Current Clinical Oncology Series Editor: Maurie Markman

Amar Safdar Editor

Principles and Practice of Cancer Infectious Diseases

💥 Humana Press

Principles and Practice of Cancer Infectious Diseases

CURRENT CLINICAL ONCOLOGY

Maurie Markman, MD, Series Editor

For other titles published in this series, go to www.springer.com/series/7631

Amar Safdar Editor

Principles and Practice of Cancer Infectious Diseases

💥 Humana Press

Editor Amar Safdar Department of Infectious Diseases, Infection Control, and Employee Health The University of Texas M.D. Anderson Cancer Center Houston, TX USA amarsafdar@gmail.com

ISBN 978-1-60761-643-6 e-ISBN 978-1-60761-644-3 DOI 10.1007/978-1-60761-644-3 Springer New York Dordrecht Heidelberg London

Library of Congress Control Number: 2011928679

© Springer Science+Business Media, LLC 2011

All rights reserved. This work may not be translated or copied in whole or in part without the written permission of the publisher (Humana Press, c/o Springer Science+Business Media, LLC, 233 Spring Street, New York, NY 10013, USA), except for brief excerpts in connection with reviews or scholarly analysis. Use in connection with any form of information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed is forbidden.

The use in this publication of trade names, trademarks, service marks, and similar terms, even if they are not identified as such, is not to be taken as an expression of opinion as to whether or not they are subject to proprietary rights. While the advice and information in this book are believed to be true and accurate at the date of going to press, neither the authors nor the editors nor the publisher can accept any legal responsibility for any errors or omissions that may be made. The publisher makes no warranty, express or implied, with respect to the material contained herein.

Printed on acid-free paper

Humana Press is part of Springer Science+Business Media (www.springer.com)

This book is dedicated for promoting excellence in care and well-being for the patients with cancer.

Preface

Patients with cancer are highly susceptible to infections. These infections are inclined to be difficult to prevent, diagnose, and treat. There are a variety of reasons for this which will be discussed in detail in the chapters of this book. The intent for this book is to provide a comprehensive review of the ever changing spectrum of the management of infectious diseases in this complex population of patients. The changes in patient demography, near-constant global migration of contagious infections, emerging resistance to standard antimicrobial therapy, and the impact of expanding repertoire of antineoplastic therapies including the anticancer biologics and stem cell transplantation have influenced these changes. This book will provide a detailed guide for assessment of risk factors for various infections, evaluating prognosis among susceptible oncology patients with complex issues related to management of opportunistic infections. Strategies to promote hosts' immune response underscore the future measures based on perspicacious insight in the disease pathogenesis; interaction between the pathogen and host's immune function and inflammatory response are given prominent discussion throughout the book. I hope the reader will become acquainted with common and less often encountered infections and importantly, develop a keen knowledge of conditions that might be mistaken as infectious diseases in patients undergoing treatment for neoplastic diseases.

Houston, TX, USA

Amar Safdar, MD

Contents

1	Infections in Patients with Cancer: Overview Amar Safdar, Gerald Bodey, and Donald Armstrong	3			
2	Infections in Hematopoietic Stem Cell Transplant Recipients Georg Maschmeyer and Per Ljungman	17			
3	Infections in Patients with Hematologic Malignancies Genovefa Papanicolaou and Jayesh Mehta	27			
4	Infections in Solid Tumor Patients Alison G. Freifeld	39			
5	Infections in Patients with Hematologic Malignancies Treated with Monoclonal Antineoplastic Therapy André Goy and Susan O'Brien	47			
6	Postsurgery Infections in Cancer Patients Emilio Bouza, Almudena Burillo, Juan Carlos Lopez-Gutierrez, and José F. Tomás-Martinez	67			
7	Management of Infections in Critically Ill Cancer Patients Henry Masur	87			
Par	Part II Clinical Syndromes				
8	Management of the Neutropenic Patient with Fever Kenneth V.I. Rolston and Gerald P. Bodey	95			
9	Controversies in Empiric Therapy of Febrile Neutropenia John R. Wingard	105			
10	Catheter-Related Infections in Cancer Patients Iba Al Wohoush, Anne-Marie Chaftari, and Issam Raad	113			
11	Intravascular Device-Related Infections: Catheter Salvage Strategies and Prevention of Device-Related Infection Nasia Safdar and Dennis G. Maki	123			
12	Pneumonia in the Cancer Patient Scott E. Evans and Amar Safdar	143			

13	Noninfectious Lung Infiltrates That May Be Confused with Pneumonia in the Cancer Patient Rana Kaplan, Lara Bashoura, Vickie R. Shannon, Burton F. Dickey, and Diane E. Stover	153		
14	Mucosal Barrier Injury and Infections Nicole M.A. Blijlevens and J. Peter Donnelly	167		
15	Bacterial Colonization and Host Immunity Coralia N. Mihu, Karen J. Vigil, and Javier A. Adachi	175		
16	Neutropenic Enterocolitis and <i>Clostridium difficile</i> Infections Amar Safdar, Bruno P. Granwehr, Stephen A. Harold, and Herbert L. DuPont	181		
17	Management of Reactivation of Hepatitis B and Hepatitis C During Antineoplastic Therapy Marta Davila and Harrys A. Torres	189		
18	Management of Genitourinary Tract Infections Amar Safdar and Maurie Markman	195		
19	Central Nervous System Infections in Cancer Patients Victor Mulanovich and Amar Safdar	207		
20	Endocarditis in Oncology Patients Sara E. Cosgrove and Aruna Subramanian	219		
21	Skin Disorders Difficult to Distinguish from Infection Sharon Hymes, Susan Chon, and Ana Ciurea	233		
Part III Major Etiologic Agents				
22	Overview of Invasive Fungal Disease in Oncology Patients Amar Safdar	257		
23	Diagnosis of Invasive Fungal Disease Dionissios Neofytos and Kieren Marr	261		
24	Invasive Candidiasis in Management of Infections in Cancer Patients Matteo Bassetti, Malgorzata Mikulska, Juan Gea-Banacloche, and Claudio Viscoli	273		
25	Management of Aspergillosis, Zygomycosis, and Other Clinically Relevant Mold Infections Konstantinos Leventakos and Dimitrios P. Kontoyiannis	283		
26	Cryptococcal Disease and Endemic Mycosis Johan A. Maertens and Hélène Schoemans	293		
27	Current Controversies in the Treatment of Fungal Infections Christopher D. Pfeiffer, John R. Perfect, and Barbara D. Alexander	301		

