

Microorganisms for Sustainability 4

Series Editor: Naveen Kumar Arora

Tapan Kumar Adhya

Bibhuti Bhusan Mishra

K. Annapurna

Deepak Kumar Verma

Upendra Kumar *Editors*

Advances in Soil Microbiology: Recent Trends and Future Prospects

Volume 2: Soil-Microbe-Plant
Interaction

 Springer

Microorganisms for Sustainability

Volume 4

Series editor

Naveen Kumar Arora, Environmental Microbiology, School for Environmental Science, Babasaheb Bhimrao Ambedkar University, Lucknow, Uttar Pradesh, India

More information about this series at <http://www.springer.com/series/14379>

Tapan Kumar Adhya • Bibhuti Bhusan Mishra
K. Annapurna • Deepak Kumar Verma
Upendra Kumar
Editors

Advances in Soil Microbiology: Recent Trends and Future Prospects

Volume 2: Soil-Microbe-Plant Interaction

 Springer

Editors

Tapan Kumar Adhya
KIIT School of Biotechnology
Bhubaneswar, Odisha, India

K. Annapurna
Division of Microbiology
Indian Agricultural Research Institute
New Delhi, India

Upendra Kumar
National Rice Research Institute
Cuttack, Odisha, India

Bibhuti Bhusan Mishra
Department of Microbiology
Orissa University of Agriculture and
Technology
Bhubaneswar, Odisha, India

Deepak Kumar Verma
Department of Agricultural & Food Engineering
Indian Institute of Technology
Kharagpur, West Bengal, India

ISSN 2512-1901

ISSN 2512-1898 (electronic)

Microorganisms for Sustainability

ISBN 978-981-10-7379-3

ISBN 978-981-10-7380-9 (eBook)

<https://doi.org/10.1007/978-981-10-7380-9>

Library of Congress Control Number: 2017964109

© Springer Nature Singapore Pte Ltd. 2017

This work is subject to copyright. All rights are reserved by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, express or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Printed on acid-free paper

This Springer imprint is published by Springer Nature

The registered company is Springer Nature Singapore Pte Ltd.

The registered company address is: 152 Beach Road, #21-01/04 Gateway East, Singapore 189721, Singapore

Contents

1	Soil Microbial Diversity: An Ecophysiological Study and Role in Plant Productivity	1
	Bighneswar Baliyarsingh, Suraja Kumar Nayak, and Bibhuti Bhusan Mishra	
2	Microbial Diversity and Soil Health in Tropical Agroecosystems	19
	Dipanti Chourasiya, Mahaveer P. Sharma, Hemant S. Maheshwari, Aketi Ramesh, Sushil K. Sharma, and Tapan Kumar Adhya	
3	Plant Growth-Promoting Microbes (PGPM) as Potential Microbial Bio-Agents for Eco-Friendly Agriculture	37
	Madhurama Gangwar, Preeti Saini, Pooja Nikhanj, and Sukhjinder Kaur	
4	Plant Growth-Promoting Rhizobacteria for Abiotic Stress Alleviation in Crops	57
	Sangeeta Paul, Ajinath S. Dukare, Bandeppa, B. S. Manjunatha, and K. Annapurna	
5	Phosphate-Solubilizing Microorganisms in Sustainable Agriculture: Genetic Mechanism and Application	81
	A. Pradhan, A. Pahari, S. Mohapatra, and Bibhuti Bhusan Mishra	
6	Arbuscular Mycorrhizal Fungi (AMF) for Sustainable Rice Production	99
	P. Panneerselvam, Upendra Kumar, T. C. K. Sugitha, C. Parameswaran, Sowarnalisha Sahoo, A. K. Binodh, Afrin Jahan, and A. Anandan	
7	Biological Nitrogen Fixation in Cereals Crops: A Bacterial Perspective	127
	S. Garcha and P. K. Maan	

8	Biological Control as a Tool for Eco-friendly Management of Plant Pathogens	153
	Mamta Sharma, Avijit Tarafdar, Raju Ghosh, and S. Gopalakrishanan	
9	Biological Control of Insect Pests for Sustainable Agriculture	189
	Satyavir S. Sindhu, Anju Sehwat, Ruchi Sharma, and Aakanksha Khandelwal	
10	Soil Organic Matter and Microbial Role in Plant Productivity and Soil Fertility	219
	Tapas Biswas and Subhas Chandra Kole	

Editors' Biography

Dr. Tapan Kumar Adhya is currently the Director, South Asia Nitrogen Centre, New Delhi, and is also working as Professor in the School of Biotechnology, KIIT University, Bhubaneswar, Odisha, India. He has more than 150 publications and 30 book chapters. He is Editor/Associate Editor of several international research journals published by Springer.

Dr. Bibhuti Bhusan Mishra is presently working as Professor and Head in the Department of Microbiology in Orissa University of Agriculture and Technology, Bhubaneswar, India.

Dr. K. Annapurna obtained her M.Sc. (1982) and Ph.D. (1986) degrees in Microbiology from Indian Agricultural Research Institute, New Delhi, India. She has been a pioneer researcher in the field of molecular ecology of legume – *Rhizobium* symbiosis, *Azospirillum*, plant growth-promoting rhizobacteria (PGPR), and initiated work on soybean rhizobial genetic diversity.

Deepak Kumar Verma is an Agriculture Science professional. He is a PhD Research Scholar with major specialization in Food Process Engineering (FPE), apart from theory and lab practicals at Agricultural and Food Engineering Department, Indian Institute of Technology (IIT), Kharagpur (WB), India, the prestigious institute of India which has world ranking between 225 and 250. His area of specialization during master's was Agricultural Biochemistry.

Dr. Upendra Kumar is working as Scientist at ICAR-National Rice Research Institute, Cuttack, India, and also served as Visiting Scientist at CSIRO, Waite Campus, Adelaide, Australia. He obtained his M.Sc. and Ph.D. in Agricultural Microbiology from IARI, New Delhi, and also received many National and International awards.

