanisms in plants may contribute to the development of plants, adapted to the environment and resistant to pathogens. During the last decades, antioxidant enzymes have been used to develop transgenic plants that have increased tolerance to several stresses. The ROS production, major antioxidant enzymes as well as nonenzymatic antioxidants involved in detoxification, and defense under stresses are the major areas to be elucidated.

The book comprises 20 chapters (review articles) written by experts, highlighting the various enzymatic and nonenzymatic antioxidants, defense mechanisms, and adaptation strategies employed by plants to avoid the stressful conditions. We are hopeful, this volume would furnish the need of all researchers who are working or have interest in this particular field.

We are highly grateful to all our contributors for accepting our invitation and for not only sharing their knowledge and research but also venerably integrating their expertise in dispersed information from diverse fields in composing the chapters and vi Preface

enduring editorial suggestions to finally produce this venture. We also thank Springer-Nature team for their generous cooperation at every stage of the book production.

Lastly, thanks are also due to well-wishers, research students, and editors' family members for their moral support, blessings, and inspiration in the compilation of this book.

Aligarh, Uttar Pradesh, India Jeddah, Saudi Arabia Tariq Aftab Khalid Rehman Hakeem

Contents

I	An Overview of Roles of Enzymatic and Nonenzymatic Antioxidants in Plant Shashi Kant Sharma, Devendra Singh, Himanshu Pandey, Raj Babar Jatav, Virendra Singh, and Devendra Pandey	1
2	Functional Characterization of the Antioxidant Enzymes in Plants Exposed to Environmental Stresses	15
3	Gene Expression and Role of Antioxidant Enzymes in Crop Plants Under Stress Mohd Yasin Bhat, Mir Zahoor Gul, and Jehangir Shafi Dar	31
4	Proteomic and Genomic Approaches for Antioxidant Enzyme-Mediated Defense Analyses in Higher Plants	57
5	Genetic Engineering Applications in Inducing Stress Tolerance in Plants Through Antioxidants	71
6	Kinase-Mediated Signaling Cascades in Plant Abiotic Stress Physiology Shalini Dhiman, Neha Handa, Sukhmeen Kaur Kohli, Mohd Ibrahim, Tamanna Bhardwaj, Dhriti Kapoor, Indu Sharma, Shelja Sareen, Renu Bhardwaj, and Bilal Ahmad Mir	109
7	Plant Peroxidases: Biomarkers of Environmental Stresses and Signaling in Plants	135

viii Contents

8	Molecular Mechanisms of Superoxide Dismutase (SODs)-Mediated Defense in Controlling Oxidative Stress in Plants	157
	Raufa Batool, Muhammad Jawad Umer, Babar Hussain, Muhammad Anees, and Zhenying Wang	107
9	Glutathione in Higher Plants: Biosynthesis and Physiological Mechanisms During Heat and Drought-Induced Oxidative Stress Akbar Hossain, Karma L. Bhutia, Biswajit Pramanick,	181
	Sagar Maitra, Ulkar Ibrahimova, Venugopalan Visha Kumari, Zahoor Ahmad, Muhammad Uzair, and Tariq Aftab	
10	Role of Tocopherol in Conferring Abiotic Stress Tolerance	
	in Plants	215
11	Plant Glutathione Transferases and Their Role in the Mitigation of Abiotic Stresses	235
12	Role of Ascorbic Acid in Alleviating Abiotic Stress in Crop Plants	259
13	CRISPR/Cas-Mediated Genome Editing Technologies in Plants for Stress Resilience	285
14	Decrypting Drought Stress Tolerance of Crop Plants via Photosynthesis and Antioxidative Defense Mechanisms	305
15	Role of Brassinosteroids (BRs) in Modulating Antioxidative Defense Mechanism in Plants Growing Under Abiotic and Biotic Stress Conditions	325

Contents ix

16	Selenium-Mediated Regulation of Antioxidant Defense System and Improved Heavy Metals Tolerance in Plants Zaid Ulhassan, Ali Raza Khan, Mohamed Salah Sheteiwy, Wardah Azhar, Yasir Hamid, Sajad Hussain, Abdul Salam, Muhammad Aqeel Kamran, Khalid Rehman Hakeem, Tariq Aftab, and Weijun Zhou	369
17	Antioxidant Defense System in Plants Against Biotic Stress Najeebul Tarfeen, Qadrul Nisa, Khair-Ul-Nisa, and Kaysar Kahlief	383
18	Revisiting the Crucial Role of Reactive Oxygen Species and Antioxidant Defense in Plant Under Abiotic Stress	397
19	Plant Life Under Changing Environment: An Exertion of Environmental Factors in Oxidative Stress Modulation Sabreena and Shahnawaz Hassan	421
20	Beneficial Role of Phytochemicals in Oxidative Stress Mitigation in Plants Seerat Saleem, Naveed Ul Mushtaq, Wasifa Hafiz Shah, Aadil Rasool, Khalid Rehman Hakeem, and Reiaz Ul Rehman	435

Editors and Contributors

About the Editors



Tariq Aftab received his Ph.D. in the Department of Botany at Aligarh Muslim University, India, and is currently an Assistant Professor there. He is the recipient of a prestigious Leibniz-DAAD fellowship from Germany, Raman Fellowship from the Government of India, and Young Scientist Awards from the State Government of Uttar Pradesh (India) and Government of India. After completing his doctorate, he has worked as Research Fellow at National Bureau of Plant Genetic Resources, New Delhi, and as Post-doctorate Fellow at Jamia Hamdard, New Delhi, India. Dr. Aftab also worked as Visiting Scientist at the Leibniz Institute of Plant Genetics and Crop Plant Research (IPK), Gatersleben, Germany, and in the Department of Plant Biology, Michigan State University, USA. He is a member of various scientific associations from India and abroad.

He has edited 15 books with international publishers, co-authored several book chapters, and published over 75 research/review papers in peer-reviewed international journals. His research interests include physiological, proteomic, and molecular studies on medicinal and crop plants.



Khalid Rehman Hakeem, Ph.D. is Professor at King Abdulaziz University, Jeddah, Saudi Arabia. After completing his doctorate (Botany; specialization in Plant Eco-physiology and Molecular Biology) from Jamia Hamdard, New Delhi, India, in 2011, he worked as a lecturer at the University of Kashmir, Srinagar, for a short period. Later, he joined Universiti Putra Malaysia, Selangor, Malaysia, and worked there as Post-doctorate Fellow in 2012 and Fellow Researcher (Associate Prof.) from 2013 to 2016. Dr. Hakeem has more than 10 years of teaching and research experience in plant eco-physiology, biotechnology and molecular biology, medicinal plant research, plant-microbe-soil interactions as well as in environmental studies. He is the recipient of several fellowships at both national and international levels; also, he has served as the visiting scientist at Jinan University, Guangzhou, China, Currently, he is involved with a number of international research projects with different government organizations.