x

28	Fungal Drug Resistance and Pharmacologic Considerationsof Dosing Newer Antifungal TherapiesRussell E. Lewis and David S. Perlin	317
29	Immunotherapy for Difficult-to-Treat Invasive Fungal Diseases Brahm H. Segal, Amar Safdar, and David A. Stevens	331
30	Cytomegalovirus in Patients with Cancer Morgan Hakki, Per Ljungman, and Michael Boeckh	341
31	Epstein-Barr Virus, Varicella Zoster Virus, and Human Herpes Viruses-6 and -8 Mini Kamboj and David M. Weinstock	359
32	Respiratory Viruses Roy F. Chemaly, Dhanesh B. Rathod, and Robert Couch	371
33	BK, JC, and Parvovirus Infections in Patients with Hematologic Malignancies Véronique Erard and Michael Boeckh	387
34	Antiviral Resistance and Implications for Prophylaxis Robin K. Avery	397
35	Management of Gram-Positive Bacterial Disease: Staphylococcus aureus, Streptococcal, Pneumococcal, and Enterococcal Infections Samuel Shelburne and Daniel M. Musher	409
36	Infections Caused by Aerobic and Anaerobic Gram-Negative Bacilli Kenneth V.I. Rolston, David E. Greenberg, and Amar Safdar	423
37	Listeriosis and Nocardiosis Heather E. Clauss and Bennett Lorber	435
38	Antibacterial Distribution and Drug–Drug Interactions in Cancer Patients Ursula Theuretzbacher and Markus Zeitlinger	443
39	<i>Mycobacterium tuberculous</i> Infection Michael Glickman	455
40	Nontuberculous Mycobacterial Infections Amar Safdar	463
41	Parasitic Infections in Cancer Patients: Toxoplasmosis, Strongyloidiasis, and Other Parasites Brian G. Blackburn and José G. Montoya	469
42	Zoonoses in Cancer Patients Donald Armstrong	481

Part IV Management of Antimicrobial Therapy

43	Antimicrobial Stewardship: Considerations for a Cancer Center Coralia N. Mihu, Alla Paskovaty, and Susan K. Seo	491
44	Controversies in Antimicrobial Stewardship Graeme N. Forrest	499
45	Prevention of Antimicrobial Resistance: Current and Future Strategies Cesar A. Arias and Adolf W. Karchmer	507
Par	rt V Infection Prevention: Antimicorbial Prophylaxis and Immunization	
46	Antibacterial, Antifungal, and Antiviral Prophylaxis in High-Risk Cancer and Stem Cell Transplant Population Marcio Nucci and John R. Wingard	521
47	Controversies in Antimicrobial Prophylaxis Ben de Pauw and Marta Stanzani	533
48	Infection Prevention – Protected Environment and Infection Control J. Peter Donnelly	541
49	Prevention of Tropical and Parasitic Infections: The Immunocompromised Traveler Francesca F. Norman and Rogelio López-Vélez	551
50	Prophylactic Vaccination of Cancer Patients and Hematopoietic Stem Cell Transplant Recipients William Decker and Amar Safdar	561
Err	atum	E1
Ind	ex	573

Contributors

Javier A. Adachi, M.D.

Department of Infectious Diseases, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Barbara D. Alexander, M.D.

Department of Medicine, Division of Infectious Diseases and International Health, Duke University Medical Center, Durham, NC, USA

Iba Al Wohoush, M.D.

Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, Texas, USA

Cesar A. Arias, M.D.

Department of Internal Medicine, Division of Infectious Diseases, University of Texas Medical School at Houston, TX, USA

Donald Armstrong, M.D.

Department of Medicine, Infectious Disease Service, Memorial Sloan–Kettering Cancer Center, New York, NY, USA

Robin K. Avery, M.D.

Department of Infectious Disease, Cleveland Clinic Foundation, Lerner College of Medicine of Case Western Reserve University, Cleveland, OH, USA

Lara Bashoura, M.D.

Department of Pulmonary Medicine, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Matteo Bassetti, M.D., Ph.D.

Division of Infectious Diseases, San Martino Hospital and University of Genoa, Genoa, Italy

Brian G. Blackburn, M.D.

Department of Internal Medicine, Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, CA, USA

Nicole M. A. Blijlevens, M.D.

Department of Haematology, Radboud University Nijmegen Medical Centre & Nijmegen University Centre for Infectious Diseases, Nijmegen, The Netherlands

Gerald P. Bodey, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Michael Boeckh, M.D.

Vaccine and Infectious Disease Division, University of Washington, Fred Hutchinson Cancer Research Center, Seattle, WA, USA

Emilio Bouza, M.D.

Clinical Microbiology and Infectious Diseases Department, Hospital General Universitario Gregorio Marañón, Universidad Complutense de Madrid; CIBER de Enfermedades Respiratorias (CIBERES), Madrid, Spain

Almudena Burrillo, M.D., Ph.D.

Clinical Microbiology Department, Hospital Universitario de Móstoles, Madrid, Spain

Anne-Marie Chaftari, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Roy F. Chemaly, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Susan Chon, M.D.

Department of Dermatology, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Ana Ciurea, M.D.

Department of Dermatology, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Heather E. Clauss, M.D.

Department of Infectious Diseases, Temple University Hospital, Philadelphia, PA, USA

Sara E. Cosgrove, M.D.

Division of Infectious Diseases, Johns Hopkins University School of Medicine, Baltimore, MD, USA

Robert Couch, M.D.

Department of Molecular Virology and Microbiology, Baylor College of Medicine, Center for Infection and Immunity Research, Houston, TX, USA

Marta Davila, M.D.

Department of Gastroenterology, Hepatology, and Nutrition, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

William Decker, Ph.D.

Department of Blood and Marrow Transplantation, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Burton F. Dickey, M.D.

Department of Pulmonary Medicine, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

J. Peter Donnelly, Ph.D.

Department of Haematology and Nijmegen Institute for Infection, Inflammation and Immunity, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands

Herbert L. DuPont, M.D.

Department of Medicine, The University of Texas, School of Public Health, Center for Infectious Diseases; Department of Internal Medicine, St. Luke's Episcopal Hospital; Department of Microbiology and Immunology, Baylor College of Medicine, Houston, TX, USA

Véronique Erard, M.D. Médecin Adjointe, Infectiologie, HFR-Fribourg, Switzerland

Scott E. Evans, M.D.

Department of Pulmonary Medicine, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Graeme N. Forrest, M.D.

Division of Infectious Disease, Portland VA Medical Center, Portland, OR, USA

Alison G. Freifeld, M.D. Department of Medicine, University of Nebraska Medical Center, Omaha, NE. USA

Juan Gea-Banacloche, M.D.

Experimental Transplantation and Immunology Branch, National Cancer Institute, Bethesda, MD, USA

Michael Glickman, M.D.

Department of Medicine, Infectious Disease Service, Memorial Sloan Kettering Cancer Center, New York, NY, USA

Andre Goy, M.D. Hematology/Oncology, Internal Medicine, Hackensack University Medical Center, Hackensack, NJ, USA

Bruno P. Granwehr, M.D.

Department of Infectious Diseases, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

David E. Greenberg, M.D.

Department of Infectious Diseases, National Institute of Allergy and Infectious Diseases, Bethesda, MD, USA

Morgan Hakki, M.D.