Chapter 1

Soil Microbial Diversity: An Ecophysiological Study and Role in Plant Productivity

**Bighneswar Baliyarsingh, Suraja Kumar Nayak,
and Bibhuti Bhusan Mishra**

Abstract Soil is considered as one of the most competent ecosystems for subsistence of microorganisms. Soil microbial community structure and activity depend largely on structure and status of the soil habitat. Diverse heterotrophic microbial communities in soil along with their complex web of interaction facilitate the cycling of micro- and macro-nutrients in soil ecosystem. The demand of sustained plant productivity is achieved through managing soil fertility. The dynamic relationships between different components, living or nonliving, of agroecosystem control the richness of plants or crops. In turn, soil organic matter is influenced by the inputs from plants and also their chemistry makes each ecosystem somewhat unique in its microbial community. Though the role of soil microbiome is widely known, we still have a limited understanding of its complexity. Thus, understanding the microbial diversity will enhance our ability of increasing agricultural production.

Keywords Soil microbiology · Soil microbial habitat · Plant-soil microbial interaction · Microbial diversity · Soil fertility

1.1 Introduction

In the recent past, growing understandings on the potential of microorganisms have given much emphasis to explore and study the active microbial population inhabiting the soil. Since the beginning of the nineteenth century, soil

B. Baliyarsingh · S. K. Nayak (✉)
Department of Biotechnology, College of Engineering and Technology, Bhubaneswar, Odisha,
India
e-mail: surajnayak3@gmail.com

B. B. Mishra
Department of Microbiology, Orissa University of Agriculture and Technology,
Bhubaneswar 751003, Odisha, India

© Springer Nature Singapore Pte Ltd. 2017

T. K. Adhya et al. (eds.), *Advances in Soil Microbiology: Recent Trends and Future Prospects*, Microorganisms for Sustainability 4,
https://doi.org/10.1007/978-981-10-7380-9_1

microorganisms have gained importance as the driver of biochemical processes that are beneficial to ecosystem. These microorganisms carry out different processes of decomposition of organic substances, transformation of elements, and also recycling of nutrients that are essential for growth of animals, plants, and crops. Some soil-inhabiting microorganisms are, however, injurious to plant and animal life, acting as pathogens affecting the host directly or releasing toxic substances in the soil ecosystem. A better understanding of the soil microbes is thus important to interpret their impacts on agriculture and environment. Hence, the soil microbiologists are not only focusing on diversity of microbes in soil but also their interaction with the environment as well as with other organisms.

1.2 Historical Perspectives of Soil Microbiology

During the mid-nineteenth century, studies of microbiologists like Louis Pasteur, Selman Waksman, and Sergei Winogradsky led the foundations for the modern soil microbiology research. Among various discoveries by Winogradsky, the father of soil microbiology, some of the notable studies are the sulfur cycle, role of CO₂ and inorganic ions on microbial growth (chemoautotrophy), and nitrification. He was also honored by naming one of the nitrifying bacteria species as *Nitrobacter winogradskii*. Nitrogen fixation can be accomplished by nonsymbiotic bacteria which was first suggested by Berthelot in the late nineteenth century. Soil microbes bring about the mineralization of litters and make easy availability of the essential nutrients for the growth and development of plants and animals. It was also emphasized that the addition of stable manure to the soil is more effective than to direct addition of inorganic nutrients.

The fixation of nitrogen by leguminous plants is one of the most important and well-studied microbial process. Studies on soil fungus and bacteria involved in nitrification and denitrification and chemical transformations of nitrogen in soil along with in composts, laid foundation for the modern era of soil microbiology, and all these processes have direct effect on plant growth and productivity. The overall relationship of soil microbial population with that of soil fertility also gave rise to the concept of inoculating desired microorganisms to soil (Kuramae et al. 2012).

Apart from agronomic importance of soil microbes, in 1939 S. Waksman and Rene Dubos found a soil actinobacteria (formerly actinomycete) *Streptomyces* sp., with antibiotic properties. Waksman was awarded Nobel Prize in 1952 for the findings on antimicrobial properties of soil microbes. Studies on the actinobacteria of the soil by Krinsky, Conn, Waksman, and Curtis, by Melin and Hayner on mycorrhizal fungi, and on the soil protozoa by Cutler widened the purview of soil microbiology. Since their discovery, soil microbes are widely studied and applied to various realms of human endeavors (Balser et al. 2010).

1.3 Habitat of Soil Microorganism

Soil is not an inert material but a dynamic and most biodiverse ecosystem as it is home to several thousand different species of organisms/microorganisms. The population, composition, and activities of soil microbes are influenced by their soil habitat. Cultivated lands are rich in organic matters and contain much higher microbial population than sandy or eroded soils. Within this habitat, the soil microbes respire, compete for food, cooperate among themselves, and respond to the changes in their inhabiting environment.

Typically, differing sizes of soil aggregates and soil pores constitute the soil microbial habitat (Sylvia et al. 2005). The texture and soil types influence the composition and population of microbial community and vice versa. Some soil microbes secrete polysaccharides, gums, and glycoproteins, which glue soil minerals together, forming the basis for soil structure. Further, fungal hyphae and plant roots bind soil aggregates together that provide favorable environment for plant growth. Rudakov (1951) suggested that active humus play an important role of cementing the soil particles into aggregates. The soil cementing substances are mostly composed of (1) compounds of uronic acids, (2) bacterial proteins or products of their lytic activities, and (3) lysates of fungal cultures and/or colloidal protein compounds synthesized by soil bacteria.