So far, Dr. Hakeem has authored and edited more than 70 books with international publishers. He also has to his credit more than 140 research publications in peer-reviewed international journals and 60 book chapters in edited volumes with international publishers.

At present, Dr. Hakeem serves as an editorial board member and reviewer of several high-impact international scientific journals.

Contributors

Aqleem Abbas State Key Laboratory of Agricultural Microbiology, Provincial Key Laboratory of Plant Pathology of Hubei Province, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan, Hubei, China

Cbaroymar Abbas Department of Biological Sciences, Karakoram International University, Gilgit, Gilgit-Baltistan, Pakistan

Tariq Aftab Department of Botany, Aligarh Muslim University, Aligarh, Uttar Pradesh, India

Plant Physiology Section, Department of Botany, Aligarh Muslim University, Aligarh, India

Divya Aggarwal MTG Learning Media Pvt. Ltd., Gurugram, Haryana, India

Zahoor Ahmad Department of Botany, University of Central Punjab, Bahawalpur Campus, Bahawalpur, Punjab, Pakistan

Nazir Ahmed Department of Crop Physiology, Sindh Agriculture University, Tandojam, Pakistan

Amina A. M. Al-Mushhin Department of Biology, College of Science and Humanities in Al-Kharj, Prince Sattam Bin Abdulaziz University, Al-Kharj, Saudi Arabia

Taghreed S. Alnusaire Biology Department, College of Science, Jouf University, Sakaka, Saudi Arabia

Muhammad Anees Zhengzhou Fruit Research Institute, Chinese Academy of Agricultural Sciences, Henan Joint International Research Laboratory of South Asian Fruits and Cucurbits, Zhengzhou, China

Muhammad Asim Plant Science Division, Pakistan Agriculture Research Council, Islamabad, Pakistan

Muhammad Mahran Aslam Nuclear Institute of Agriculture (NIA) Tandojam, Sindh, Pakistan

Wardah Azhar Institute of Crop Science, Ministry of Agriculture and Rural Affairs Key Laboratory of Spectroscopy Sensing, Zhejiang University, Hangzhou, China

Raufa Batool State Key Laboratory for Biology of Plant Diseases and Insect Pest, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing, China

Renu Bhardwaj Department of Botanical and Environmental Sciences, Guru Nanak Dev University, Amritsar, Punjab, India

Tamanna Bhardwaj Department of Botanical and Environmental Sciences, Guru Nanak Dev University, Amritsar, Punjab, India

Mohd Yasin Bhat Department of Plant Sciences, School of Life Sciences, University of Hyderabad, Hyderabad, Telangana, India

Karma L. Bhutia Department of Agricultural Biotechnology & Molecular Breeding, College of Basic Science and Humanities, Dr. Rajendra Prasad Central Agricultural University, Pusa, Bihar, India

Sadaruddin Chachar Department of Biotechnology, Sindh Agriculture University, Tandojam, Pakistan

Jehangir Shafi Dar Centre of Research for Development, University of Kashmir, Hazratbal, Srinagar, Jammu and Kashmir, India

Shalini Dhiman Department of Botanical and Environmental Sciences, Guru Nanak Dev University, Amritsar, Punjab, India

Anupam Dikshit Biological Product Laboratory, Department of Botany, University of Allahabad, Prayagraj, Uttar Pradesh, India

Saif ud Din Department of Biological Sciences, Karakoram International University, Gilgit, Gilgit-Baltistan, Pakistan

Arneeb Tariq Department of Botany, Government College Women University, Faisalabad, Pakistan

Uttam Kumar Ghosh Department of Agronomy, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

Mir Zahoor Gul Department of Biochemistry, University College of Sciences, Osmania University, Hyderabad, Telangana, India

Khalid Rehman Hakeem Department of Biological Sciences, Faculty of Science, King Abdulaziz University, Jeddah, Kingdom of Saudi Arabia

Yasir Hamid Ministry of Education (MOE) Key Lab of Environmental Remediation and Ecosystem Health, College of Environmental and Resources Science, Zhejiang University, Hangzhou, China

Neha Handa Division of Research and Development, Department of Research Facilitation, Lovely Professional University, Phagwara, Punjab, India

Shahnawaz Hassan Department of Environmental Science, University of Kashmir, Srinagar, Jammu and Kashmir, India

Akbar Hossain Bangladesh Wheat and Maize Research Institute, Dinajpur, Bangladesh

Md. Saddam Hossain Department of Agronomy, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

Babar Hussain Department of Biological Sciences, Karakoram International University, Gilgit, Pakistan

Instituto de Investigación para el Desarrollo Sustentable de Ceja de Selva, Universidad Nacional Toribio Rodríguez de Mendoza de Amazonas, Chachapoyas, Peru

Institute of Agricultural Resources and Regional Planning, Chinese Academy of Agricultural Sciences, Beijing, China

Sajad Hussain Institute of Ecological Agriculture, Sichuan Agricultural University/Sichuan Engineering Research Centre for Crop Strip Intercropping System, Chengdu, People's Republic of China

Moh Ibrahim Department of Botanical and Environmental Sciences, Guru Nanak Dev University, Amritsar, Punjab, India

Ulkar Ibrahimova Institute of Molecular Biology and Biotechnologies, Azerbaijan National Academy of Sciences, Baku, AZ, Azerbaijan

Rashid Iqbal Department of Agronomy, Faculty of Agriculture and Environment, The Islamia University of Bahawalpur, Bahawalpur, Punjab, Pakistan

Md. Nahidul Islam Department of Agro-Processing, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

Raj Babar Jatav G.H. Raisoni University, Saikheda, Borgaon, Madhya Pradesh, India

Areej Javaid Department of Botany, Government College Women University, Faisalabad. Pakistan

Khadija Javeed Department of Environmental Science, PMAS-Arid Agriculture University, Rawalpindi, Punjab, Pakistan

The State Key Laboratory for Bio-Pesticides Engineering of Plant Disease Biocontrol and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Science, Beijing, China

Gunja Jha Department of Biological Sciences, GD Goenka University, Gurugram, Haryana, India

Saket Jha Biological Product Laboratory, Department of Botany, University of Allahabad, Prayagraj, Uttar Pradesh, India

Kaysar Kahlief Department of Management Studies, Lovely Professional University, Phagwara, Punjab, India

Muhammad Aqeel Kamran Ministry of Education (MOE) Key Lab of Environmental Remediation and Ecosystem Health, College of Environmental and Resources Science, Zhejiang University, Hangzhou, China

Dhriti Kapoor Department of Botany, School of Bioengineering and Biosciences, Lovely Professional University, Phagwara, Punjab, India

Ravinderjit Kaur Department of Zoology, S.R. Govt. College for Women, Amritsar, India

Khair-Ul-Nisa Department of Environmental Science, University of Kashmir, Srinagar, India

Anjali Khajuria Department of Zoology, Central University of Jammu, Jammu, Jammu and Kashmir, India

Ali Raza Khan Institute of Crop Science, Ministry of Agriculture and Rural Affairs Key Laboratory of Spectroscopy Sensing, Zhejiang University, Hangzhou, China