Division of Infectious Diseases, Oregon Health & Science University, Portland, OR, USA

Stephen A. Harold

Department of Medicine, The University of Texas, School of Public Health, Center for Infectious Diseases, Houston, TX, USA

Sharone Hymes, M.D.

Department of Dermatology, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Mini Kamboj, M.D.

Department of Medicine, Infectious Disease Service, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

Rana Kaplan, M.D.

Department of Medicine, Pulmonary Medicine Service, Memorial Sloan–Kettering Cancer Center, New York, NY, USA

Adolf W. Karchmer, M.D.

Division of Infectious Diseases, Beth Israel Deaconess Medical Center, Boston, MA, USA

Dimitrios P. Kontoyiannis, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Konstantinos Leventakos, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Russell E. Lewis, Pharm. D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Per Ljungman, M.D.

Hematology Center, Karolinska University, Stockholm, Sweden

Juan Carlos Lopez-Gutierrez, M.D.

Department of Pediatric Surgery, Hospital Universitario Lu Paz, Universidad Autonoma de Madrid, Spain

Rogelio López-Vélez, M.D.

Tropical Medicine and Clinical Parasitology Unit, Department of Infectious Diseases, Ramón y Cajal Hospital, Instituto Ramón y Cajal de Investigación Sanitaria (IRYCIS), Madrid, Spain

Bennett Lorber, M.D.

Department of Medicine, Section of Infectious Diseases, Temple University School of Medicine, Philadelphia, PA, USA

Johan A. Maertens, M.D., Ph.D.

Department of Hematology, Acute Leukemia and Stem Cell Transplantation Unit, University Hospitals Leuven, Leuven, Belgium

Dennis G. Maki, M.D.

Department of Medicine, University of Wisconsin Hospital and Clinics, Madison, WI, USA

Maurie Markman, M.D.

Department of Gynecologic Oncology, The University of Texas M.D. Anderson Cancer Center, Houston, Texas, USA

Kieren Marr, M.D. Division of Infectious Diseases, The Johns Hopkins Hospital, Baltimore, MD, USA

Georg Maschmeyer, M.D.

Department of Hematology, Oncology and Palliative Care, Klinikum Ernst von Bergmann, Potsdam, Germany

Henry Masur, M.D.

Critical Care Medicine Department, Clinical Center, National Institutes of Health, Bethesda, MD, USA

Jayesh Mehta, M.D.

Hematopoietic Stem Cell Transplant Program, Robert H. Lurie Comprehensive Cancer Center, Northwestern University Medical Center, Chicago, IL, USA

Coralia N. Mihu, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Malgorzata Mikulska, M.D.

Division of Infectious Diseases, San Martino Hospital and University of Genoa, Genoa, Italy

José G. Montoya, M.D.

Department of Internal Medicine, Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, CA, USA; *Toxoplasma* Serology Laboratory, Palo Alto Medical Foundation, Palo Alto, CA, USA

Victor Mulanovich, M.D.

Infectious Diseases Department, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Daniel M. Musher, M.D.

Departments of Medicine, Microbiology and Immunology, Baylor College of Medicine, Infectious Diseases Section, Veterans Affairs Medical Center, Houston, TX, USA

Dionissios Neofytos, M.D.

Division of Infectious Diseases, The Johns Hopkins Hospital, Baltimore, MD, USA

Francesca F. Norman, M.D.

Tropical Medicine and Clinical Parasitology Unit, Department of Infectious Diseases, Ramón y Cajal Hospital, Madrid, Spain

Marcio Nucci, M.D.

Department of Internal Medicine, Hematology Unit Head, Mycology Laboratory, Hospital Universitário Clementino Fraga Filho – Federal University of Rio de Janeiro, Rio de Janeiro, Brazil

Susan O'Brien, M.D.

Department of Leukemia, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Genovefa Papanicolaou, M.D.

Infectious Diseases Service, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

Alla Paskovaty, Pharm.D.

Infectious Diseases Service, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

Ben de Pauw, M.D.

Institute of Haematology and Clinical Oncology "Lorenzo e Ariosto Seràgnoli", Sant'Orsola-Malpighi Hospital, University of Bologna, Bologna, Italy

John R. Perfect, M.D.

Department of Medicine, Division of Infectious Diseases and International Health, Duke University Medical Center, Durham, NC, USA

David S. Perlin, Ph.D.

Department of Clinical Sciences and Administration, College of Pharmacy, University of Houston, Texas Medical Center Campus, Houston, TX, USA; Department of Infectious Disease, Infection Control, and Employee Health, The University of Texas/M.D. Anderson Cancer Center, Houston, TX, USA

Christopher D. Pfeiffer, M.D.

Department of Medicine, Division of Infectious Diseases and International Health, Duke University Medical Center, Durham, NC, USA

Issam Raad, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Dhanesh B. Rathod, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Kenneth V. I. Rolston, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Amar Safdar, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Nasia Safdar, M.D.

Section of Infectious Diseases, Department of Medicine, University of Wisconsin Medical School, Madison, WI, USA

Hélène Schoemans, M.D.

Department of Hematology, Acute Leukemia and Stem Cell Transplantation Unit, University Hospitals Leuven, Leuven, Belgium

Brahm H. Segal, M.D.

Department of Medicine and Immunology, Roswell Park Cancer Institute, Department of Medicine, School of Medicine and Biomedical Sciences, University of Buffalo, Elm & Carlton Streets, Buffalo, NY, USA

Susan K. Seo, M.D.

Infectious Diseases Service, Memorial Sloan-Kettering Cancer Center, New York, NY, USA

Vickie R. Shannon, M.D.

Department of Pulmonary Medicine, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Samuel Shelburne, M.D.

Department of Infectious Diseases, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Marta Stanzani, M.D.

Institute of Haematology and Clinical Oncology "Lorenzo e Ariosto Seràgnoli", Sant' Orsola-Malpighi Hospital, University of Bologna, Bologna, Italy

David A. Stevens, M.D.

Department of Medicine, Stanford University School of Medicine Division of Infectious Diseases, Santa Clara Valley Medical Center, Saratoga, CA, USA

Diane E. Stover, M.D.

Department of Medicine, Pulmonary Medicine Service, Memorial Sloan-Kettering, New York, NY, USA

Aruna Subramanian, M.D.

Division of Infectious Diseases, Johns Hopkins University School of Medicine, Baltimore, MD, USA

Ursula Theuretzbacher, Ph.D. Center for Anti-Infective Agents, Vienna, Austria

José Francisco Tomaś-Martinez, M.D. Department of Hematology, The University of Texas M.D. Anderson International España, Madrid, Spain

Harrys A. Torres, M.D.