Similarly the diversity of soil microbial community is influenced by physical parameters like temperature, humidity and seasonal variations (Lipson 2007), soil acidity or alkalinity (pH), oxygen levels, and availability of nutrients. The soil fertility can be indirectly correlated with the overall microbial biomass which depends on availability and quality of carbon, termed as soil organic carbon (SOC). The amount of SOC available within the initial 1m depth of soil is more than two to three times the carbon amount present aboveground (Brady and Weil 2002). Thus most biologically active region of the soil is believed to be the upper 20–30 cm of soil where abundance of soil microbial communities are present due to rich SOC (Fierer et al. 2007; Jobbágy and Jackson 2000; Veldkamp et al. 2003). Carbon availability often declines with depth, as does overall microbial biomass. Also many microorganisms exist in topsoil having rich carbon sources, than in the subsoil. They are especially abundant in the area close to plant roots (called the rhizosphere), where sloughed-off cells and root exudates provide carbon sources. It is established that there is decrease in fungal-to-bacterial ratios with increase in depth. Approximately 3×10^4 bacteria, 1.5×10^5 fungi, 6×10^4 algae, 1×10^4 protozoa, 5×10^5 nematodes, and 3×10^4 earthworms are estimated to be present in the soil (Pankhurst 1997).

Soil fertility is one of the significant parameters of natural or managed agroecosystem to achieve sustained productivity. To access the soil quality, changes in chemical, physical, and biological properties are monitored which are the indirect measure of productivity and diversity. Biological indicators considered to access soil quality are microbial biomass, soil respiration, soil enzyme activities, earthworm numbers, etc. The metabolic quotient (qCO_2) is the estimate of the

amount of CO₂-C released per unit of microbial biomass in time that represents the thriving status of the soil microbes. Besides, biochemical indicators such as microbial enzymes are also useful in understanding soil fertility. The biological reactions most commonly studied were metabolism of nitrogenous compounds which includes ammonification, nitrification, denitrification, and the fixation of nitrogen. Microbial indicators have advantage in forecasting early to any changes in soil quality as it is being susceptible to minor changes in ecosystem.

In addition to structural diversification of soil microbes, soil organisms are naturally active during certain period of the year. Most are active when the soil is warm and moist, like during late spring and early summer. Moreover minerals, aggregation of the soil particles, and soil porosity influence diversification of bacterial community in soil microhabitats (Certini et al. 2004; Carson et al. 2007). The uniqueness of each soil ecosystem is due to typical microbial communities performing diverse activities and multifaceted interactions with their habitat (Wixon and Balsler 2009) (Fig. 1.1).

1. The soil microbes provide basic macroelements of nutrition, viz., carbon, nitrogen, and phosphorous for growth and development of plants.
2. The microorganisms show indirect way of controlling the production and regulation of growth-specific phytohormones.
3. Symbiotic association with plants or plant parts (e.g., leguminous plant roots) is maintained by very special group of microbes.
4. Not all soil-dwelling microbes show beneficial interaction; some have harmful effects on higher plants. The microbes either show competition for uptake of basic nutrients from the soil or depend on the higher plants as parasites or attacking them by producing toxic chemicals.
5. Indirect injury to plant growth is observed when various bacteriophages feed on useful bacteria.
6. On the other hand, the immediate vicinity of plant parts creates a microenvironment to the microorganisms for their growth and proliferation.
7. The plant residues and root exudates are the rich source of nutrients, consumed by the microorganisms.
8. Plants metabolites may have antagonistic effects on the growth and sustainability of microorganisms.

Various groups of bacteria, fungi, and actinobacteria either penetrate the roots of plants and live there or live in close proximity of the root system. The concept of the “rhizosphere” is introduced to assign the close association between soil microorganisms and the root systems of higher plants. The “root region” encompasses both the root surface and the rhizosphere which are the zones of high microbiological activity. Typically this type of association may be considered as midway between true symbiosis, observed between root-nodule bacteria with the leguminous plants and the phenomena of parasitism. Accumulation of manure, lowering of the concentration of certain mineral nutrients, partial desiccation of the soil, and increase in soil carbonates following root excretion are the major causes of rhizosphere effect. The tuberization and protein formation in certain seeds in plants are

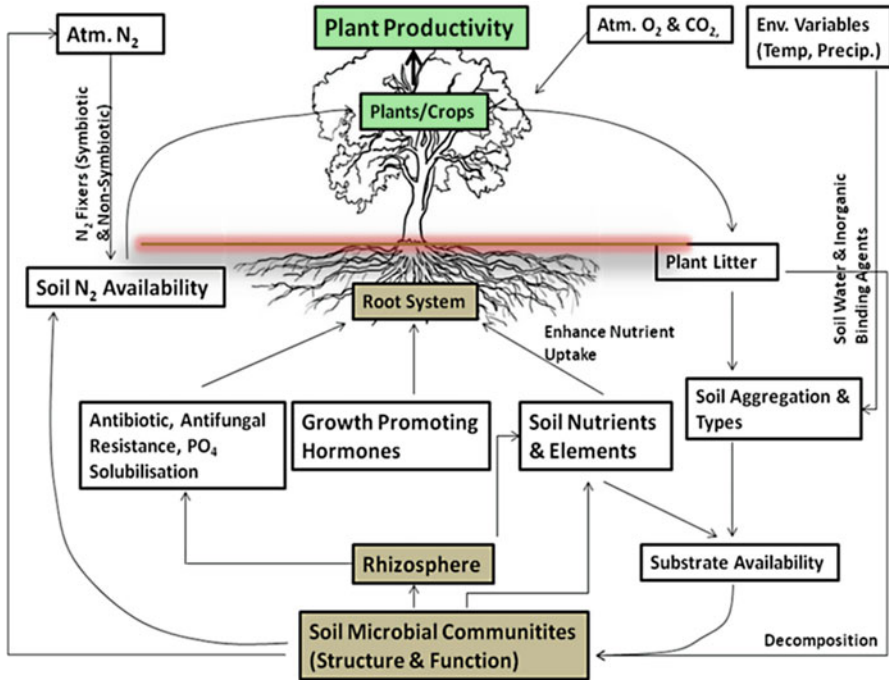


Fig. 1.1 Dynamic interactions between microbes and plants

induced by some fungi. Garrett (1951) emphasized the necessity to distinguish between rhizosphere effects and typical characteristics of living roots with that of the diseased root microenvironment.