Md. Arifur Rahman Khan Department of Agronomy, Bangabandhu Sheikh Mujibur Rahman Agricultural University, Gazipur, Bangladesh

Sher Wali Khan Department of Biological Sciences, Karakoram International University, Gilgit, Gilgit-Baltistan, Pakistan

Sukhmeen Kaur Kohli Department of Botanical and Environmental Sciences, Guru Nanak Dev University, Amritsar, Punjab, India

Sandeep Kour Nematology Lab, Department of Zoology, Guru Nanak Dev University, Amritsar, India

Deepak Kumar Nematology Lab, Department of Zoology, Guru Nanak Dev University, Amritsar, India

Venugopalan Visha Kumari Department of Agronomy, Bidhan Chandra KrishiVishwavidalya, Mohanpur, West Bengal, India

Sagar Maitra Department of Agronomy, Centurion University of Technology and Management, Paralakhemundi, India

Bilal Ahmad Mir Department of Botany, School of Life Sciences, Satellite Campus, Kargil, University of Kashmir, Srinagar, Jammu and Kashmir, India

Naveed Ul Mushtaq Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Rashda Naheed Department of Botany, Government College Women University, Faisalabad, Punjab, Pakistan

Qadrul Nisa Division of Plant Pathology, SKUAST-K, Shalimar, Srinagar, India

Puja Ohri Nematology Lab, Department of Zoology, Guru Nanak Dev University, Amritsar, India

Devendra Pandey Central Institute for Subtropical Horticulture, Lucknow, Uttar Pradesh, India

Himanshu Pandey Dr. Y.S. Parmar University of Horticulture and Forestry, U.H.F. Nauni, Solan, Himachal Pradesh, India

Deepu Pandita Government Department of School Education, Jammu, Jammu and Kashmir, India

Biswajit Pramanick Department of Agronomy, Dr. Rajendra Prasad Central Agricultural University, Pusa, Bihar, India

Huma Maqbool Rai Department of Botany, Government College Women University, Faisalabad, Pakistan

Shameem Raja Department of Botany, Government College Women University, Faisalabad, Pakistan

Umar Rao Institute of Plant Breeding and Biotechnology, MNS-University of Agriculture, Multan, Pakistan

Aadil Rasool Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Reiaz Ul Rehman Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Samiya Rehman Department of Biochemistry, University of Okara, Okara, Punjab, Pakistan

Luqman Riaz College of Life Sciences, Henan Normal University, Xinxiang, China

Sabreena Department of Environmental Science, University of Kashmir, Srinagar, Jammu and Kashmir, India

Sana Saeed Department of Botany, Government College Women University, Faisalabad, Pakistan

Abdul Salam Institute of Crop Science, Ministry of Agriculture and Rural Affairs Key Laboratory of Spectroscopy Sensing, Zhejiang University, Hangzhou, China

Seerat Saleem Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Shelja Sareen Department of Biotechnology, BBK DAV College for Women, Amritsar, Punjab, India

Wasifa Hafiz Shah Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Iram Shahzadi Department of Botany, Government College Women University, Faisalabad, Pakistan

Indu Sharma Department of Botany, Sant Baba Bhag Singh University, Jalandhar, Punjab, India

Nandni Sharma Nematology Lab, Department of Zoology, Guru Nanak Dev University, Amritsar, India

Shashi Kant Sharma Junagadh Agriculture University, Junagadh, Gujarat, India

Mohamed Salah Sheteiwy Department of Agronomy, Faculty of Agriculture, Mansoura University, Mansoura, Egypt

Mohee Shukla Biological Product Laboratory, Department of Botany, University of Allahabad, Prayagraj, Uttar Pradesh, India

Mohammad Aquil Siddiqui Nuclear Institute of Agriculture (NIA) Tandojam, Sindh. Pakistan

Devendra Singh Motilal Nehru National Institute of Technology, Allahabad, Prayagraj, Uttar Pradesh, India

Parul Singh Department of Biological Sciences, GD Goenka University, Gurugram, Haryana, India

Ravikant Singh Department of Biotechnology, Swami Vivekanand University, Sagar, Madhya Pradesh, India

Virendra Singh Maulana Azad Medical College, New Delhi, India

Mona H. Soliman Botany and Microbiology Department, Faculty of Science, Cairo University, Giza, Egypt

Biology Department, Faculty of Science, Taibah University, Al-Sharm, Yanbu El-Bahr, Yanbu, Saudi Arabia

Muhammad Muneeb Subhani Centre of Agriculture Biochemistry and Biotechnology, Faisalabad, Punjab, Pakistan

Tahira Tabassum Department of Biology/Faculty of Life Sciences, University of Okara, Okara, Pakistan

Hafiza Naila Tabbasum Centre of Agriculture Biochemistry and Biotechnology, Faisalabad, Punjab, Pakistan

Inayatullah Tahir Department of Botany, School of Biological Sciences, University of Kashmir, Srinagar, Jammu and Kashmir, India

Najeebul Tarfeen Centre of Research for Development (CORD), University of Kashmir, Srinagar, India

Fozia Farhat Department of Botany, Government College Women University, Faisalabad, Pakistan

Naveed Ul Mushtaq Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, India

Reiaz Ul Rehman Department of Bioresources, School of Biological Sciences, University of Kashmir, Srinagar, India

Zaid Ulhassan Institute of Crop Science, Ministry of Agriculture and Rural Affairs Key Laboratory of Spectroscopy Sensing, Zhejiang University, Hangzhou, China

Muhammad Jawad Umer State Key Laboratory of Cotton Biology/Institute of Cotton Research, Chinese Academy of Agricultural Sciences (ICR, CAAS), Anyang, China

Muhammad Uzair Department of Botany, University of Central Punjab, Bahawalpur Campus, Bahawalpur, Punjab, Pakistan

Zhenying Wang State Key Laboratory for Biology of Plant Diseases and Insect Pest, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing, China

Weijun Zhou Institute of Crop Science, Ministry of Agriculture and Rural Affairs Key Laboratory of Spectroscopy Sensing, Zhejiang University, Hangzhou, China

Saman Zulfiqar Department of Botany, The Government Sadiq College Women University, Bahawalpur, Pakistan

An Overview of Roles of Enzymatic and Nonenzymatic Antioxidants in Plant

Shashi Kant Sharma, Devendra Singh, Himanshu Pandey, Raj Babar Jatav, Virendra Singh, and Devendra Pandey

Abstract

The postindustrial age radically altered global climate conditions, posing a difficult task for plants and crops to thrive under stress environments like high temperatures, salt, waterlogging, heavy metals, drought, and so on. A small period of poor weather had a substantial impact on the development and growth of plants, eventually influencing crop quality, yield, and agricultural sustainability as a whole. Plant cells produce free oxygen (O2) radicals and their derivatives, known as reactive oxygen species (ROS), as by-products of other reactions in such hostile environments. Furthermore, these ROS molecules are used as signaling molecules in plants for signal transduction in response to changing environmental conditions. The cytoplasmic balance that triggers the antioxidant defense mechanisms is disrupted as a result of the excessive accumulation of ROSs inside the cell. Plants have developed a complicated ROS scavenging system to avoid sensitive cellular components from being damaged