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, Houston, TX, USA

Karen J. Vigil, M.D. University of Texas Health Science Center, Houston, TX, USA

Claudio Viscoli, M.D. Division of Infectious Diseases, San Martino Hospital and University of Genoa, Genoa, Italy

David M. Weinstock, M.D. Department of Medical Oncology, Dana-Farber Cancer Institute, Boston, MA, USA

John R. Wingard, M.D. Department of Medicine, University of Florida, Gainesville, FL, USA

Markus Zeitlinger, M.D. Department of Clinical Pharmacology, Medical University of Vienna, Vienna, Austria

Part I Overview and Special Population

Chapter 1 Infections in Patients with Cancer: Overview

Amar Safdar, Gerald Bodey, and Donald Armstrong

Abstract Patients with neoplastic disease are often highly susceptible to severe infections. The following factors influence the types, severity, and response to therapy of these infections: (1) Changing epidemiology of infections; (2) cancerand/or treatment-associated neutropenia; (3) acquired immune deficiency states such as cellular immune defect; (4) recent development of new-generation diagnostic tools including widely available DNA amplification tests; (5) effective intervention for infection prevention; (6) empiric or presumptive therapy during high-risk periods; (7) availability of new classes of highly active antimicrobial drugs; (8) strategies to promote hosts' immune response; and (9) future measures. This introductory chapter intended for the reader to become familiar with the important historical milestones in the understanding and development in the field of infectious diseases in immunosuppressed patients with an underlying neoplasms and patients undergoing hematopoietic stem cell transplantation.

Keywords Cancer • Infection • Neutropenia • Immune defects • Diagnosis • Therapy

Patients with neoplastic disease are often highly susceptible to severe infections. These are inclined to be difficult to prevent, diagnose, and treat. There are a variety of reasons for this which will be discussed in detail in the chapters of this book. We will introduce this volume by reviewing the history and background of such infections, where we believe major advances have been made and what we believe will be necessary to effectively prevent and manage such infections in the future. The following factors influence the types, severity, and response to therapy of these infections: (1) Changing epidemiology of infections; (2) cancer- and/or treatmentassociated neutropenia; (3) acquired immune deficiency states such as cellular immune defect; (4) recent development of new-generation diagnostic tools including widely available DNA amplification tests; (5) effective intervention for infection prevention; (6) empiric or presumptive therapy during high-risk periods; (7) availability of new classes of highly active antimicrobial drugs; (8) strategies to promote hosts' immune response; and (9) future measures.

Historical Perspective

The introduction of chemotherapeutic regimens has expanded the population at risk, since many of these agents affect host defenses, most often causing neutropenia. However, even in acute leukemia, the malignancy with the highest frequency of infection, very little was published about infectious complications until the second half of the twentieth century. The paucity of published data is illustrated by a book on acute leukemia, published in 1958, which made no mention of infectious complications [1]. Indeed, at that time, some physicians attributed fevers in leukemia patients to a general hypermetabolic condition caused by the neoplasm.

By the 1950s, several antineoplastic agents became available which caused at least transient improvement in some malignant diseases. Nitrogen mustard caused responses in Hodgkin disease, aminopterin caused responses in acute leukemia, and methotrexate cured choriocarcinoma in women. The subsequent use of multiple drug combinations in acute lymphocytic leukemia and Hodgkin disease represented major advances [2]. Another important advance was the use of platelet transfusions to control and prevent hemorrhage in acute leukemia patients with thrombocytopenia [3]. In an autopsy study, the frequency of hemorrhage as a cause of death in acute leukemia patients decreased from 67 to 37% due to the use of platelet transfusions [4]. Unfortunately, infection remained a major cause of death. There have been many reviews of the subjects over the years, some with international contributors and continuity which are references here [5–11].

A. Safdar (🖂)

Department of Infectious Diseases, Infection Control, and Employee Health, The University of Texas M.D. Anderson Cancer Center, 1515 Holcombe Boulevard, Houston, TX 77030, USA e-mail: asafdar@mdanderson.org

Epidemiological Factors

Exposures to organisms in the distant as well as recent past should be considered in patients with neoplastic disease. Latent infections may be activated in the presence of waning immunity whether it be due to the disease itself or to the treatment. The classic example of this is reactivation of latent tuberculous in patients with treatment-induced helper T-cell dysfunction. Additional latent infections which may be activated, for example, are histoplasmosis, coccidiomycosis, disease caused by the Herpes group of viruses, toxoplasmosis, strongyloidiasis, and others. These demand consideration and many such as TB, herpes simplex, and strongyloidiasis can be effectively treated prophylactically. Recent travel or residence and hospitalization may expose patients to organisms which may incubate such as malaria after travel to an endemic area or colonization due to drug-resistant bacteria such as Klebsiella, Pseudomonas, and Stenotrophomonas species acquired during a previous hospitalization. Questions to investigate epidemiologic factors should include exposures at home along with work, habits, and hobbies. Also, a detailed history of recent and remote travel and recreational activities may provide clues for an otherwise improbable diagnosis. All of these can be a source of infection, some of which can be avoided with appropriate patient education.

Hosts' Susceptibility

It is not surprising that the frequency of infection is related to the type of underlying malignancy and most infections occur in patients who are failing to respond to their cancer therapy. Surveys in the 1960s found that about 80% of patients with acute leukemia, 75% with lymphoma, but less than 40% of patients with metastatic carcinoma developed infection [12, 13]. There are a wide variety of factors that may impact on the susceptibility of cancer patients to infection [11]. Local factors such as tumor masses that may obstruct the bronchial tree or urinary tract and necrotic tumors in the gastrointestinal tract can result in infection. In an autopsy study of children with metastatic carcinoma, 80% of cases of pneumonia were associated with pulmonary metastases, aspiration, or tracheostomy [14]. Antibiotic therapy is often of limited efficacy in these types of tumors, unless the local predisposing factor can be removed.

Immunological Factors

Neutropenia is the most important predisposing factor and can be due to the disease or its therapy. While there were some reports of the role of neutropenia in infection, a detailed

analysis of 52 patients with acute leukemia was published in 1966 [15]. This study demonstrated that the risk of infection was related to the degree and duration of neutropenia. The risk increased when the neutrophil count was less than 1,000/ mm³, but increased substantially when it was below 500/ mm³. Also, the risk of developing infection increased the longer the duration of neutropenia. One hundred percent of episodes of severe neutropenia (<100 PMN/mL) lasting 3 weeks or longer were accompanied by identifiable infection compared to 65% of episodes lasting one week. Neutropenia diminishes the likelihood of detecting characteristic manifestations of infection. One study compared physical findings of infection in a group of patients with severe neutropenia with a group with adequate neutrophil counts [16]. Only 8% of patients in the former group with pneumonia were able to produce purulent sputum compared to 84% in the latter group. Similarly, among patients with urinary tract infections, pyuria was found in 11 and 97%, respectively. In an autopsy study, it was demonstrated that many pulmonary infections were not detected on routine chest radiographs antemortem [17]. Likewise, among patients with gram-negative bacillary pneumonia, 85% of those with initially abnormal chest radiographs had >1,000 neutrophils/mL, whereas 81% with normal roentgenograms had <1,000 neutrophils/mL [18]. The lack of signs of infection in febrile neutropenic patients impairs the physician's ability to determine whether or not fever is due to infection. In one study of fever in neutropenic patients, physicians were required to conclude whether infection was present or not before instituting therapy [19]. The physician's initial diagnosis (infection or fever of unknown origin) was incorrect in 33% of the cases.