The diversified soil microbes play pivotal roles in an array of terrestrial ecosystem functions such as nutrient recycling, sustaining plant growth, water purification, carbon storage, maintenance of soil structure, degradation of xenobiotics, nitrogen fixation, or competitor for pathogens to successfully establish desired microbial populations (Insam 2001). Similarly, the decomposition and nutrient recycling are carried out by diverse group of microbes in a cascading network of chemical process. By decomposition processes, soil microbes liberate simpler carbon compounds, nitrogen, CO₂, and minerals from dead organic material which are ultimately consumed by higher plants.

The decomposition carried out by soil microbes is either aerobic (aerobic bacteria and fungi) or anaerobic microbes. The aerobic decomposition of plant residues by the microbes produces humic acid in the soil. The abundance of humic acid and fulvic acid facilitates the gluing properties of humus. In addition, various protopectinase enzymes were produced by soil microbes like sporogenous *Bacilli*, *Bacillus polymyxa*, *B. radiobacter*, *B. mycoides*, *B. laterosporus*, and *Clostridium macerans*. Some fungi also take part in the decomposition of plant residues by producing the humic acids. *Trichoderma lignorum*, *Mucor intermedius*, and

Mortierella isabellina are few examples of fungi, mostly involved in the process of soil structure formation (Harris et al. 1966).

The solubilization of soil minerals, especially the carbonates, phosphates, and zeolites, is carried out by microorganisms with the help of CO₂; nitrous, nitric and sulfuric acids; and organic acids. Soil toxins constitute the toxins produced from soil microbes as a result of their activities. Thus the soil solution represents very dynamic part of the soil where most of the biochemical processes occur. The fouling properties of soil can be treated with heat, volatile antiseptics, or simple lime to increase productivity. Reports indicate that the germination of plant seeds and seed growth is favored by the soil microorganisms. For example, rapid evolution of CO₂ by microbial respiration creates anaerobic conditions unfavorable to oxidation and favorable for germination. Recent studies depict that microorganisms may produce hormones and/or similar substances which promotes plant growth.

1.4 Influence of Plants on Soil Microorganisms

Although, there are few experimental investigations that suggest the influence of plant diversity on soil microorganisms (Bargett and Shine 1999; Wardle et al. 1999; Stephan et al. 2000), there are reasons to expect that plant diversity can affect microbial population in the soil ecosystem. It is due to the lack of experimental procedure to accesses the direct influence of the growing plant or plant products on soil microbes. Mostly the effect of plants on soil microbes is studied indirectly by (a) measuring the numbers of microorganisms, (b) nitrifying and denitrifying capacity of the soil, and (c) oxidizing power of the soil in terms of aeration or CO₂ production. The effects of growing plants on the structure and function of soil microorganisms can be represented as:

1. The growth of soil bacteria and fungi is favored by the soluble organic and inorganic compounds like formic, oxalic, and malic acids, certain reducing and nonreducing sugars, phosphatides, and various nitrogenous compounds secreted by the plants.
2. In addition to these plants in the form of dead roots and root hairs, epidermal cells and other waste products help the growth of microorganisms.
3. Various soil minerals, including nitrates, phosphates, and potassium salts, are continuously removed by the growing plants resulting in change in soil solution and modification of microbial activities.
4. Plants excrete considerable CO₂ into the soil which increases the solubility of certain inorganic soil constituents and thereby changing the composition of the soil atmosphere.
5. Soil porosity and structure are modified and affected by the plant diversity.
6. Sometimes, plants can hinder the growth of soil microbes by removing a considerable amount of moisture from soil.

Feedback mechanisms between plant and microbial communities control the richness of plant species and productivity. For example, soil bacteria are stimulated by cowpeas, field peas, vetch, and soybean. Among the bacteria, particularly, the *Radiobacter* sp. increased in number when legumes are cultivated. In contrast, continuous production of single type of plant in a soil leaves residues that may result in a change in the chemical composition of the soil leading to microbial disequilibrium. This is evident from the fact that the pathogenic fungi population increases in soil habitat by continuous farming of wheat, flax, or clover. While the alfalfa roots have only a slight stimulating effect upon filamentous fungi, the eggplants had a significant effect (Starkey 1931). A range of stimulatory effects on soil microbes by cultivation of alfalfa, rye, and vetch plant are observed ranging least on actinobacteria, slightly on fungi, and highest toward soil bacteria.

1.5 Microbial Diversity and Soil Biological Processes

Soil is reckoned as a storehouse of microbial activity, though the space inhabited by active microorganisms is approximately less than 5% of the total occupied area. Despite their small volume, soil microorganisms are key players in the global cycling of organic matter, reworking organic residues or mineralizing them to CO₂, H₂O, N₂, P, S, and other nutrients (Table 1.1). Hence, soil microbes are playing the most vital role in plant growth regulation and contributing thereby to the interdependence of diversity – fecundity (Pradhan et al. 2014; Mitchell 2003; van der Heijden et al. 2008). Major microbiological activity is confined to the clusters of accumulated organic matter and rhizosphere. Thus, there is a growing demand to study on microbial community composition in various microhabitats in nature.

The soil as a habitat exhibits utmost variability leading to adjacent microhabitats differing widely in their physical and chemical properties. Diversified heterotrophic microbial communities residing in soil control the ecosystem carbon (C) and nitrogen (N) cycling (Fig. 1.2) through various processes that control ecosystem and represent a potential mechanistic link in between plant diversity and ecosystem function. The growth-limiting resource availability plays a key role in determining biotic community composition (Tilman 1982, 1987). The resource availability for soil microbial communities is incarcerated through organic compounds in litters which can be used for cellular energy generation (Smith and Paul 1990). Changes in plant diversity could alter the yield as well as the range of organic compounds that limit and thus control the composition as well as microbial community function.