Junagadh Agriculture University, Junagadh, Gujarat, India

D. Singh (⊠)

Motilal Nehru National Institute of Technology, Allahabad, Prayagraj, Uttar Pradesh, India

Dr. Y.S. Parmar University of Horticulture and Forestry, U.H.F. Nauni, Solan, Himachal Pradesh, India

R. B. Jatav

G.H. Raisoni University, Saikheda, Borgaon, Madhya Pradesh, India

V. Singh

Maulana Azad Medical College, New Delhi, India

Central Institute for Subtropical Horticulture, Lucknow, Uttar Pradesh, India

S. K. Sharma

[©] The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2022

T. Aftab, K. R. Hakeem (eds.), Antioxidant Defense in Plants,

by reactive oxygen species. Enzymatic antioxidants, like catalase (CAT), peroxidase (POD), superoxide dismutase (SOD), glutathione peroxidases (GPX), and ascorbate peroxidases (APX), and nonenzymatic antioxidants, like glutathione, ascorbate, tocopherols, and phenolic compounds, are important antioxidants that play key roles in eliminating superoxide (O_2^-) and (H_2O_2) . The antioxidant capacity of plants is the sum of the activities of all enzymatic and nonenzymatic antioxidant systems. This chapter seeks to provide fundamental information on enzymatic and nonenzymatic antioxidants, their occurrence, characteristics, and the antioxidant defense system involved in reactive oxygen species (ROS) detoxification under various stresses, as well as their interactions with cellular components.

Keywords

 $ROS \cdot Abiotic \ stress \cdot Oxidative \ stress \cdot Enzymatic \ antioxidants \cdot Nonenzymatic \ antioxidants$

1.1 Introduction

Based on their biochemical nature, antioxidants are divided into two categories: enzymatic and nonenzymatic. While substantial anabolic and catabolic reactions are occurring, these chemicals are engaged in the detoxification of free radicals or reactive oxygen species (ROS). Both classes of antioxidants are capable of efficiently neutralizing ROS and converting them into relatively stable nontoxic molecules, preventing oxidative damage to cellular apparatuses. As a result, antioxidants are the most important first line of defense against the oxidative stress-induced cell damage. Antioxidants, both enzymatic and nonenzymatic, are electron-rich compounds that readily share electrons with highly energetic ROS and free radicals, stabilizing cellular randomness.

Furthermore, they may interfere with the oxidizing chain reaction in order to reduce free radical damage (Apel and Hirt 2004). Antioxidants are also known as ROS scavengers because they use dynamic and synergistic processes to keep the intracellular concentration of ROS in check. It (antioxidant) is a substance that may scavenge reactive oxygen species (ROS) without being converted into a harmful radical (Noctor and Foyer 1998). As a result, antioxidant enzymes are crucial for sustaining good cellular and systemic health and well-being. All highly reactive, oxygen-containing molecules, including the free radicals, are referred to as ROS. The hydroxyl radical (OH $^-$), singlet oxygen, superoxide anion radical, $\rm H_2O_2$ (hydrogen peroxide), hypochlorite radicals, lipid peroxides, and nitric oxide radical are all examples of reactive oxygen species. All have the ability to react with membrane lipids, enzymes, and other molecules, resulting in the loss of critical cellular structures and functions and a variety of negative consequences for plants and animals. As previously stated, free radicals or ROS are highly reactive compounds that are released directly or as a by-product during normal metabolic processes in

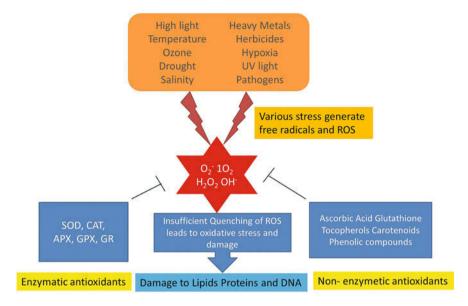


Fig. 1.1 Illustrative representation of different agents generating free radicals and reactive oxygen species in plants and different antioxidants scavenging agents

various cellular compartments like mitochondria, chloroplast, peroxisomes, and apoplast (Panieri and Santoro 2015), but their synthesis is accelerated during extreme conditions. Because these compounds have a lone pair of electrons, they are very unstable and thus highly reactive (Del Río and López-Huertas 2016). ROS-induced oxidative stress is a well-controlled process, and the balance in-between the ROS and its quenching define a plant's and any other organism's well-being. If antioxidants reach a condition of disequilibrium, meaning they are unable to destroy free radicals, the cell and tissue suffer oxidative damage. The degree of oxidative stress caused by free radicals/ROS is determined by their concentration, kind, synthesis site, and developmental stage (Møller et al. 2007). Plants exposed to ROS for a long time period can suffer considerable damage to their cell machinery and biomolecules, including protein oxidation, lipid peroxidation (Mittler 2002), PCD (programmed cell death), and inhibition of the enzymes and also damages nucleic acid, which can lead to tissue necrosis or plant death (Fig. 1.1) (Pérez-Pérez et al. 2012).

1.2 Enzymatic Antioxidants

Several enzymes, like GPX, SOD, glutathione reductase, APX, and CAT (catalase), are the enzymatic components of the antioxidant defense system found in diverse subcellular compartments. In plants under oxidative stress, these enzymatic antioxidant molecules are critical for maintaining cellular homeostasis.

1.2.1 SOD (Superoxide Dismutases)

SODs are enzymes which can catalyze the dismutation/partitioning of O₂⁻ to H₂O₂ and molecular oxygen (O₂). Because O₂⁻ is a typical consequence of oxygen metabolism, SOD is thought to be the first line of defense against the reactive O₂ species-induced damage. By neutralizing the superoxide radical, the Haber-Weiss reaction eliminates the possibility of hydroxyl ion production. These enzymes are classed as metalloenzymes based on metal cofactors, subcellular distribution, and protein folds. Cu-/Zn-containing SODs have been found in prokaryotic and eukaryotic organisms, and in plant cells, they can be found in the cytoplasm, extracellular space, or chloroplasts and can also be found in lysosomes, cytoplasm, and nuclear compartments in mammalian cells. Plant cytoplasm and chloroplasts have been found to have Fe-containing SODs. MnSODs are found in all kingdoms but particularly in eukaryotic mitochondria, where they play a critical role in preserving mitochondria by scavenging ROS (Pilon et al. 2011) and initiating cellular differentiation (Moller 2001, 2012). According to Feng et al. (2016), SODs are found in different organisms, such as mammals, yeast, bacteria, and plants. Multiple genes producing SODs exist in plants, and their expression can be influenced by developmental stage, tissue type, and environmental cues (Scandalios 2005; Menezes-Benavente et al. 2004). There are nine SOD genes in tomatoes, which are unevenly distributed across 12 chromosomes and include four Cu/ZnSODs, one MnSOD, and three FeSODs (Feng et al. 2016). Various investigations revealed that different types of SOD genes had varied levels of expression under harsh environmental conditions. Underwater stress, for example, the expression patterns of the banana genes like MaCSD1B and MaMSD1A, which are involved in SOD production, were utterly incompatible (Feng et al. 2015).