White blood cell (WBC) transfusions were initiated in an effort to improve the outcome of infections in severely neutropenic patients. Since it was difficult to collect sufficient neutrophils from normal donors, initially, patients with chronic myelogenous leukemia with high neutrophil counts were used as donors [20]. Later, the development of the continuous cell separating machine made it possible to collect adequate cells from normal donors [21]. Studies demonstrated that there was a direct relationship between the number of cells transfused and the increment in the recipient's neutrophil count. In one study of 128 neutropenic patients who had fever unresponsive to antibiotic therapy, 49% responded after WBC transfusions, including patients with pneumonia and gram-negative bacillary septicemia [22]. Unfortunately, potential adverse effects occurred in some recipients. In one study when WBC transfusions were administered with amphotericin B, 64% of patients developed acute dyspnea, respiratory deterioration, and new pulmonary infiltrates compared to only 6% of patients who did not receive amphotericin B [23]. Several other studies failed to observe this toxicity. Another potential adverse event primarily for bone marrow transplant recipients was

acquisition of cytomegalovirus (CMV) infection [24]. Reports of graft-versus-host disease (GVHD) in a few recipients has led to routine irradiation transfused cells, but questions have been raised about adverse effects of radiation on the function of the transfused neutrophils. In a review of seven prospective randomized trials of WBC transfusions in neutropenic patients with infection, it was concluded that the transfusions were of some benefit in five studies but the number of patients in each study was small [25]. A problem with many was the ignoring of the number of neutrophils administered; hence, some patients received an inadequate dose. The use of WBC transfusions diminished by the 1980s because there was inadequate evidence of their efficacy from prospective comparative studies. However, there has been a resurgence of interest in increasing available neutrophils since recombinant myeloid growth factor granulocyte-colony-stimulating factor (G-CSF) has become available. Administration of G-CSF to donors improves the number of neutrophils collected as well as increases their activity against infection [26].

Protected Environment. Because of the risk of infection during periods of chemotherapy-induced neutropenia, efforts were made to provide a sterile environment for these patients. The first type of unit was a bed surrounded by a plastic canopy with filtered air (Fig. 1.1). Later, laminar air flow rooms were designed [27]. These units provided filtered air, sterile water supply, sterile room, specially prepared food, and toilet facilities. The patients were given specifically prepared "sterile" food and prophylactic oral and topical antibiotics. These rooms, air, food, and patients were carefully monitored for microbial contamination [28, 29]. The program reduced the frequency of infection and permitted the use of more

intensive chemotherapy in the premyeloid growth factor era. Unfortunately, more intensive chemotherapy in this setting did not result in higher remission rates for several malignancies including acute leukemia [30], lymphoma [31], and sarcoma [32]. One review of protected environment entitled "Protected Environment are discomforting and expensive and do not offer meaningful protection" summarized the discussion as follows "The one constant in almost every controlled study is that life has not been prolonged, remission induction increased, nor remission prolonged" [8].

In the late 1940s and early 1950s, patients with neoplasms were originally found to be infected with organisms from the flora in their nasopharynx and the gastrointestinal tract due to neutropenia caused by their disease or subsequent therapy. Exceptions were those with cellular immune defects due to the neoplasm such as Hodgkin's disease, who might present with cryptococcosis or those with multiple myeloma who might present with pneumococcal septicemia because of their decreased production of normal immune globulins. In the neutropenic patient, the organisms invading from the nasopharynx were usually Streptococcus pyogenes or Staphylococcus aureus (penicillin susceptible). From the orointestinal tract, Escherica coli and Klebsiella or Proteus species were responsible; these bacteria were sensitive to most available antibiotics during early 1950s. Gradually, but steadily, resistance developed in most of the organisms except S. pyogenes. S. aureus resistant to penicillin and Pseudomonas aeruginosa resistant to all antimicrobials except polymyxin appeared in the late 1950s [4, 33, 34]. Antimicrobial resistance developed over the years among the orointestinal isolates and the Gram-positive cocci increased to become predominate by the 1980s with MRSA and penicillin-resistant



Fig. 1.1 First type of protective environment for severely neutropenic patients. Note, sleeves in the side of canopy to perform tasks on patient and chambers at the foot that irradiated items placed into unit

alpha streptococci appearing. Many of the effective anticancer treatment regimens result in neutropenia so that these types of infection remain a major problem in patients with neoplastic disease.

In contrast, patients with cellular immune defects due to their basic disease or its therapy are prey to a different array of organisms. Predisposing diseases include Hodgkin's disease, T lymphocyte lymphomas and leukemias, and hairy cell leukemia. Various transplantation procedures and GVHD along with treatments for them including cyclosporine, antithymocyte globulin, tacrolimus and adrenocorticosteroids induce defects which result in such opportunistic infections. The diseases are due to organisms from all categories including Salmonella spp., Histoplasma capsulatum, Leishmania spp., and CMV. In the early 1980s and with the advent of the AIDS epidemic, investigators with access to laboratories where T cells could be measured began systematic studies that revealed that patients with levels in the 200 range or lower would develop one or more of these opportunistic infections, especially PCP. It became apparent that as the T cells fell, it could be predicted which organisms would cause disease [8, 35]. Now with the measurements of endogenous cytokines, T-cell subset populations, and functional analysis, this is even more predictable and offers opportunities for treatment and prevention.

B-cell defects have been well described occurring in certain groups of patients with certain underlying neoplastic diseases such as multiple myeloma and chronic lymphocytic leukemia or those after bone marrow transplantation. In these instances, the organisms to be anticipated are *Streptococcus pneumoniae*, *Hemophilus influenzae*, *Neiseriae meningitimus*, or late after transplantation, Echoviruses. Vaccine studies in this group of patients and others are underway to try to achieve protection.

An altered integument allows access to a large variety of organisms to invade patients with neoplastic disease. Areas at risk include the entire orointestinal tract where chemotherapy-induced mucusitis with ulcers allow organisms' entry into tissues and the bloodstream. Intravascular catheters allow direct entry into the bloodstream and other catheters such as bladder, intraperitoneal or intracranial devices are sources of infection especially in the neutropenic patient. In addition, life-threatening infections may result from infusion of blood products or transplanted organs. These may vary from HIV and HTLV-I [36] to *Salmonella* spp., *Candida* spp., and *Trypanasoma cruzi* among others.

Knowing the immunological defect in a patient with neoplastic disease suspected of having an infection is extremely important. From the clinical picture, the appropriate tests can be done to confirm the diagnosis, and if indicated, empiric therapy can be started. A fine example of this is the empiric therapy of the neutropenic patient with appropriate antibiotics for anticipated organisms in the clinical setting such as a particular hospital. In the early 1960s, a clinical study from the NCI documented the association of the fall of the neutrophil count with the rise of the severe infections [15]. An example of a population at risk for a specific infection due to an immune defect was the prevention of Pneumocystis pneumonia in children with acute lymphoblastic leukemia carried out at St. Judes Hospital in Memphis TN [37]. Almost 100% protection was achieved. Knowledge of the perturbations in immune function following bone marrow transplantation has enabled clinicians to use preemptive therapy for suspected infections such as those caused by CMV.