1.5.1 Bacteria and Actinobacteria

The sustainability of soil function is the result of bacterial diversity. Endemism and clonality, which have an intrinsic spatial dimension (Grundmann and Normand

Table 1.1 Major roles of diverse soil microorganisms

Domain/phylum/ group of microorganism	Important species	Association with host	Major role of these species/ strains	References
Archaea	<i>Nitrosophaera viennensis</i> , <i>Methanosarcina</i> sp., <i>Methanoseta</i> sp.,	No association	Contributing to ammonia oxidation in soil	Aislabie and Deslippe (2013)
			Weathering minerals as well as nutrient cycling in soil	
			Carry out N ₂ fixation	
			Curtaiment of atmospheric N ₂ to NH ₄ ⁺	
Bacteria	<i>Rhizobium</i> sp., <i>Bradyrhizobium</i> sp., <i>Mesorhizobium</i> sp.,	Symbiotic	Fixation of atmospheric nitrogen in leguminous plant in root nodules	Giordano and Hirsch (2004)
			Present inside the host root system	
			Fixes atmospheric nitrogen usable form such as ammonia	
Actinobacteria	<i>Pseudomonas</i> sp., <i>Bacillus</i> sp., <i>Klebsiella</i> sp., <i>Azotobacter</i> sp., <i>Azospirillum</i> sp., <i>Azomonas</i> sp.,	Asymbiotic	Remain in close proximity of root system	Glick et al. (1999)
			Fixation of atmospheric nitrogen	
			Solubilization of minerals	
Verrucomicrobia	<i>Arthrobacter</i> sp., <i>Rhodococcus</i> sp., <i>Streptomyces</i> sp., <i>Rubrobacter</i> sp., <i>Terrabacter</i> sp., <i>Acidimicrobium</i> sp., <i>Chloroflexi</i> sp., <i>Planctomyces</i> sp., <i>Gemmatimonadetes</i> sp., <i>Micromonospora</i> sp., <i>Streptomyces</i> sp., <i>Streptosporangium</i> sp., <i>Thermobifida</i> sp., <i>Frankia</i> sp., <i>Nocardia</i> sp. <i>Kitasatospora</i> sp., <i>Methylacidiphilum</i> SoIV, <i>Methylacidiphilum</i> KamI, <i>Verrucomicrobium spinosum</i>	No association	Antagonistic efficiency against different fungal root pathogen	Ahemad and Kibret (2014); Bhattacharyya and Jha (2012) and Franco-Correa and Chavarro-Anzola (2014)
			Efficient IAA producers (plant growth hormone)	
			Nitrogen fixation and associative activity	
			No association	
			No association	
			No association	

Yeast	<p><i>Kluyveromyces waltii</i>, <i>Scyaromycopsis cataeensis</i>, <i>Pachyichospora transvaalensis</i> <i>Rhodotorula</i> sp., <i>Azotobacter</i> sp., <i>Cryptococcus</i> sp., <i>Lipomyces</i> sp., <i>Meyerozyma</i> sp.</p>	Various association	<p>Utilization of bacterial, plant root exudates for synthesis of plant proteo- tants and other useful substances involved in plant growth Known producers of extracellular polymeric substances Produces xylitol, phosphate solubilization and associated activities, and soil aggregation</p>	Vishniac (1995), Cho et al. (2001), Nakayan et al. (2009)
Cyanobacteria	<p><i>Nostoc</i> sp., <i>Anabaena</i> sp., <i>Leptolyngbya</i> sp.</p>	Symbiotic and nonsymbiotic	<p>Improved the soil C, soil N, and exopolymetric substance contents of the soil Releases amino acids and proteins, polysaccharides and carbohydrates, vitamins, and phytohormones as elicitor molecules for plant growth promotion Increases resistance of plants against biotic and abiotic stresses</p>	Bano and Iqbal (2016)
Fungi	<p><i>Phoma</i> sp., <i>Trichoderma</i> sp., <i>Penicillium</i> sp., <i>Rhizoctonia</i> sp., <i>Pythium</i> sp.</p>	Various association	<p>Improve plant growth Induce systemic resistance (ISR) against plant pathogens Enhances biomass production and promotes lateral root growth Niche exclusion, antibiosis, predation, mycoparasitism, and ISR induction</p>	Murali et al. (2012), Whipps (2001) and Mauchline et al. (2002)

(continued)

Table 1.1 (continued)

Domain/phylum/ group of microorganism	Important species	Association with host	Major role of these species/ strains	References
AMF	<i>Acaulospora morrowiae</i> , <i>Glomus etunicatum</i> , <i>Rhizophagus irregularis</i> , <i>Gigaspora rosea</i>		Increases the soil nutrient availability Increase yield by enhancing host resource uptake by sharing Improved water relations and increases antagonism against plant pathogens Phytoremediation of polluted soil	Ellouze et al. (2014) and Lovelock et al. (2004)