Furthermore, SODs with the same cofactor may not necessarily have the same role in various species. MnSOD expression was not affected by oxidative stress in *Arabidopsis*, but it was affected significantly by drought and cold stress in wheat and salt stress in pea (Baek and Skinner 2003). Additionally, alternative splicing and miRNAs have been implicated in the regulation of SOD gene expression (Lu et al. 2010). Various plant species, such as *Populus trichocarpa*, *Sorghum bicolor*, *Musa acuminata*, and *Arabidopsis thaliana*, have been found to contain the SOD genes (Srivastava et al. 2009).

1.2.2 CAT (Catalases)

These are tetrameric enzymes, with iron as a prosthetic group attached to each monomer; they catalyze the energy-efficient disproportionation of H_2O_2 into water molecules (H_2O) and molecular oxygen (Regelsberger et al. 2002; Zamocky et al. 2008). In contrast to H_2O_2 , it has a lower affinity for R-O-O-R (organic peroxides) and also has a higher turnover rate among antioxidant enzymes and is unique among antioxidant enzymes because it does not need a reducing counterpart. Unfavorable conditions necessitate increased energy generation, and expenditure by plants and

other organisms leads to an increase in catabolic reactions, which yields H_2O_2 . CATs are predominantly attached to peroxisomes, where they execute β -oxidation of fatty acid, photorespiration, and purine catabolism, all of which produce a considerable amount of H_2O_2 (Mittler 2002). Hydrogen peroxide (H_2O_2) is also a key signaling molecule in plant development and plant response to the environment (Mhamdi et al. 2010). Plants have numerous CATs, each produced by a separate gene, that respond differently to different stressors that are known to cause ROS. Recent investigations reveal that CAT is also prevalent in other subcellular compartments like the mitochondria, chloroplast, and cytosol, albeit no evidence of substantial CAT activity has been observed (Mhamdi et al. 2010). CAT1 is mostly expressed in plant pollen and seeds; on the other side, CAT2 is not only expressed in photosynthetic tissues (mostly) but also expressed in seeds and roots, while CAT3 is predominantly expressed in vascular tissues and leaf in angiosperms (McClung 1997; Frugoli et al. 1996).

1.2.3 APX (Ascorbate Peroxidases)

Ascorbate peroxidases are the heme peroxidase superfamily that is involved in the recycling of AsA and the response to environmental stress in plants (Ishikawa and Shigeoka 2008; Lazzarotto et al. 2011). These enzymes catalyze the conversion of H₂O₂ to water and monodehydroascorbate (MDHA) using ascorbate (AsA) as an electron donor (Caverzan et al. 2012). Plants have five different isoforms of APX, which are found in diverse subcellular compartments like the cytosol, mitochondria, peroxisomes, and chloroplast (Sharma and Dubey 2005). These enzymes are divided into groups based on the amino acids they contain and the cell compartments in which they are found. Several abiotic stressors in plants have varied effects on the APX genes (Rosa et al. 2010; Caverzan et al. 2014). APXs, CATs, and SODs must be in balance to determine the effective intracellular level of H₂O₂ and oxygen, and variations in this equilibrium appear to trigger compensation processes (Apel and Hirt 2004; Scandalios 2002, 2005). Under a normal and stressful environment, APX is a key component of the ASC-GSH (ascorbate-glutathione) cycle, which eliminates excess H₂O₂ from plant cells (Mittler and Zilinskas 1991). Because APX is more broadly distributed and also has a higher affinity for hydrogen peroxide than CAT, it is a more effective H₂O₂ scavenger during stressful situations. APX is a chloroplastic isoenzyme expressed by a single gene in higher plants (apx1).

1.2.4 GPx (Glutathione Peroxidases)

The term glutathione peroxidases collectively describe the group of phylogenetically related nonheme and thiol-containing peroxidase enzymes. It was called after GPx-1, the mammalian tetrameric seleno enzyme, which was the first to be defined and reported. More than 700 members of the GPx family have been identified so far, overall domains of life. G. C. Mills discovered its activity in red blood cells in 1957,

where it protected hemoglobin from oxidative degradation. The biological function of GPxs is to catalyze the $\rm H_2O_2$ reduction or lipid hydroperoxides into $\rm H_2O$ or the corresponding alcohols by using GSH (glutathione) as a reducing agent. The catalytic moiety of selenium was later identified as selenocysteine, the 21st naturally occurring amino acid. Selenocysteine was recommended as a way to ensure a rapid reaction with the hydroperoxide and glutathione reducibility. GPx1 is found in the mitochondria, chloroplasts, and cytosol of a wide range of plants and animals, and it serves as an antioxidant in several cellular compartments. In human beings, there are eight distinct glutathione peroxidase isoforms (GPx1-8) that have been found. The mechanism of reaction comprises oxidation of the selenol of a selenocysteine residue by $\rm H_2O_2$. The RSeOH (selenenic acid) group is formed as a result of this action. A two-step mechanism converts selenenic acid back to selenol, starting with a reaction with GSH to create the GS-SeR and $\rm H_2O$. The GS-SeR intermediate is reduced back to selenol by a second GSH molecule, releasing a by-product GS-SG.

1.3 Nonenzymatic Antioxidants

Antioxidants that are generally nonenzymatic in nature are found in all subcellular parts of plants. These antioxidants help to detoxify reactive O_2 species and free radicals and can also help to reduce the substrates from antioxidant enzymes (Mittler 2002). The primary cellular redox buffers GSH and ascorbate, along with carotenoids, tocopherol, and polyphenolic compounds, are different forms of nonenzymatic defense pathways (Scandalios 2002).

1.3.1 Vitamin C (Ascorbic Acid)

Ascorbate is a well-known vitamin having anti-oxidizing properties that have been found in a variety of organelles and even apoplast. It occurs in different reduced and oxidized forms, as ascorbic acid (90% of the ascorbate pool) and mono- and dehydroascorbic acid (Smirnoff 2011). The ratio of oxidized to reduced ascorbate is a key element that influences plant oxidative stress resistance (Conklin et al. 2000; Cruz-Rus et al. 2012). Glutathione reductase, dehydroascorbate reductase, and monodehydroascorbate reductase are among the NAD(P)H-dependent enzymes that keep AsA in its reduced state (Mittler 2002; Foyer and Noctor 2011). According to the Horemans et al. (2000), the mitochondria are the major site for ascorbate production, from which it is transferred to other cell organs via a proton-electron gradient. Due to its ability to transfer electrons in enzymatic and nonenzymatic processes, AsA is a critical component for ROS detoxification in the aqueous phase. AsA can quickly remove O_2^- , hydroxide ions, and $1O_2$, and therefore, it can reduce H₂O₂ to H₂O by the ascorbate peroxidase mechanism, protecting membranes (Blokhina et al. 2003). In the ascorbate-glutathione cycle, APX utilizes two AsA molecules to decrease H₂O₂ to water, with the intermediate monodehydroascorbate, which is a short-lived radical that is further disproportionated into dehydroascorbate (DHA) and AsA. Furthermore, it plays a crucial role in the dynamic and control of the cell cycle, the advancement of the G1 to S stage of the cell division pathway, and cellular elongation, as well as many photosynthetic functions (Smirnoff 2011). Although the exact biosynthetic process for ascorbate is unknown, it is speculated that D-glucose may be used as a precursor.