Finally, immune defects involving innate and adaptive immune responses may occur in patients who have received prolonged courses of chemotherapy, neoadjuvant antineoplastic monoclonal antibody therapy, or immunosuppressive agents for treatment of GVHD following allogeneic stem cell transplantation.

Diagnostic Evaluation of Infection

There have been remarkable advances in diagnostic tests for the evaluation of infection in the past five decades, especially in diagnostic microbiology allowing us to make earlier and more specific microbial diagnoses. Gram stains, invented in 1884 by Hans Christian Gram in Denmark, and variations on dye techniques are still routine and useful for early presumptive diagnoses, but immunological methods using direct fluorescent antibody stains have been developed and are regularly used especially for viruses. In unusual circumstances such as suspected polyoma virus infection, electron microscopy may be used. New culture methods include isolator lysis centrifugation tubes which are used for continuous around-the-clock monitoring employing a fluorescent carbon dioxide detection system. An automated broth system can be used for quantitation by colony counts of centrifuged sediments and these systems are more sensitive for the isolation of some fungi, mycobacteria and Bartonella species. In addition, the broth can be examined by nucleic acid probes and HPLC for rapid organism identification. Automated broth Minimum Inhibitory Concentration (MIC) antimicrobial susceptibility tests yield more rapid results which can be entered into online computer systems for clinicians and recorded for antimicrobial susceptibility patterns for hospital infection control. To help select antimicrobial regimens for empiric therapy, these data can also be available for local and national Health Departments as well as the hospital.

Polymerase chain reaction (PCR) techniques to recognize copies of nucleic acid fragments in various specimens have been developed and are being used. Many are undergoing FDA approval and some may be available only in special laboratories. These techniques may well replace earlier tests using antigen detection by poly or monoclonal antibodies and chemical tests for specific cellular elements such as arabinatol, beta D-glucan, or galactomanans of fungi.

Antibody tests are much easier to perform since the enzyme-linked antibody (ELISA) test has replaced the compliment fixation (CF) test, and for specificity, the Western blot has become the "gold standard". However, for cancer patients and those following allogeneic stem cell transplantation, serologic diagnosis may provide limited information regarding active versus remotely acquired disease. Furthermore, a negative serology cannot be interpreted with certainty due to potential defects in B-cell function.

Radiologic testing with CT scans and MRIs has better defined anatomic lesions for presumptive diagnoses, and recent advances in safe tissue sampling can be used by interventional radiology techniques for specific diagnoses. Bronchoalveolar lavages have virtually replaced open lung biopsies for investigating pulmonary lesions; however, similar to diagnostic reliability of serologic diagnosis, a negative BAL sample smear or culture dose not exclude the possibility of opportunistic lung infection. Radioactive labeling of the patient's own neutrophils and injecting them for localizing foci of infection can sometimes be helpful as can technetium scans. Efforts to localize infected sites using antibody for specific organisms are presently under study and this method could also offer treatment opportunities. Similarly, PET scan are now commonly used for tumor burden and disease recurrence monitoring; this new technology appears promising as an adjuvant diagnostic tool.

Pathogens of Interest

Most infections occurring in patients with nonhematological malignancies are caused by organisms commonly associated with the site of the tumor or nosocomial pathogens except when on chemotherapy. Infections in patients with hematological malignancies are usually caused by organisms that are prevalent in association with specific deficiencies in host defense mechanisms or are due to nosocomial pathogens. Only a few examples will be presented in this discussion, primarily focused on those infections prevalent in neutropenic patients.

Bacterial Infections

Early studies of infection in patients receiving chemotherapy for hematological malignancies found that *S. aureus* developed resistance to penicillin. It became the predominant cause of fatal infection in neutropenic patients [4]. Once effective antibiotics became available for treatment of penicillinresistant S. aureus, gram-negative bacilli emerged as the most common cause of fatal infections. Pseudonomas aeruginosa became a major cause of infections, especially among neutropenic patients [29, 37, 38]. Although polymyxin B and colistin were very active in vitro against the pathogen, they were ineffective for therapy in neutropenic patients and were of limited benefit in other patients. Their efficacy in neutropenic patients depended upon the recovery from neutropenia. The availability of carbenicillin, the first β lactam with anti-pseudomonal activity, had a dramatic impact on the therapy of life-threatening Pseudomonas infections [39]. Other gram-negative bacilli emerged as significant pathogens, including Klebiella spp. and Serratia marcescens. Cephalothins were the first β lactam available for the treatment of some of these infections [40]. Over the years, multiplicity of antibiotics has been developed including potent broad-spectrum cephalosprosins, carbapenims, and fluoroquinolones [41]. Despite these important advances, bacterial infections remain a serious threat to cancer patients, due in large part to the ability of organisms to develop resistance to multiple antibiotics. Recent increase in nonpseudomonal nonfermentative Gram-negative bacteria such as Stenotrophomonas maltophilia has been associated with difficult-to-treat healthcare-associated infections; these bacteria may also cause less severe community-acquired infections [42]; high-dose trimethoprim-sulfamethoxazole remains the treatment of choice, although occasionally a multidrug-resistant organism poses a serious challenge [43]. Emergence and spread of extended-spectrum beta-lactamases (ESBL) Enterobacteriaceae and recently identified carbapenemases producing *Klebsiella* species (KPC) and spreading to other gram-negative disease-associated bacteria herald alarming limitation in choice for effective antimicrobial therapy against these new groups of MDR-gram-negative bacterial infections [44].

Listeria monocytogenes was one of the first bacterial infections reported as occurring more frequently in patients with cellular immune defects [8, 45] and it continues to be a problem [46]. It soon became apparent that *Salmonella* spp., *Nocardia asteroids*, and *Rhodococcus equi* were also opportunistic bacterial pathogens in this setting. *Mycobacterium hemphilum* [47] was thereafter established as a *Mycobacterium to be anticipated in T-cell-deficient patients, in addition to the classic example of M. tuberculous* [48] and subsequently *M. avium-intracellulare* complex.

Principles of Antibiotic Therapy in Neutropenic Patient

This discussion will be limited to general principles. Discussion of specific antibiotic therapies is presented in other chapters of this book. After multiple antibiotics became available and the potential for emergence of resistance became apparent, it became the standard practice to withhold antibiotic therapy in the febrile patient until the infecting pathogen was identified. However, early studies of antibiotic therapy for fever in neutropenic patients clearly indicated the importance of instituting antibiotic therapy promptly to neutropenic patients when they become febrile. It has been demonstrated that mortality rates increase substantially if therapy is not administered promptly. The choice of initial antibiotic therapy should provide broad-spectrum antibacterial coverage against gram-positive cocci and gram-negative bacilli. Most infections are caused by aerobic gram-negative bacilli and anaerobic infections tend to be uncommon. It is of critical importance for physicians caring for neutropenic patients to be aware of the common pathogens causing infections at their hospitals and their antimicrobial susceptibilities so that appropriate antibiotic regimens will be selected. Antibiotics that are bactericidal should be selected when possible. The greatest experience has been obtained with broad-spectrum β lactams and aminoglycosides. Aminoglycosides are less effective as single agents in neutropenic patients and should not be used alone [49].