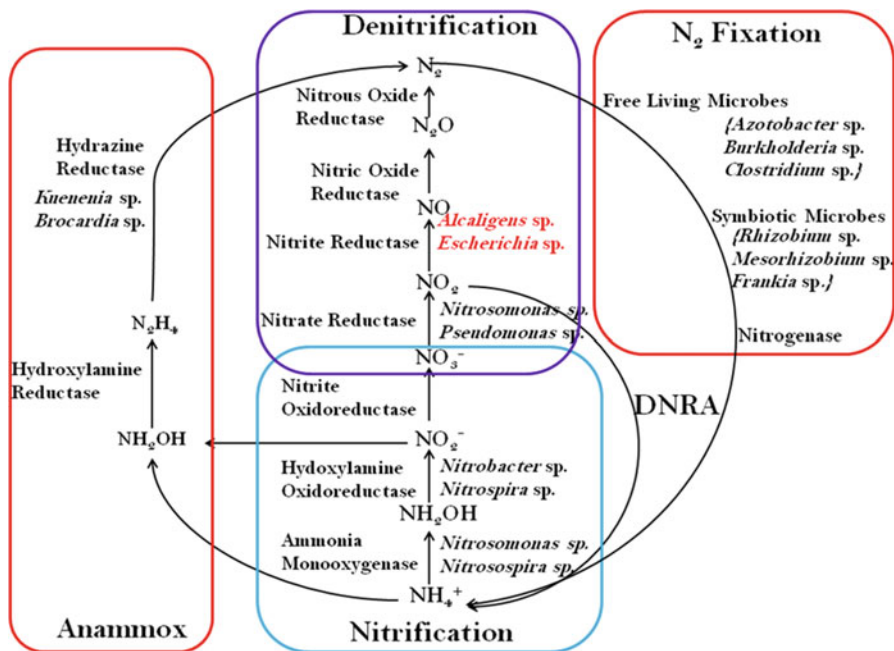


Fig. 1.2 Soil microbes involved in biological nitrogen cycles

2000), may have significant implications in soil function. Soil texture influencing bacterial community structure, even if the change is minor and with unchanged mineral composition of soil, probably results in the scattered arrangement of bacteria (Hattori and Hattori 1976). Bacterial diversity and community structure are influenced by a range of physicochemical properties of the soil matrix, including pore size, particle size, and the availability of water and carbon (Carson et al. 2009). At low pore connectivity, the maximal bacterial diversity is probably mediated by lots of interacting factors which favors the coexistence and decreases the competitive interactions (Carson et al. 2010).

Actinobacteria, being a slow grower in the soil, may colonize roots slowly but successfully. Mostly bacteria and actinobacteria bring rhizospheric change through altering the composition of small molecules like sugars, amino acids, fatty acids, flavonoids, and strigolactones leading to significant modification in the plant (Badri and Vivanco 2009) phenology. In turn, microbes inhabiting the rhizosphere release phytohormones, small molecules, or volatile compounds and regulate plant growth and root morphogenesis (Castro-Sowinski et al. 2007).

1.5.2 *Fungi*

Several advanced, independent soil microbiological methods for charactering the microbial biomass and soil microbial community structure uncovered that living and dead soil bacteria were associated with the clay fraction, whereas lower eukaryotes as fungi and their exoenzymes involved in the degradation of complex organic compounds were found in association with soil particulate organic matter. The plant species richness is also positively correlated with arbuscular mycorrhizal fungi (AMF) richness (Klironomos et al. 2000). AMF contribute positively to the plant productivity due to diverse relationship with increase in soil microbial diversity leading to an increase in per capita plant productivity (van der Heijden et al. 2008). The soil health is benefited by extensive mycelia networks of AMF with the glomalin secretion leading to the improvement of structural stability and quality of soil. At the instance of resource competition, AMF increases the yield by improving the resource uptake as phosphorous (P) along with carbon (C) and nitrogen (N) (Koide 1991). From an agroecological point of view, the ecological function imparted by AMF signifies the important impact these groups of symbiotic organisms have on the agricultural productivity and sustainability (Ellouze et al. 2014).

1.5.3 *Archaea*

There is abundant presence of archaea in soil, predominantly by the members of the phylum *Crenarchaeota*, mostly found and isolated below the topsoil. In general, these are grouped in the class of anaerobic soil microbes. Some members from *Euryarchaeota* phylum especially methanogens are active in the absence of O₂ created due to water logging (Angel et al. 2012). *Methanosaeta*, *Methanocella*, and *Methanosarcina* sp. comprising methanogens are widely found in soil microhabitat. Being strict anaerobes, they break down the organic molecules to CH₄ and CO₂ and participate in anaerobic food chain. Although, the pathways used by these methanogens for production of CH₄ from methylated compounds vary that include either reduction of CO₂ and methanol, cleavage of acetate, or reduction of methylated compounds from methane (Singh et al. 2017). Both *Methanosarcina* sp. and *Methanosaeta* sp. reduce acetate to produce methane. The oxidizing archaea like ammonia-oxidizing archaea (AOA) are more prominent at the subsoils (Di et al. 2010). Under high ammonia substrate concentrations, AOA dominates soil microbial community.

1.5.4 Cyanobacteria

In many agricultural fields, cyanobacteria are one of the vital inhabitants, where they efficiently contribute toward biological nitrogen fixation, help “P” solubilization, and improve crop productivity as well as soil fertility. The nitrogen-fixing cyanobacteria are helpful in crop production with saving partially or entirely mineralized nitrogen. Novel association between agriculturally important plants, N₂-fixing cyanobacteria, and cereal plants is now immersing (Spiller et al. 1993). Cyanobacteria, as inoculants in rice production, have enhanced soil fertility and improved soil structure, besides enhancing crop yields. Highest populations of *Anabaena* sp. ($1 \times 10^6 \text{ g}^{-1}$ soil) and *Nostoc* sp. ($9.1 \times 10^4 \text{ g}^{-1}$ soil) were recorded in paddy field soil, when supplemented with nitrogen. *Calothrix* sp., *Hapalosiphon* sp., and *Westiellopsis* sp. were among the other isolates from the rhizosphere region. Out of the nonheterocystous cyanobacteria from the rhizosphere, *Oscillatoria* sp. and *Phormidium* sp. were prevalent, but not in all rice cultivars, *Scytonema* sp. was found to be common in saline soil (El-Ayouty et al. 2012). Ninety-eight percent of Indian soils require inorganic phosphorus (chemical or biological) through phosphorus fertilization. Due to the low solubility and chemical fixation of phosphorus present in soil, only 0.1% is available to the crop plants. The soil microorganisms act as a biological combat system by solubilizing the insoluble inorganic phosphorous to its simplest form and making it available to the plants. The cyanobacteria involved in phosphate solubilization (PS) are *Anabaena* sp., *Calothrix* sp., *Nostoc* sp., *Scytonema* sp., and *Tolypothrix ceylonica*.