1.3.2 Glutathione

Glutathione (GSH) oxidation by ROS results in the formation of GSSG, which is present in all plant cell compartments. In cellular compartments, GSH, together with its oxidized counterpart, GSSG, maintains a redox balance. It has been revealed that the GSH/GSSH pair is involved in gene regulation and cell cycle pathways (Mittler 2002). The antioxidant glutathione and ascorbic acid are prolific and stable, and they have the right redox potential to react with different varieties of substrate and compounds. Glutathione is a tripeptide molecule made up of three amino acids, glutamine, cysteine, and glycine, that are found in all plant cell parts, including the cytoplasm, vacuoles, chloroplasts, mitochondria, and endoplasmic reticulum (Millar et al. 2003). In most plant cells, glutathione is the important source of nonprotein thiols. Glutathione is appropriate for a wide range of metabolic actions in all organisms due to the presence of a thiol group and its reactivity. The reduced form of glutathione (GSH) is found at higher concentrations in chloroplasts. Unlike the ascorbate biogenesis system, the glutathione biosynthetic pathway is wellestablished and identical throughout all domains of life. The amino acids are linked to produce the full tripeptide in two ATP-dependent stages catalyzed by GSHS and γ -ECS. These events take place in chloroplastic and non-chloroplastic segments, and glutathione concentrations and redox status play a crucial role in different pathways. GSH is used to reduce DHA both in an enzymatic and nonenzymatic way in the ascorbate-glutathione pathway, and it is then oxidized to GSSG. Glutathione reductase catalyzes the regeneration of GSH from GSSG, with NADPH as the reducing agent. The cysteine residue in GSH's tripeptide has a high reactivity potential. GSH scavenges harmful H₂O₂ by reacting nonenzymatically with O₂⁻, OH⁻, and 1O₂. The ability of GSH to replenish another potent antioxidant, ascorbic acid, via the ascorbate–glutathione cycle gives it a crucial role in antioxidative defense (Millar et al. 2003). It has been observed that the antioxidant property of glutathione was utilized to achieve transgenic lines of tobacco (Foyer and Noctor 2005), which can withstand oxidative stress (Del Río and López-Huertas 2016). GSH has been involved in sensing alterations in redox equilibrium and transferring these alterations to appropriate target proteins, in addition to being a co-substrate and reducing agent in defense against reactive oxygen species.

1.3.3 Vitamin E

All types of tocopherols are methylated fat-soluble phenols that have similar to vitamin E (Sharma et al. 2012). They primarily function as antioxidants in plants, algae, and mammals, but they may also have additional functions. In lipid bilayers, α -tocopherol is the most effective for the removal of peroxyl radicals. Tocopherols are the most powerful scavengers of free radicals. At the energy of 323 kJ mol⁻¹, the hydroxyl bond present in vitamin E becomes weaker than in the majority of phenols and readily liberates hydrogen atom and thereby helps in quenching peroxyl radicals and other free radicals, decreasing their harmful effect (Lide 2006). The produced tocopheryl radical is generally nonreactive, and through redox interaction with a hydrogen donor such as ascorbate or other antioxidants, it reverts to tocopherol (Igamberdiev and Hill 2004; Traber and Stevens 2011). Tocopherols are integrated into cell membranes and thereby protect the chloroplast membrane from oxidative damage due to their fat-soluble nature (Blokhina et al. 2003). α-Tocopherols are significantly bioactive and prominent antioxidants in the chloroplast lamina and are primarily utilized for defending them against the photooxidative effect. It has been observed that a single α-tocopherol molecule may neutralize one 20 singlet oxygen and also act as reusable chain reaction terminators for PUFA radicals synthesized during oxidation of lipids (Hare et al. 1998; Wu and Tang 2004; Ledford and Niyogi 2005). Photosynthesis and other metabolic processes in chloroplasts produce ROS, which causes lipid peroxidation in plant cells. To cope up with a range of abiotic stress conditions, the content of α-tocopherol in photosynthetic plant tissues increases dramatically (Noctor 2006). The ability of α-tocopherols to scavenge and quench ROS aids in the modulation of signal transduction and the stabilization of membranes (Kruk et al. 2005; Noctor 2006). They work as a free radical capturing process by preventing the chain extension stage in lipid autooxidation. Plants respond to oxidative stress by expressing genes involved in tocopherol production (Table 1.1) (Giacomelli et al. 2007; Wu and Tang 2004).

1.3.4 Carotenoids

Carotenoids, also known as tetra terpenoids, are pigments that naturally occur in plants and microorganisms (Otles and Cagindi 2008). To date, more than 750 natural carotenoids have been discovered. These chemicals give different vegetables and fruits their distinct colors. Xanthophylls and carotenes are the two broad categories of carotenoids. Carotenoids are classified into two broad subclasses, xanthophylls and carotenes; the former class contains oxygen, whereas the latter class is purely hydrocarbons and contains no oxygen (Ngamwonglumlert et al. 2017). Carotenoids found in the human diet may help to prevent cancer, age-related muscle degeneration, atherosclerosis, and other disorders. These pigments are lipid-soluble and are absorbed together with fats through the gut tract. Carotenoids have a variety of beneficial activities in plants, including attracting pollinators, indicating fruit development, assisting in photosynthesis, and protecting cells from light-induced damage

Table 1.1 ROS and their characteristics

ROS	Site of generation	Mode of action	Interaction with protein	Interaction with DNA	Scavenging systems
02_	Membranes, mitochondria, chloroplast	The reaction occurs with a double bond having molecules such as iron-sulfur proteins	Through Fe-center	Excessively low	Super oxide dismutase
_HO	Membranes, chloroplast, mitochondria	Highly reactive with different biomolecules	Fast reacting	Quick reacting	Flavonoid and proline
H_2O_2	Membranes, mitochondria, chloroplasts peroxisomes	Oxidizes proteins and help in the generation of $$\operatorname{\textbf{Reacts}}$$ with Cys residue OH $^-$ through O_2^-	Reacts with Cys residue	Excessively low	CAT and flavonoid
O ₂	Membranes, mitochondria chloroplasts	Oxidizes proteins, polyunsaturated fatty acid, and deoxyribonucleic acid	Highly reactive for Trp, Met, His, Tyr, and Cys amino acids moties	Polyunsaturated fatty acid	Carotenoids and vitamin E

in plants and in photosynthetic bacteria and algae (Lerfall 2016). The ability of β -carotene to quench free radicals such as O_2^- , OH^- , and $1O_2$ without undergoing any changes or degradation is largely due to its highly delocalized polyene backbone or conjugated double bond structure, which is primarily responsible for its antioxidant behavior. Carotenoids found in higher concentrations in particular tissues and organs can protect lipids against oxidative damage.