Some studies have indicated that synergistic combinations that provide high serum cidal levels such as a β lactam plus an aminoglycoside are more effective than single agents [50]. However, aminoglycosides have potential nephrotoxicity, which are more prevalent in the elderly and patients with cancer such as multiple myeloma or cancer therapy induced reduced renal reserves.

Various regional, national, and international groups have met and are still meeting to study questions of treatment and how to conduct studies to evaluate treatment of bacterial infections. These have included The Infectious Diseases Society of America [51], The European Organization for Research and Treatment of Cancer [52], and The Immunocompromised Host Society [53]. For empirical antibacterial treatment, it is evident that regimens should be aimed at the most prevalent organisms with reliable knowledge of their susceptibility infecting the patient at a given hospital. It must be stressed that continued efforts at prevention, e.g., scrupulous hygiene, are most important.

Patients with fever of unknown origin that persists after several days of broad-spectrum of antibiotic therapy represent a difficult problem. Careful reevaluation and collection of additional appropriate diagnostic tests need to be performed and additional therapeutic measures should be considered. These may include other antibacterial, antifungal, or antiviral agents. Antifungal agents should be given serious consideration in these patients. Some investigators have advocated that antibiotic therapy be continued in patients with documented infections until the neutrophil count recovers. There is considerable evidence to indicate that this is unnecessary and can encourage superinfection. A more appropriate approach is to discontinue the therapeutic agents, watch carefully.

Mycobacterial Infections

Tuberculous is a well-recognized, albeit uncommon, complication even in patients with severe cellular immune defect [48]. Patients with solid organ cancer may be as susceptible to active *Mycobacterium tuberculous* infection as patients with hematologic malignancy and those undergoing hematopoietic stem cell transplantation [54]. It remains important to realize that tuberculous, being an indolent disease, may be mistaken for a slowly progressing neoplasm and may lead to unnecessary large excisions that can be avoided by initial fine needle aspiration and biopsy of the suspected mass [55].

Nontuberculous mycobacterial disease due to slowgrowing mycobacteria is on the rise. Cancer patients with Mycobacterium intracellulare lung infections are often postmenopausal women [56], with a selective defect in interferon gamma production or presence of interferon gamma inhibitor [57, 58]. Rapidly growing mycobacterial (RGM) lung disease is uncommon and mostly seen in patients undergoing chemotherapy and in individuals with pervious pulmonary involvement with cancer [59]. Mycobacterium chelonae and Mycobacterium fortuitum were the prominent RGM associated with lung disease [59, 60]; recently, Mycobacterium abscessus has been a predominate RGM pulmonary pathogen [61]. M. abscessus infections are difficult-to-treat due to high level of drug-resistance [61] and issues related with drug intolerance. Patients with severe cellular immune defects have significantly poor outcome with disseminated RGM end-organ infection [62], with the exception of Mycobacterium mucogenicum catheter-associated infection that responds to prompt removal of the infected catheter and a short course of combination antimicrobial therapy [61].

Fungal Infections

Fungal infections emerged as a significant complication of patients with hematological malignancies after effective chemotherapy became available. The major predisposing factors to these infections were determined to be prolonged neutropenia and adrenocorticosteroid therapy, which interferes with macrophage function. These infections are also prevalent among HSCT recipients who develop graft vs. host disease and receive adrenocorticosteroid therapy.

As early as the mid-1950s, an increasing proportion of patients with acute leukemia developed fungal infections, predominantly candidiasis and aspergillosis [63]. In recent years, infections caused by Zygomycetes, *Fusrium* species, and *Scedasporium* species have become increasingly frequent [64, 65].

There are multiple species of Candida, with different antifungal susceptibilities and patterns of infection [56, 66–68]. Superficial candidiasis occurs in cancer patients receiving radiation therapy and those with impaired T-cell function. Infections involved the oropharynx, esophagus, larynx, urinary tract, and gastrointestinal tract and serve as the origin of disseminated infection, especially in those with neutropenia and long-term intravenous catheters. Disseminated infection is often difficult to diagnose because there may be few signs and symptoms except fever and progressive debilitation and the organism is often not cultured from blood specimens. About 10% percent of patients have multiple skin lesions [69]. There is a chronic form of disseminated candidiasis that occurs in neutropenic patients, which persists after neutrophil recovery and is characterized by persistent fever, debilitation, weight loss, and in some patients, hepatosplenomegaly and right upper quadrant pain [70–72].

Mortality rates have been as high as 70% among patients treated with amphotericin B. Fluconazole prophylaxis has been associated with a significant increase in drug-resistant *Candida krusei* and *Candida glabrata* breakthrough disseminated infections [73–75]. Other alternatives are lipid formulations of amphoterician B and echinocandins. Neutrophil recovery is a critical factor in recovery from candidiasis. Prolonged therapy with fluconazole has been effective for chronic candidiasis and recent experience suggests that anti-inflammatory agents may be useful.

Aspergillosis. The major sites of infection are the lungs and sinuses. Disseminated infection is uncommon. Infection is acquired by inhalation of spores and epidemics have occurred during construction in hospitals. The hyphae invade blood vessels causing thrombosis and infraction and can erode through facial planes, cartilage, and bone. Patients with pulmonary infection may present with symptoms suggesting acute pulmonary embolism. Characteristic nodular infiltrates can be detected on pulmonary CT scans "Halo sign" when radiographs are normal [76]. Culture specimens are often negative, but blood galactamannan tests are helpful in establishing the diagnosis and evaluating treatment response [77]. Sinus infections often present with black eschars on the nose or palate. Progressive infection causes proptosis, endophthalmitis, or cerebral infraction. Therapy consists of effective new Aspergillus active triazole-based drugs such as voriconazole and posaconazole, and echinocandins such as caspofungin and micafungin in combination or as a single agent [78]. Lipid formulations of amphoterician B are also used in combination with other mold-active drugs. Neutrophil recovery and discontinuation of systemic immunosuppressive therapy, especially adrenal costicosteroids, are important for recovery from the infection. Surgical resection of the infected tissue may benefit some patients and resection of residual cavitary lesions may be necessary to prevent pulmonary hemorrhage and late-recurring bacterial superinfections.