1.5.5 Yeast

Some selective variety of yeasts promote plant growth by means of pathogen suppression, phytohormone production, phosphate solubilization, N and S oxidation, siderophore production, and stimulation of mycorrhizal root colonization. Strains have been identified as biofertilizer yeasts (Eman et al. 2008). In vitro nitrification can be achieved through selective members of *Candida* sp., *Geotrichum* sp., *Rhodotorula* sp., *Saccharomyces* sp., and *Williopsis* sp. *Saccharomyces* sp. and *Williopsis* sp. were able to oxidize elemental sulfur in vitro for production of phosphate (PO_4^{3-}), sulfate (SO_4^{2-}), and tetrathionate ($\text{S}_4\text{O}_6^{2-}$) (Al-Falih and Wainwright 1995). Among yeasts, *Rhodotorula minuta*, *Saccharomyces cerevisiae*, and *Torula thermophila* cause inorganic phosphate solubilization. Other phosphate-solubilizing yeasts are *Hansenula* sp., *Klockera* sp., *Rhodotorula* sp., and *Debaryomyces* sp., and the most efficient was *D. hansenii*. *Schwanniomyces occidentalis* exhibited PS activity on Mussoorie rock phosphate (Gaur 1990). Yeasts also synthesize antimicrobials and other metabolites required for plant growth, from polysaccharides and amino acids released by bacteria and root exudates. Yeasts are rich source of tryptophan a

precursor of indole acetic acid (IAA) that stimulates cell elongation and division (Agamy et al. 2013).

1.6 Evaluation of Soil Fertility by Measuring Microbiological Activities

Various methods have been developed by researchers worldwide to determine the soil fertility on the basis of its microbiological activity.

1. A general mechanism of assessment of soil fertility is by adding moistened soil samples with 60 or 70% of water-holding capacity, with specific nutrient solution, followed by incubation at 20–30 °C for 7–30 days and observes biological changes.
2. The biological reactions involving metabolism of nitrogenous compounds are analyzed to evaluate the soil fertility. The common biological reactions like ammonification, nitrification, denitrification, and nitrogen fixation are used in soil fertility analysis.
3. The abundance of certain organisms in the soil and presence of other organisms can be examined by various microbiological culture methods. Measuring the changes in concentration of certain important plant nutrients in the soil with respect to microbial activity and abundance provides idea about soil fertility.

The major difficulty involved in the use of such methods is the dependency of soil microbial reaction on many variables in a natural scenario. This comprises of different intrinsic soil conditions, weather condition effects, and soil management.

1.7 Conclusion

Soil microbiology is a multidisciplinary branch of science which deals with the study of microorganisms associated with soil and their interaction. The soil, being the natural habitat for the survival and growth of microorganisms, has always given great emphasis to study its physicochemical and biological characteristic features. It is very important to understand the relationship of microorganisms with respect to higher plants as well as to exploit the ecto- and endo-microbial association for a sustainable soil microbial ecosystem. Soil microbiology defines a brief idea about microbes nurtured by nature and the chemical properties of the soil depending upon the geographical limits and climatic changes. It should make an exciting study in the field of agriculture and environmental sciences to create an intricate road map by extracting knowledge behind the complex microbiological activities, interacted with the environmental cues that would dramatically improve plant productivity and thus increase socioeconomical values.

References

- Agamy R, Hashem M, Alamri S (2013) Effect of soil amendment with yeasts as bio-fertilizers on the growth and productivity of sugar beet. *Afr J Agri Res* 8(1):46–56. <https://doi.org/10.5897/AJAR12.1989>
- Ahemad M, Kibret M (2014) Mechanisms and applications of plant growth promoting rhizobacteria: current perspective. *J King Saud Univ Sci* 26:1–20. <https://doi.org/10.1016/j.jksus.2013.05.001>
- Aislabie J, Deslippe JR (2013) Soil microbes and their contribution to soil services. *Soil Microbial Diversity*
- Al-Falih AM, Wainwright M (1995) Nitrification, S oxidation and P-solubilization by the soil yeast *Williopsis californica* and by *Saccharomyces cerevisiae*. *Mycol Res* 99:200–204. [https://doi.org/10.1016/S0953-7562\(09\)80886-1](https://doi.org/10.1016/S0953-7562(09)80886-1)
- Angel R, Claus P, Conrad R (2012) Methanogenic archaea are globally ubiquitous in aerated soils and become active under wet anoxic conditions. *ISME J* 6:847–862. <https://doi.org/10.1038/ismej.2011.141>
- Badri DV, Vivanco JM (2009) Regulation and function of root exudates. *Plant Cell Environ* 32:666–681
- Balser TC, Wixon D, Moritz LK, Lipps L (2010) The microbiology of natural soils. In: Dixon GR, Tilston EL (eds) *Soil microbiology and sustainable crop production*. Springer, Berlin, pp 27–57
- Bano SA, Iqbal SM (2016) Biological nitrogen fixation to improve plant growth and productivity. *Int J Ag Innov Res* 4(4):2319–1473
- Bargett RD, Shine A (1999) Linkages between plant litter diversity, soil microbial biomass and ecosystem function in temperate grasslands. *Soil Biol Biochem* 31:317–321. [https://doi.org/10.1016/S0038-0717\(98\)00121-7](https://doi.org/10.1016/S0038-0717(98)00121-7)
- Bhattacharyya PN, Jha DK (2012) Plant growth-promoting rhizo-bacteria (PGPR): emergence in agriculture. *World J Microbiol Biotechnol* 28:1327–1350. <https://doi.org/10.1007/s11274-011-0979-9>
- Brady NC, Weil RR (2002) *The nature and properties of soils*, 13th edn. Prentice Hall, Upper Saddle River
- Carson JK, Rooney D, Gleeson DB, Clipson N (2007) Altering the mineral composition of soil causes a shift in microbial community structure. *FEMS Microbiol Ecol* 61:414–423. <https://doi.org/10.1111/j.1574-6941.2007.00361.x>
- Carson JK, Campbell L, Rooney D, Clipson N, Gleeson DB (2009) Minerals in soil select distinct bacterial communities in their microhabitats. *FEMS Microbiol Ecol* 67:381–388. <https://doi.org/10.1111/j.1574-6941.2008.00645>
- Carson JK, Gonzalez-Quinones V, Murphy DV, Hinz C, Shaw JA, Gleeson DB (2010) Low pore connectivity increases bacterial diversity in soil. *Appl Environ Microbiol* 76(12):3936–3942. <https://doi.org/10.1128/AEM.03085-09>
- Castro-Sowinski S, Herschkovitz Y, Okon Y, Jurkevitch E (2007) Effects of inoculation with plant growth-promoting rhizobacteria on resident rhizosphere microorganisms. *FEMS Microbiol Lett* 276:1–11. <https://doi.org/10.1111/j.1574-6968.2007.00878.x>
- Certini G, Campbell CD, Edwards AC (2004) Rock fragments in soil support a different microbial community from the fine earth. *Soil Biol Biochem* 36:1119–1128. <https://doi.org/10.1016/j.soilbio.2004.02.022>
- Cho DH, Chae HJ, Kim EY (2001) Synthesis and characterization of a novel extracellular polysaccharide by *Rhodotorula glutinis*. *Appl Biochem Biotech* 95:183–193
- Di H, Cameron KC, Shen JP, Winefield CS, O’Callaghan M, Bowatte S et al (2010) Ammonia-oxidizing bacteria and archaea grow under contrasting soil nitrogen conditions. *FEMS Microbiol Ecol* 72:386–394. <https://doi.org/10.1111/j.1574-6941.2010.00861.x>
- El-Ayouty YMF, Ghazal M, El-Etr WT, EL-Abdeen HAZ (2012) Effect of Cyanobacteria Inoculation associated with different nitrogen levels on some Sandy and Calcareous soils properties and wheat productivity. *Nat Sci* 10(12):233–240