1.3.5 Phenolic Compounds

Tannins, flavonoids, lignins, and stilbenes are examples of phenolic compounds, which constitute a varied group of naturally occurring secondary metabolites common in plants. Multiple phenol rings distinguish these compounds, making them suited for free radical scavenging in both de novo and in vitro conditions. Over 8000 phenolic bioactive substances have been detected in different plant families. Phenylalanine/shikimic acid is the common intermediate precursor for all plant phenolic compounds. Under in vitro conditions, polyphenols have been shown to be more potent antioxidants than ascorbate and tocopherols. Phenols are the most significant dietary elements for humans, providing bitterness, color, astringency, flavor, odor, and oxidative stability in food (Schroeter et al. 2002). Antioxidative activities of polyphenols are characterized by the presence of readily available donor hydrogen or electron (Rice-Evans et al. 1997). In another way, phenols alter the peroxidation kinetics of lipid membrane and packaging, thereby protecting the membrane integrity (Schroeter et al. 2002). Furthermore, phenolics have been implicated in the H₂O₂ scavenging cascade in plant cells. Polyphenols play different functions in plants, which include pigmentation to plants; increase and decrease of plant growth regulators, for example, auxin; UV protectants against ionizing light; deterrence to herbivores; phytoalexins; and signaling compounds in ripening and other plant developmental activities (Huber et al. 2003; Lattanzio et al. 2006).

1.4 Conclusion

Antioxidants are created normally, but they are triggered and upregulated in stressful situations, which help to retain the structural firmness of cell organelles while probably reducing oxidative damage. Plant defense is aided by a number of antioxidant enzymes. The production and activation of ROS scavenging enzyme systems in transgenic plants to increase their tolerance to a variety of stress conditions. Furthermore, because numerous enzymes and their different isoforms are involved and reactive oxygen species is only one of the major factors of plant resistance to unfavorable environmental and biotic stimuli, further research is needed in this field. The increasing number of articles addressing superoxide dismutase, common antioxidant enzyme, ascorbate peroxidase, glutathione peroxidase enzyme, and glutathione reductase enzymes demonstrates these enzymes' favorable responses to biotic and abiotic stressors. These findings highlight the need to investigate these

enzymes in order to better understand their role in the scavenging of hazardous cell products in a variety of species and the relationship between biological processes and oxidative stress.

References

- Apel K, Hirt H (2004) Reactive oxygen species: Metabolism, oxidative stress and signal transduction. Annu Rev Plant Biol 55:373–399
- Baek KH, Skinner DZ (2003) Alteration of antioxidant enzyme gene expression during cold acclimation of near-isogenic wheat lines. Plant Sci 165(6):1221–1227
- Blokhina O, Virolainen E, Fagerstedt KV (2003) Antioxidants, oxidative damage and oxygen deprivation stress: A review. Ann Bot 91(2):179–194
- Caverzan A, Passaia G, Rosa SB, Ribeiro CW, Lazzarotto F, Margis-Pinheiro M (2012) Plant responses to stresses: role of ascorbate peroxidase in the antioxidant protection. Genet Mol Biol 35(4):1011–1019
- Caverzan A, Bonifacio A, Carvalho FEL, Andrade CMB, Passaia G, Schünemann M, Maraschin FS, Martins MO, Teixeira FK, Rauber R, Margis R, Silveira JAG, Margis-Pinheiro M (2014) The knockdown of chloroplastic ascorbate peroxidases reveals its regulatory role in the photosynthesis and protection under photo-oxidative stress in rice. Plant Sci 214:74–87
- Conklin PL, Saracco SA, Norris SR, Last RL (2000) Identification of ascorbic acid-deficient Arabidopsis thaliana mutants. Genetics 154(2):847–856
- Cruz-Rus E, Amaya I, Valpuesta V (2012) The challenge of increasing vitamin C content in plant foods. Biotechnol J 7(9):1110–1121
- Del Río LA, López-Huertas E (2016) ROS generation in peroxisomes and its role in cell signaling. Plant Cell Physiol 57(7):1364–1376
- Feng X, Lai Z, Lin Y, Lai G, Lian C (2015) Genome-wide identification and characterization of the superoxide dismutase gene family in *Musa acuminata* cv. Tianbaojiao (AAA group). BMC Genomics 16:823
- Feng K, Yu J, Cheng Y, Ruan M, Wang R, Ye Q, Zhou G, Li Z, Yao Z, Yang Y, Zheng Q, Wan H (2016) The SOD gene family in tomato: Identification, phylogenetic relationships, and expression patterns. Front Plant Sci 7:1279
- Foyer CH, Noctor G (2005) Redox homeostasis and antioxidant signaling: a metabolic interface between stress perception and physiological responses. Plant Cell 17(7):1866–1875
- Foyer CH, Noctor G (2011) Ascorbate and glutathione: the heart of the redox hub. Plant Physiol 155(1):2–18
- Frugoli JA, Zhong HH, Nuccio ML, McCourt P, McPeek MA, Thomas TL, McClung CR (1996) Catalase is encoded by a multigene family in *Arabidopsis thaliana* (L.) Heynh. Plant Physiol 112(1):327–336
- Giacomelli L, Masi A, Ripoll DR, Lee MJ, van Wijk KJ (2007) *Arabidopsis thaliana* deficient in two chloroplast ascorbate peroxidases shows accelerated light-induced necrosis when levels of cellular ascorbate are low. Plant Mol Biol 65(5):627–644
- Hare PD, Cress WA, Van Staden JV (1998) Dissecting the roles of osmolyte accumulation during stress. Plant Cell Environ 21(6):535–553
- Horemans N, Foyer CH, Asard H (2000) Transport and action of ascorbate at the plant plasma membrane. Trends Plant Sci 5(6):263–267
- Huber B, Eberl L, Feucht W, Polster J (2003) Influence of polyphenols on bacterial biofilm formation and quorum-sensing. Zeitschrift Fur Naturforschung. C. J Biosci 58(11–12):879–884
- Igamberdiev AU, Hill RD (2004) Nitrate, NO and haemoglobin in plant adaptation to hypoxia: An alternative to classic fermentation pathways. J Exp Bot 55(408):2473–2482

Ishikawa T, Shigeoka S (2008) Recent advances in ascorbate biosynthesis and the physiological significance of ascorbate peroxidase in photosynthesizing organisms. Biosci Biotechnol Biochem 72(5):1143–1154