Patients at risk of developing cryptococcosis have impaired cellular immunity or are receiving adrenal corticosteroids; hence, patients with CLL or lymphoma or HSCT recipients are at greatest risk. Infection is acquired by inhalation of organisms; hence, the lung is the primary site of infection, although less than 40% of infected patients present with symptoms of pneumonia. The infection can progress rapidly leading to death. Over 50% of cancer patients develop meningitis and some have widely disseminated infection. The latex agglutination test detects cryptococcal antigen in cerebrospinal fluid or blood of infected patients [79]. Optimal treatment consists of initial systemic therapy with amphotericen B plus low-dose flucytosine [80]; for patients with mild-to-moderate infection, high-dose oral fluconazole may be given for maintenance therapy.

Zygomycosis, caused by molds of the order Mucorales, are increasing in frequency [81]. These infections share the same characteristics as aspergillosis, but mortality rates exceed 70% despite amphotericin B therapy. Newer azoles such as posaconazole may be effective therapy [82]. Over 80% of Trichosporon infections are disseminated and the organism can be cultured from blood specimens of most patients. Other infections include endophthalmitis, pneumonia, meningitis, and osteomyelitis [83]. Optimal therapy may be a combination of amphotericin B and fluconazole, but the mortality rate is high in neutropenic patients despite therapy; high-dose voriconazole may be effective in patients with disseminated or hepatosplenic Trichosporon species infection [84]. Breakthrough Trichosporon infection may occur in patients receiving mold-active drugs such as echinocandins or oral broad-spectrum triazoles [85, 86].

Fusarium spp. cause infections predominantly in the sinuses and lungs. Fusariosis like *Aspergillus* species infection are angioinvasive; pulmonary nodular or mass-like disease is indistinguishable from other mold infections [87]. About 75% of infections in neutropenic patients disseminate and the organism often can be cultured from blood specimens. Nearly half of patients are fungemic and up to 80% or more present may develop multiple (>10) nodular skin lesions that develop necrotic center; skin biopsy is diagnostic and should be performed promptly. Mortality remains high despite the availability of highly active triazole drugs against this organism [87].

Unresolved immune suppression continues to influence treatment response among cancer and hematopoietic stem cell transplant (HSCT) recipients with systemic fungal disease [88]. Various strategies including donor granulocyte transfusions in patients with severe neutropenia have not shown significant improvement in outcomes in recent clinical trials [89]. Combined therapy using effective antifungal agents plus recombinant cytokines to boost macrophage, helper, and cytotoxic lymphocyte functions have been explored; a nonrandomized study using granulocyte-macrophage colony-stimulating factor (GM-CSF) and interferon gamma (IFN γ) which were safe and appeared to have a favorable impact in patients receiving donor granulocyte transfusions [90]. Safety of IFN γ has been a concern due to potential cytokine-induced graft compromise and/or GVHD in recipients of allogeneic HSCT; these concerns were not observed in our patients with life-threatening fungal infections [91], although larger, randomized studies are needed to explore this important issue further. Similarly, drugs that may promote pathogen-directed immune capture by introducing configurational changes in these pathogens are being explored [92, 93].

Viral Infections

For many years, little attention was focused on viral infections in cancer patients due to the lack of rapid diagnostic tests and effective therapy. For example, only in recent years have community respiratory viral infections been recognized as potentially serious to immunocompromised patients. Table 1.1 lists most of these viral infections and available therapy. Many acute viral infections represent reactivation of long-standing latent infection.

Human herpes viruses are among the most common causes of viral infections in cancer patients and are associated with significant morbidity and mortality in severely immunocompromised hosts. Herpes simplex viruses cause oropharyngeal and esophageal disease and may disseminate to other organs. Reactivation of varicella-zoster virus occurs mainly in patients with leukemia and lymphoma and can result in localized infection (shingles), disseminated cutaneous infection, pneumonia, encephalitis, hepatitis, or small bowel disease [94]. CMV infection is most often due to reactivation of latent infection, but has also been attributed to transmission by white blood cell transfusions [24, 95]. It is a special risk to HSCT recipients who may receive infected tissue. CMV may cause hepatitis, meninoencephalitis, pneumonitis, or gastroenteritis [96, 97]. The disease has immunosuppressive effects that increase the risk of other infection. Prophylaxis or preemptive therapeutic strategies are necessary for patients undergoing stem cell transplantation [98]. Epstein–Barr virus can cause a fulminant fatal lymphoproliferative disorder in occasional patients following allogeneic stem cell transplantation. Immunocompromised cancer patients occasionally develop interstitial pneumonitis, encephalitis, or hepatitis due to human herpes virus 6 infections.

Progressive multifocal leukoencephalopathy is a demyelinating disease of the brain caused by the JC virus which occurs infrequently among patients with CLL and Hodgkin disease. The disease is due to reactivation of latent infection that is prevalent in normal adults. Symptoms include visual disturbances, speech defects, and mental deterioration leading to dementia and coma with 80% of patients dying within one year. Parvovirus B19 may cause anemia in cancer patients which may be followed by severe polyarthritis. Most patients have been infected with polyomavirus (BK) virus that persists in the genitourinary tract and is a major cause of hemorrhagic cystitis in HSCT recipients [99].

Community respiratory viral infections cause about 30% of respiratory infections in cancer patients during winter and spring and can be a serious threat to transplant recipients and patients with acute leukemia who may develop viral pneumonia or superinfection with bacteria or fungi [100, 101]. Epidemics have occurred in transplant and leukemia units. Some of these patients have very prolonged viral shedding after resolution of symptoms. Viruses causing infection include influenza A and B, respiratory syncytial virus (RSV), parainfluenza (PIV), and adenovirus. In stem cell transplant recipients following PIV and RSV infections, pulmonary obstructive defects were recently recognized; these may be severe and complete resolution may take longer than 12 months after the initial viral infection [102]. Novel respiratory viruses recently recognized to cause serious lifethreatening disease include human metapneumovirus, human cornonavirus NL63 and HKU1, agent of severe acute respiratory syndrome (SARS), and human bocavirus [103, 104]. Adenovirus also causes gastrointestinal infection, hepatitis, hemorrhagic cystitis, pancreatitis, and encephalitis; fatal disseminated adenovirus infections are seen in adults and pediatric patients with profound cellular immune defects such as cordblood transplant recipients with GVHD [105].

Parasitic Infections

Neuro-hepatic toxoplasosis is more common in cancer and transplant recipients in the northeastern United States, whereas strongyloidiasis infestation rates are mostly seen in habitants of southeast and south-central US states. Similarly, amebiasis and giardiasis are infrequently seen in patients from rural residences who consume water from shallow contaminated wells. Latent Toxoplasma gondii infection is difficult to diagnose on the bases of travel, food consumption, or history of domestic feline exposure; serology may be diagnostic, although in patients with B-cell defects PCR analysis may be needed. Malaria is mostly seen in patients traveling to endemic regions without prophylaxis or receiving ineffective chemoprophylaxis due to drug-resistant strains of Plasmodium species. "Airport malaria" has also been seldom reported in patients who reside near airports with frequent international flights. Transfusion malaria has been observed in patients with neoplastic diseases and should be considered and explored in the presence of unexplained fever [106]. Chaga's disease has also been transmitted by transfusions