- Ellouze W, Taheri AE, Bainard LD, Yang C, Bazghaleh N, Navarro-Borrell A, Hanson K, Hamel C (2014) Soil fungal resources in annual cropping systems and their potential for management. *BioMed Res Int*. <https://doi.org/10.1155/2014/531824>
- Eman AAA, Saleh MMS, Mostaza EAM (2008) Minimizing the quantity of mineral nitrogen fertilizers on grapevine by using humic acid, organic and bio-fertilizers. *Res J Agric Biol Sci* 4:46–50
- Fierer N, Bradford MA, Jackson RB (2007) Toward an ecological classification of soil bacteria. *Ecology* 88(6):1354–1364. <https://doi.org/10.1890/05-1839>
- Franco-Correa M and Chavarro-Anzola V (2014) Chapter 10: Actinobacteria as plant growth-promoting. Actinobacteria – basics and biotechnological applications. <https://doi.org/10.5772/61291>
- Garrett SD (1951) Ecological groups of soil fungi: a survey of substrate relationships. *New Phytol* 50:149–166
- Gaur AC (1990) Phosphate solubilizing microorganisms as biofertilizers. Omega Scientific Publication, New Delhi
- Giordano W, Hirsch AM (2004) The expression of MaEXP1, a *Melilotus alba expansin* gene, is upregulated during the sweet clover-*Sinorhizobium meliloti* interaction. *Mol Plant Microbe Interact* 17(6):613–622. <https://doi.org/10.1094/MPMI.2004.17.6.613>
- Glick BR, Patten CL, Holguin G, Penrose GM (1999) Biochemical and genetic mechanisms used by plant growth promoting bacteria. Imperial College Press, London
- Grundmann GL, Normand P (2000) Microscale diversity of the genus *Nitrobacter* in soil on the basis of analysis of genes encoding rRNA. *Appl Environ Microb* 66(10):4543–4546
- Harris RF, Chesters G, Allen ON (1966) Dynamics of soil aggregation. *Advances in agronomy* 18:107–169
- Hattori T, Hattori R (1976) The physical environment in soil microbiology: an attempt to extend principles of microbiology to soil microorganisms. *Crit Rev Microbiol* 4:423–460. <https://doi.org/10.3109/10408417609102305>
- Insam H (2001) Developments in soil microbiology since the mid 1960s. *Geoderma* 100:389–402. <https://doi.org/10.1007/s00374-010-0442-3>
- Jobbágy EG, Jackson RB (2000) The vertical distribution of soil organic carbon and its relation to climate and vegetation. *Ecol Appl* 10(2):423–436. [https://doi.org/10.1890/1051-0761\(2000\)010\[0423:TVDOSO\]2.0.CO;2](https://doi.org/10.1890/1051-0761(2000)010[0423:TVDOSO]2.0.CO;2)
- Klironomos JN, McCune J, Hart M, Neville J (2000) The influence of arbuscular mycorrhizae on the relationship between plant diversity and productivity. *Ecol Lett* 3:137–141. <https://doi.org/10.1046/j.1461-0248.2000.00131.x>
- Koide RT (1991) Nutrient supply, nutrient demand and plant response to mycorrhizal infection. *New Phytol* 117:365–386. <https://doi.org/10.1111/j.1469-8137.1991.tb00001.x>
- Kuramae EE, Yergeau E, Wong LC, Pijl AS, van Veen JA, Kowalchuk GA (2012) Soil characteristics more strongly influence soil bacterial communities than land-use type. *FEMS Microbiol Ecol* 79:12–24
- Lipson DA (2007) Relationship between temperature responses and bacterial community structure along seasonal and altitudinal gradients. *FEMS Microb Ecol* 59:418–427. <https://doi.org/10.1111/j.1574-6941.2006.00240.x>
- Lovelock CH, Patterson PG, Walker RH (2004) Services marketing: an Asia pacific perspective, 3rd edn. Prentice Hall, Sydney
- Mauchline TH, Kerry BR, Hirsch PR (2002) Quantification in soil and the rhizosphere of the nematophagous fungus *Verticillium chlamydosporium* by competitive PCR and comparison with selective plating. *Appl Environ Microbiol* 68:1846–1853. <https://doi.org/10.1128/AEM.68.4.1846-1853.2002>
- Mitchell CE (2003) Trophic control of grassland production and biomass by pathogens. *Ecol Lett* 6:147–155. <https://doi.org/10.1046/j.1461-0248.2003.00408.x>