- Kruk J, Holländer-Czytko H, Oettmeier W, Trebst A (2005) Tocopherol as singlet oxygen scavenger in photosystem II. J Plant Physiol 162(7):749–757
- Lattanzio V, Lattanzio VML, Cardinali A (2006) Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects. Phytochem: Adv Res 5:23–67
- Lazzarotto F, Teixeira FK, Rosa SB, Dunand C, Fernandes CL, de Vasconcelos Fontenele AV, Silveira JAG, Verli H, Margis R, Margis-Pinheiro M (2011) Ascorbate peroxidase related (APx-R) is a new heme-containing protein functionally associated with ascorbate peroxidase but evolutionarily divergent. New Phytol 191(1):234–250
- Ledford HK, Niyogi KK (2005) Singlet oxygen and photo-oxidative stress management in plants and algae. Plant Cell Environ 28(8):1037–1045
- Lerfall J (2016) Carotenoids: Occurrence, properties and determination. In: Caballero B, Finglas PM, Toldrá F (eds) Encyclopedia of food and health. Elsevier, pp 663–6669
- Lide DR (ed) (2006) CRC handbook of chemistry and physics, 87th edn. CRC Press Press
- Lu Y, Feng Z, Bian L, Xie H, Liang J (2010) miR398 regulation in rice of the responses to abiotic and biotic stresses depends on CSD1 and CSD2 expression. Funct Plant Biol 38(1):44–53
- McClung CR (1997) Regulation of catalases in Arabidopsis. Free Radic Biol Med 23(3):489–496 Menezes-Benavente L, Teixeira FK, Alvim Kamei CLA, Margis-Pinheiro M (2004) Salt stress induces altered expression of genes encoding antioxidant enzymes in seedlings of a Brazilian indica rice (*Oryza sativa* L.). Plant Sci 166(2):323–331
- Mhamdi A, Queval G, Chaouch S, Vanderauwera S, Van Breusegem FV, Noctor G (2010) Catalase function in plants: a focus on Arabidopsis mutants as stress-mimic models. J Exp Bot 61(15): 4197–4220
- Miller AF (2012) Superoxide dismutases: ancient enzymes and new insights. FEBS Lett 586(5): 585-595
- Millar AH, Mittova V, Kiddle G, Heazlewood JL, Bartoli CG, Theodoulou FL, Foyer CH (2003) Control of ascorbate synthesis by respiration and its implications for stress responses. Plant Physiol 133(2):443–447
- Mittler R (2002) Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci 7(9):405–410 Mittler R, Zilinskas BA (1991) Purification and characterization of pea cytosolic ascorbate peroxidase. Plant Physiol 97(3):962–968
- Moller IM (2001) Plant mitochondria and oxidative stress: electron transport, NADPH turnover, and metabolism of reactive oxygen species. Annu Rev Plant Physiol Plant Mol Biol 52:561–591
- Møller IM, Jensen PE, Hansson A (2007) Oxidative modifications to cellular components in plants. Annu Rev Plant Biol 58:459–481
- Ngamwonglumlert L, Devahastin S, Chiewchan N (2017) Natural colorants: pigment stability and extraction yield enhancement via utilization of appropriate pretreatment and extraction methods. Crit Rev Food Sci Nutr 57(15):3243–3259
- Noctor G (2006) Metabolic signalling in defence and stress: the central roles of soluble redox couples. Plant Cell Environ 29(3):409–425
- Noctor G, Foyer CH (1998) Ascorbate and glutathione: keeping active oxygen under control. Annu Rev Plant Physiol Plant Mol Biol 49:249–279
- Otles S, Cagindi O (2008) Carotenoids as natural colorants. In: Socaciu C (ed) Food colorants: chemical and functional properties. CRC Press Press, pp 51–70
- Panieri E, Santoro MM (2015) ROS signaling and redox biology in endothelial cells. Cell Mol Life Sci 72(17):3281–3303
- Pérez-Pérez ME, Lemaire SD, Crespo JL (2012) Reactive oxygen species and autophagy in plants and algae. Plant Physiol 160(1):156–164
- Pilon M, Ravet K, Tapken W (2011) The biogenesis and physiological function of chloroplast superoxide dismutases. Biochim Biophys Acta 1807(8):989–998

- Regelsberger G, Jakopitsch C, Plasser L, Schwaiger H, Furtmüller PG, Peschek GA, Zámocký M, Obinger C (2002) Occurrence and biochemistry of hydroperoxidases in oxygenic phototrophic prokaryotes (cyanobacteria). Plant Physiol Biochem 40(6–8):479–490
- Rice-Evans CA, Miller NJ, Paganga G (1997) Antioxidant properties of phenolic compounds. Trends Plant Sci 2(4):152–159
- Rosa SB, Caverzan A, Teixeira FK, Lazzarotto F, Silveira JAG, Ferreira-Silva SL, Abreu-Neto J, Margis R, Margis-Pinheiro M (2010) Cytosolic APx knockdown indicates an ambiguous redox responses in rice. Phytochemistry 71(5-6):548-558
- Scandalios JG (2002) The rise of ROS. Trends Biochem Sci 27(9):483-486
- Scandalios JG (2005) Oxidative stress: molecular perception and transduction of signals triggering antioxidant gene defenses. Braz J Med Biol Res 38(7):995–1014
- Schroeter H, Boyd C, Spencer JP, Williams RJ, Cadenas E, Rice-Evans C (2002) MAPK signaling in neurodegeneration: influences of flavonoids and of nitric oxide. Neurobiol Aging 23(5): 861–880
- Sharma P, Dubey RS (2005) Drought induces oxidative stress and enhances the activities of antioxidant enzymes in growing rice seedlings. Plant Growth Regul 46(3):209–221
- Sharma P, Jha AB, Dubey RS, Pessarakli M (2012) Reactive oxygen species, oxidative damage, and antioxidative defense mechanism in plants under stressful conditions. J Bot 2012:1–26
- Smirnoff N (2011) Vitamin C: the metabolism and functions of ascorbic acid in plants. Advances in botanical research. In: Rebeille F, Douce R (eds) Biosynthesis of vitamin S in plants: Vitamin S B6, B8, B9, C, E, K, vol 2, 1st edn. Academic Press, pp 107–177
- Srivastava V, Srivastava MK, Chibani K, Nilsson R, Rouhier N, Melzer M, Wingsle G (2009) Alternative splicing studies of the reactive oxygen species gene network in Populus reveal two isoforms of high-isoelectric-point superoxide dismutase. Plant Physiol 149(4):1848–1859
- Traber MG, Stevens JF (2011) Vitamins C and E: beneficial effects from a mechanistic perspective. Free Radic Biol Med 51(5):1000–1013
- Wu YS, Tang KX (2004) MAP kinase cascades responding to environmental stress in plants. Acta Bot Sin 46:127–136
- Zamocky M, Furtmüller PG, Obinger C (2008) Evolution of catalases from bacteria to humans. Antioxid Redox Signal 10(9):1527–1548